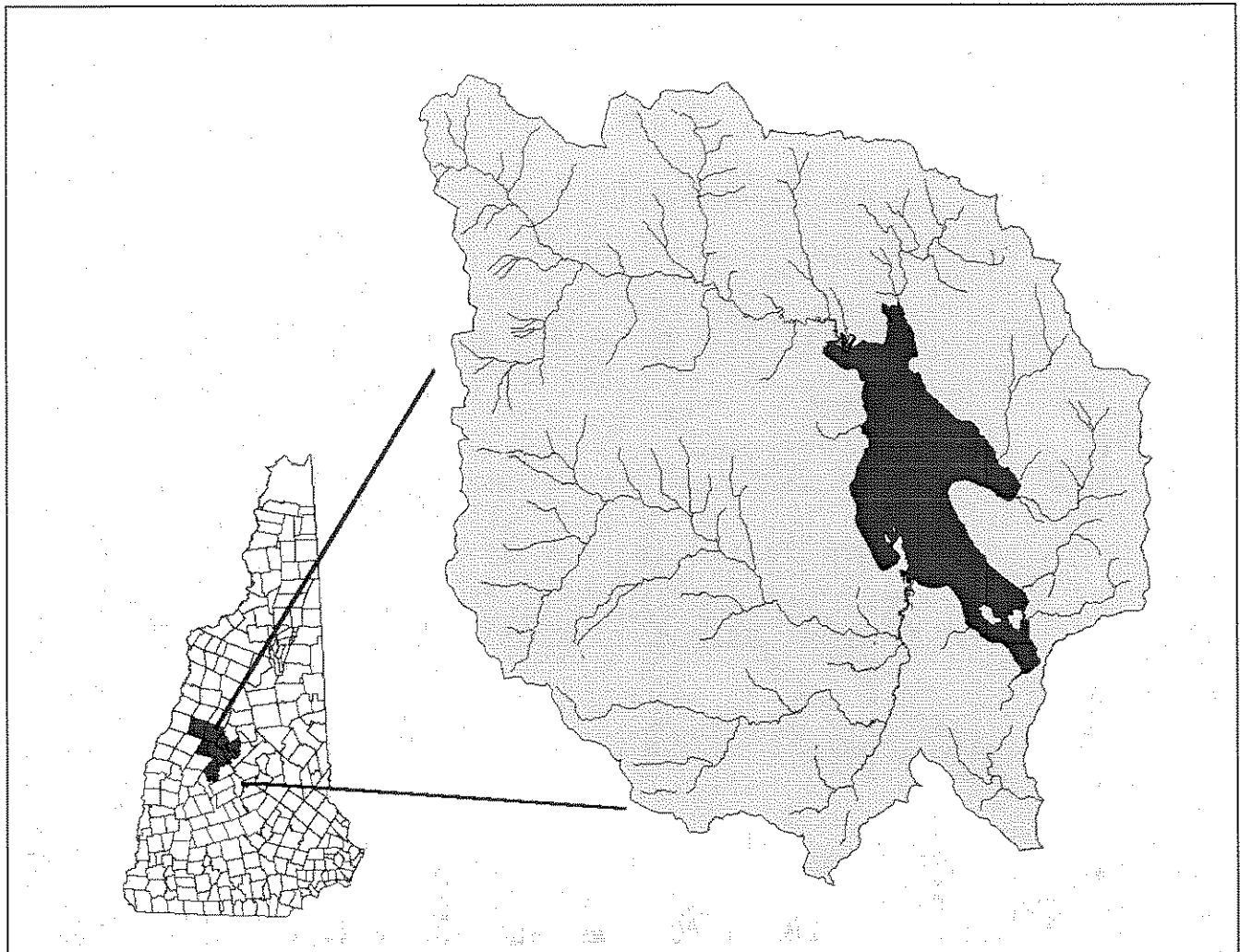


NEWFOUND LAKE

Water Quality Monitoring: 2005

Summary and Recommendations

NH LAKES LAY MONITORING PROGRAM



By: Robert Craycraft & Jeffrey Schloss

Center for Freshwater Biology
University of New Hampshire

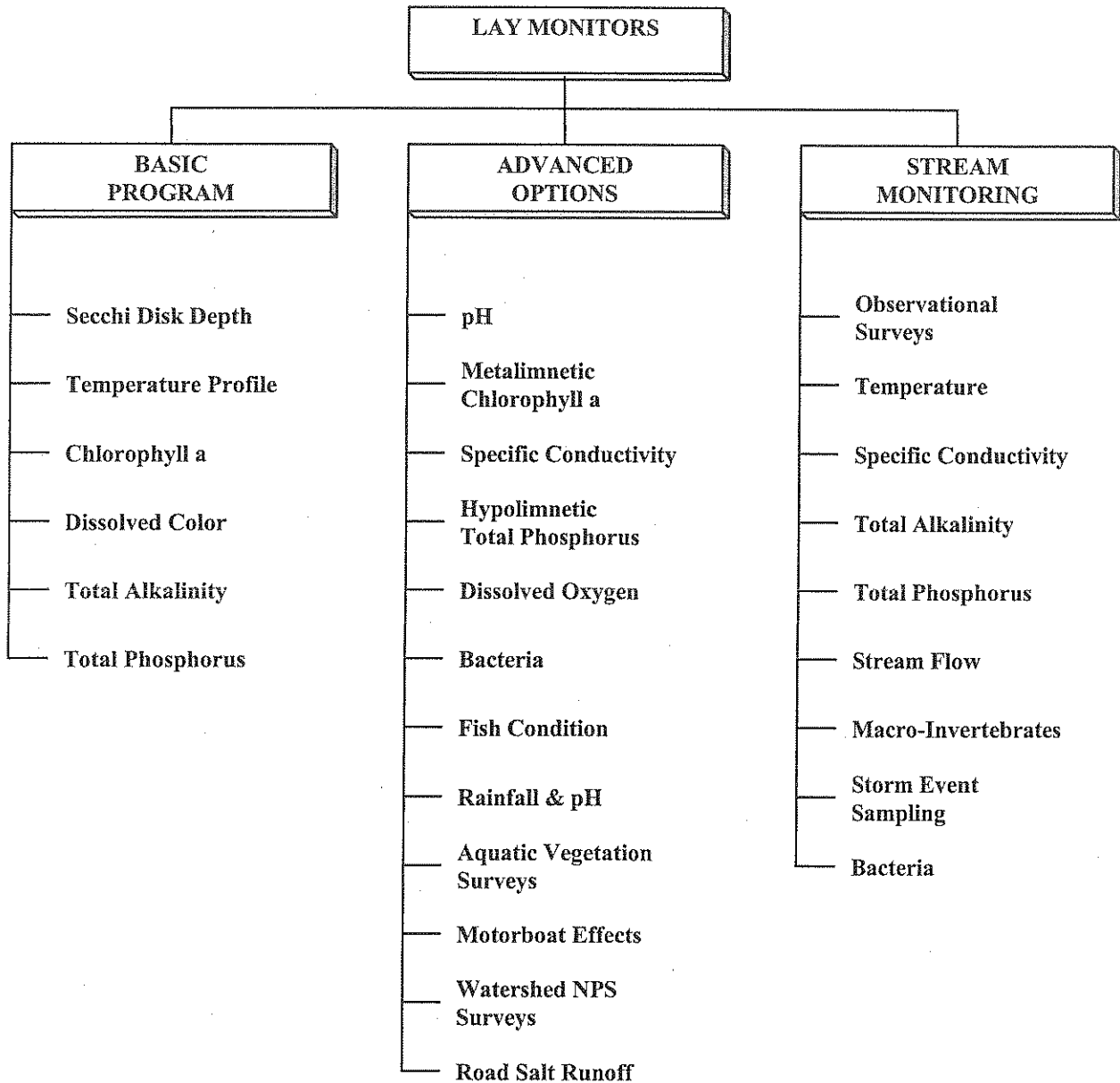


UNIVERSITY of NEW HAMPSHIRE
COOPERATIVE EXTENSION

To obtain additional information on the NH Lakes Lay Monitoring Program (NH LLMP) contact the Coordinator (Jeff Schloss) at 603-862-3848 or Assistant Coordinator (Bob Craycraft) at 603-862-3696.

PARAMETERS SAMPLED

NH LAKES LAY MONITORING PROGRAM



Center for Freshwater Biology (CFB) corroboration with the lay monitor data includes assessment of 1) physical parameters (water transparency, temperature profiles, light transmission profiles and water color); 2) chemical parameters (dissolved oxygen profiles, "free" carbon dioxide, total alkalinity, pH, total phosphorus and specific conductivity profiles); 3) biological parameters (chlorophyll a, phytoplankton community and zooplankton community). Note: in addition to the above parameters, other measurements are often collected at the discretion of the CFB or at the request of the lake association.

PREFACE

This report contains the findings of a water quality survey of Newfound Lake, Towns of Bristol, Hebron, Alexandria and Bridgewater New Hampshire, conducted in the summer of 2005 by the University of New Hampshire **Center for Freshwater Biology (CFB)** in conjunction with the Newfound Lake Region Association.

The report is written with the concerned lake resident in mind and contains a brief, non-technical summary of the year 2005 results as well as more detailed "Introduction" and "Discussion" sections. Graphic display of data is included, in addition to listings of data in appendices, to aid visual perspective.

ACKNOWLEDGMENTS

The year 2005 was the twentieth year that Newfound Lake was monitored in conjunction with the **New Hampshire Lakes Lay Monitoring Program (LLMP)**. The volunteer monitors involved in the water quality monitoring effort are highlighted in Table 1 while Lyn Egsgard and Bill Fay coordinated the volunteer monitoring activities on Newfound Lake and acted as liaisons to the **Center for Freshwater Biology (CFB)**. The CFB congratulates the volunteer monitors on the quality of their work, and the time and effort put forth. We invite other interested residents to join the Newfound Lake water quality monitoring effort in 2006 and expand upon the growing database. Funding for the water quality monitoring program was provided by the Newfound Lake Region Association.

The **Center for Freshwater Biology** is a not-for-profit research program coordinated by Jeffrey Schloss and Robert Craycraft. Members of the CFB summer field team included Ashlee Cieslak, Laura Morcom and Michelle Williams while Thais Fournier, Adam Karr, Kellie Norris, Cassandra Payne and Julie Shelly provided additional assistance in the fall compiling and organizing the water quality data.

The CFB acknowledges the University of New Hampshire Cooperative Extension for funding and furnishing office and storage space while the College of Life Sciences and Agriculture provided laboratory facilities and additional storage space. The CFB would like to thank the **Caswell Family Foundation** for their generosity in providing long-term support for undergraduate assistantships. A gift from the **Samuel P. Pardoe Foundation** allowed for an update of our volunteer temperature profiling equipment, as well as, financial support to develop a data server for our LLMP web site while the **United States Geological Survey**, through the **University of New Hampshire Water Resources Research Center**, provided some resources for staff support. Additional sup-

Table 1: Newfound Lake Volunteer Monitors (2005)

Monitor Name
Joe Allison
Stephanie Bednay
James Benson
Richard & Linda Beyer
Charlie Braman
Greg Carlson
Nick Chichetto
John & Nancy Dineen
Chris & Gavin Divelluss
Caleb Donahue
Olivia Donahue
Ralph Donahue
Kyle Donovan
Gavin Dorcas
John & Lyn Egsgard
Bill & Jeanette Fay
Priscilla Gemmill
Christian Griffin
Alec Hamilton
Matthew Hill
Jeffrey Hillier
George Humphrey
Ben Jenkins
Ken Lonske
Steve McLoy
Alexander Olson
Nat Proctor
Andrew Riely
Gail Ward

port for administering the NH LLMP comes from the **United States Department of Agriculture Cooperative State Research, Education and Extension Service** through support from the New England Regional Water Quality Program (<http://www.usawaterquality.org/newengland/>).

Participating groups in the LLMP include: The Center Harbor Bay Conservation Commission, Dublin Garden Club, Eaton Conservation Commission, Governor's Island Club Inc., Green Mountain Conservation Group, Meredith Bay Rotary Club, The New Hampshire Audubon Society, North River Lake Water Quality Audit Committee, Walker's Pond Conservation Society, the associations of Baboosic Lake, Bow Lake Camp Owners, Chocorua Lake, Crystal Lake, Dublin Lake, Goose Pond, Great East Lake, Lake Kanasatka Watershed, Langdon Cove, Long Island Landowners, Lovell Lake, Mendum's Pond, Merrymeeting Lake, Milton Ponds Lake Lay Monitoring, Mirror Lake (Tuftonboro), Moultonborough Bay, Lake Winnepesaukee, Naticook Lake, Newfound Lake, Nippo Lake, Silver Lake (Madison), Squam Lakes, Sunset Lake, Swains Lake, Lake Wentworth, Winnisquam Drive, and the towns of Alton, Amherst, Enfield, Madison, Meredith, Merrimack, Milton, Strafford, Whitefield and Wolfeboro.

Newfound Lake

2005 Non-Technical Summary

Water quality data were collected by the Newfound Lake volunteer monitors between April 8 and October 5, 2005, while a more in depth water quality survey of the Newfound Lake sampling stations was conducted by the **Center for Freshwater Biology (CFB)** on August 2, 2005 to augment the volunteer monitoring data. Generally speaking, the 2005 Newfound Lake water quality remained excellent as characterized by the high water clarity, the low levels of microscopic plant “algal” growth and low phosphorus (nutrient) levels (Table 2).

The following section discusses the 2005 and historical Newfound Lake water quality data while a complete listing of the 2005 Newfound Lake water quality data is included in Appendix A. The following section includes a discussion of variations among the sampling stations as well as a discussion of trends that have been observed. *Refer to Appendix B for a description of the Box and Whisker plots that are included in the Annual Report for only the second time.*

Table 2: 2005 Newfound Lake Seasonal Average Water Quality Readings and Water Quality Classification Criteria used by the New Hampshire Lakes Lay Monitoring Program.

Parameter	Oligotrophic “Pristine”	Mesotrophic “Transitional”	Eutrophic “Enriched”	Newfound Lake Average (range)	Newfound Classification
Water Clarity (meters)	> 4.0	2.5 - 4.0	< 2.5	7.1 meters (range: 3.4 – 9.3)	Oligotrophic
Chlorophyll a (ppb)	< 3.0	3.0 - 7.0	> 7.0	1.6 ppb (range: 0.6 – 5.4)	Oligotrophic
Phosphorus (ppb) *	< 15.0	15.0 - 25.0	> 25.0	6.9 ppb (range: 5.3 – 11.8)	Oligotrophic

* Data collected by the Center for Freshwater Biology in the surface waters (epilimnion) on August 2, 2005.

1) **Water Clarity (measured as Secchi Disk transparency)** – The Newfound Lake Secchi Disk transparency values were again high during the 2005 sampling season and generally remained visible deeper than 4.0 meters (13.2 feet) that is considered the boundary between a clear “pristine” and more nutrient enriched “transitional” New Hampshire Lake (Figures 10-21). The shallowest 2005 Secchi Disk transparency measurements were documented during the months of June during and following periods of above average rainfall.

An inter-comparison of the sampling sites indicates Site 2 Mayhew continued to exhibit the shallowest water transparency (Figure 30) of the in-lake sampling stations while the clearest median water transparency was documented at Site 3 Pasquaney (Figure 30). Each of the six sampling stations where Secchi Disk transparency measurements were recorded are characterized by a wide range of water transparency measurements that reflect the seasonal variability in water transparency readings (Figure 30).

The annual Secchi Disk comparisons for Sites 2 Mayhew, 3 Pasquaney and 4 Loon Reef indicate a trend of increasing water transparency (i.e. clearer water) between the late 1990s and 2004 that was preceded by a general trend of decreasing water transparency values that was documented between the mid

and the late 1990s (Figures 22, 24 & 26). The median annual water transparency data documented at Site 5 Beachwood also exhibits a general trend of increasing water transparency between 2000 and 2004 (Figure 28).

The 2005 median Newfound Lake Secchi Disk transparency measurements decreased at 2 Mayhew, 3 Pasquaney, 4 Loon Island and 5 Beachwood (Figures 22, 24, 26 & 28) and likely reflect the above average rainfall during the months of April, May and June that is oftentimes associated with elevated concentrations of sediments and nutrients that enter the lake. Annual water quality variations such as those experience in 2005 are common to our New Hampshire Lakes, are oftentimes associated natural climatic fluctuations, and are not necessarily a cause for alarm. On the other hand, such water quality fluctuations serve as a reminder that our lakes are susceptible to increased erosion and nutrient loading during periods of heavy/above average rainfall.

2) Microscopic plant abundance “greenness” (measured as chlorophyll α) – The 2005 seasonal chlorophyll α concentrations generally remained below the level of 3 parts per billion (ppb) that is considered the boundary between an unproductive “pristine” and more nutrient enriched “transitional” lake. Short-term chlorophyll α “spikes” were documented at Sites 6 Fowler on June 26 and July 26, 2005 during which the chlorophyll α concentrations exceeded 3.0 ppb.

A comparison among the in-lake sampling locations indicates the median chlorophyll α concentrations were highest (i.e. greenest water) at the 2 Mayhew and 3 Pasquaney sampling locations and lowest (i.e. least amount of greenness) at the 8 Follansbee Cove sampling location (Figure 31).

The annual chlorophyll α comparisons for Sites 2 Mayhew, 4 Loon Island and 5 Beachwood indicate the chlorophyll α concentrations decreased slightly relative to the 2004 concentrations (Figures 23, 27 & 29). Interestingly, the chlorophyll α concentrations have gradually increased at Site 2 Mayhew between 2001 and 2004 while the water clarity measurements increased (i.e. clearer water) over the four year span (Figures 22 & 23) and suggest the composition of the algal community might be changing. Such changes can occur naturally but can also be associated with changes in land-use and a corresponding shift in the amount of nutrients that are reaching the lake.

The 2005 Newfound Lake, Site 3 Pasquaney, median chlorophyll α concentration increased to the highest (i.e. greenest water) level documented since the year 2000 (Figure 24). The elevated 3 Pasquaney chlorophyll α concentrations corresponded to some of the shallower water transparency measurements documented at the site in recent years (Figures 24 & 25).

Continued in-lake monitoring during the 2006 season, as well as a complementary 18 month stream study that is scheduled to begin in June 2006, will better assess the nutrient load that is entering Newfound Lake through the tributary inlets and will provide additional insight into Newfound Lake’s response to the nutrient loading.

3) Background (dissolved) water color: often perceived as a “tea” color in more highly stained lakes –

The 2005 Newfound Lake dissolved color concentration of 15.8 chloroplatinate units (cpu) is characteristic of a slightly “tea” colored lake (Table 3). Dissolved color, or true color as it is sometimes called, is indicative of dissolved organic carbon levels in the water (a by-product of microbial decomposition). Small increases in water color from the natural breakdown of plant materials in and around a lake are not considered to be detrimental to water quality. However, increased color can diminish water transparency, and hence, change the public perception of water quality.

Table 3. Dissolved Color Classification Criteria used by the New Hampshire Lakes Lay Monitoring Program.

Range	Classification
0 - 10	Clear
10 - 20	Slightly colored
20 - 40	light tea color
40 - 80	tea colored
> 80	highly tea colored

4) Total Phosphorus: the nutrient considered most responsible for elevated microscopic plant growth in our New Hampshire Lakes. – Total phosphorus samples were collected by the University of New Hampshire Center for Freshwater Biology on August 2, 2005 and ranged from 4.5 to 11.8 parts per billion (ppb) at that time. The August 2, 2005 total phosphorus readings remained well below the concentration of 15 ppb that is considered the boundary between an unproductive and a more nutrient enriched “transitional” New Hampshire Lake. Supplemental tributary samples were collected by the Newfound Lake volunteer monitors between April 8 - April 22 and ranged from 3.2 and 15.1 ppb. While variable, the tributary total phosphorus samples were not alarmingly high and did not suggest any gross water quality problems at the time of sampling. *Refer to Appendix A for a complete listing of the tributary data.*

5) Resistance against acid precipitation (measured as total alkalinity) –

The 2005 Newfound Lake alkalinity measured 2.8 milligrams per liter (mg/l) and is considered typical of a lake with a moderate vulnerability to acid precipitation according to the standards devised by the New Hampshire Department of Environmental Services (Table 4). Generally speaking, the geology of the region does not contain the mineral content (e.g. limestone) which increases the buffering capacity in our surface waters. Thus, lakes in the vicinity (e.g. Squam Lake, Lake Winnepesaukee) have naturally low alkalinities. While low, the 2005 Newfound Lake alkalinity remained sufficient to neutralize acid inputs during the summer months.

Table 4. Alkalinity Classification Criteria used by the New Hampshire Department of Environmental Services

Range	(i) Classification
< 0	Acidified
0 - 2	Extremely Vulnerable
2.1 - 10.0	Moderately Vulnerable
10.1 - 25.0	Low Vulnerability
> 25.0	Not Vulnerable

Lake acidity (measured as pH) - The August 2, 2005 Newfound Lake pH data, collected in the surface waters by the **Center for Freshwater Biology**, ranged from 6.7 to 6.9 and remained well within the tolerable range for most aquatic organisms.

6) **Dissolved salts: measured as specific conductivity** – Specific Conductivity levels documented in Newfound Lake, by the **Center for Freshwater Biology**, were low and ranged from 38.1 to 44.3 micro-Siemans (*uS*) when measured on August 2, 2005. High specific conductivity values can be an indication of problem areas around a lake where failing septic systems, heavy fertilizer applications and sedimentation are contributing “excessive” nutrients into the lake. High specific conductivity readings can also be an indication of excessive road salt applications within the Newfound Lake watershed.

7) **Temperature and dissolved oxygen profiles** – Temperature profiles were collected by the volunteer monitors and indicate Newfound Lake becomes stratified into three distinct thermal layers during the summer months; a warm upper water layer, the **epilimnion**, overlies a layer of deep cold-water known as the **hypolimnion** while a layer of rapidly decreasing temperatures, the **metalimnion**, serves as a transition zone between the upper warmwater and deep coldwater layers. The formation of thermal stratification limits the replenishment of oxygen in the deeper waters and under sub-optimal conditions can coincide with oxygen depletion near the lake-bottom.

Dissolved oxygen concentrations required for a healthy fishery – Dissolved oxygen concentrations documented by the **Center for Freshwater Biology** remained above the concentration of 5 milligrams per liter down to the lake bottom at each of the deep Newfound Lake sampling stations on August 2, 2005 (Table 5 and Figures 32-34). The dissolved oxygen concentration of 5 milligrams per liter is considered the minimum oxygen concentration required for the successful growth and reproduction of most coldwater fish including trout and salmon. Similar to historical years of sampling, the dissolved oxygen concentrations measured at Site 2 Mayhew were lower near the lakebottom, relative to the dissolved oxygen concentrations measured at the other sampling stations. However, unlike past years of sampling, the Site 2 Mayhew dissolved oxygen concentrations remained above 5 milligrams per liter down to the lakebottom (Figure 32).

Table 5. August 2, 2005 Newfound Lake Dissolved Oxygen (DO) Concentrations and corresponding water quality classification criteria.

Sampling Station	DO Range (ppm) *	Classification
2 Mayhew	5.7 – 7.4 ppm	“transitional”
3 Pasquaney	11.6 – 11.7 ppm	“pristine”
5 Beachwood	11.7 – 11.8 ppm	“pristine”
7 Cockermouth	11.4 – 11.9 ppm	“pristine”
8 Follansbee	11.5 – 12.1 ppm	“pristine”

* Classification based on Dissolved oxygen Concentrations in the bottom waters (hypolimnion). Dissolved oxygen concentrations > 5 ppm are often considered typical of a “pristine” lake while dissolved oxygen concentrations < 2.0 ppm are considered typical of an “enriched” lake. Dissolved oxygen concentrations between 2.0 and 5.0 ppm are considered typical of a moderately productive “transitional” lake.

Generally speaking, the year 2005 dissolved oxygen concentrations remained within the optimal range for the Newfound Lake fishery with the single exception of the declining dissolved oxygen concentrations near the lake bottom of Site 2 Mayhew that were nearing the concentrations considered stressful to coldwater fish.

8) Comparisons between the **Center for Freshwater Biology** and lay monitor data indicate the volunteer monitors are doing an excellent job of collecting water quality data.

Based on the current and historical water quality data, Newfound Lake would be considered an unproductive “pristine” New Hampshire lake. However, continued development around the lake, heavy fertilizer applications and aging septic systems pose a threat to the high water quality characteristic of Newfound Lake, particularly when local landowners do not take the appropriate precautions to minimize the transport of pollutants (i.e. sediments and the nutrient phosphorus) into the lake. Thus, a first step towards preserving high water quality in Newfound Lake is to take action at the local level and do your part to minimize the number of pollutants that enter Newfound Lake. Whenever possible, **maintain riparian buffers** (vegetative buffers adjacent to the water body). These buffers will biologically “take up” nutrients before they enter the lake and will also provide physical filters which allow materials to settle out before reaching the lake. Preserving buffer strips and avoiding shorefront disturbance also reduces the potential for aquatic plant colonization, including variable milfoil and other invasive weeds. No area is immune to the infestation of invasive weeds like milfoil, but plants are less likely to root in undisturbed shoreline areas. Riparian buffers should be considered for both Newfound Lake as well as the stream inlets that are responsible for the majority of the overland runoff that enters Newfound Lake. **Reduce fertilizer applications.** Most residents apply far more fertilizers than necessary which can be a costly expense to the homeowner and can also be detrimental to the lake since the same nutrients that make our lawns green will also stimulate plant growth in our lakes. **Make sure your septic system is well maintained** and have it pumped out on a regular basis. An improperly functioning septic system can contribute “excessive” nutrients into the lake and result in early failure, costing thousands of dollars to repair or replace. Future volunteer monitoring efforts should be directed at pinpointing problematic regions around the lake where corrective and educational efforts should be focused.

COMMENTS AND RECOMMENDATIONS

- 1) We recommend that each participating association, including the Newfound Lake Region Association, continue to develop its database on lake water quality through continuation of the long-term monitoring program. The database currently provides information on the short-term and long-term cyclic variability that occurs in the lake and through continued monitoring would enable more reliable predictions of both short-term and long-term water quality trends.

- 2) We recommend initiating lake sampling early in the season (April/May) to document Newfound Lake's reaction to the nutrient and acid loadings that typically occur during and after spring thaw. Sampling should include alkalinity, chlorophyll α , dissolved color and Secchi Disk transparency measurements. Phosphorus samples are also recommended from both the in-lake and the tributary sampling sites. When tributary samples are collected, streamflow measurements should be included whenever possible.

- 3) Frequent "weekly" water quality samples, necessary to assess the current condition of Newfound Lake, should continue to be collected whenever possible. Weekly sampling of chlorophyll α , Secchi Disk transparency, dissolved color and alkalinity is recommended while the addition of monthly total phosphorus samples would also be useful to track variations in nutrient loading during the summer months.

- 4) The Newfound Lake Region Association might will be expanding the 2006 volunteer water quality monitoring effort to include an extensive tributary sampling program that will better assess whether or not localized water quality problems exist within the Newfound Lake watershed. The tributary sampling program will involve the collection of weekly water quality measurements at 18 tributaries to identify the sources of nutrients that enter the lake. Supplemental stream sampling will also be undertaken to "bracket" different stream reaches to better assess whether there are identifiable problems at certain points along selected tributary segments. Should potential "hot spots" be identified, future educational efforts could be focused on those priority regions. Should you be interested in volunteering you time to this effort please contact Boyd Smith, Newfound Lake Region Association executive director, or contact Bob Craycraft of the University of New Hampshire at 862-3696 or via email at bob.craycraft@unh.edu.

TABLE OF CONTENTS

PREFACE.....	I
ACKNOWLEDGMENTS	II
NEWFOUND LAKE - 2005 NON-TECHNICAL SUMMARY	IV
COMMENTS AND RECOMMENDATIONS	IX
TABLE OF CONTENTS	X
REPORT FIGURES	XII
INTRODUCTION.....	1
The New Hampshire Lakes Lay Monitoring Program.....	1
Importance of Long-term Monitoring	3
Purpose and Scope of This Effort	4
CLIMATIC SUMMARY - 2005	6
Water Quality and the Weather	6
Precipitation (2005).....	7
Water Quality Impacts	9
Water Transparency and Dissolved “tea” Colored Water	9
Sediment Loading.....	10
Nutrient Loading	10
Microscopic “Algal” and Macroscopic “Weed” Plant Growth.....	11
DISCUSSION OF LAKE AND STREAM MONITORING MEASUREMENTS.....	12
Thermal Stratification in the Deep Water Sites	12
Water Transparency.....	12
Chlorophyll <i>a</i>	13
Turbidity *	13
Dissolved Color.....	13
Total Phosphorus.....	14
Streamflow	14
pH *.....	14
Alkalinity.....	14
Specific Conductivity *	15
Dissolved Oxygen and Free Carbon Dioxide *	15
Underwater Light *	16
Indicator Bacteria *	16
Phytoplankton *	17
Zooplankton *	17
Macroinvertebrates *	18
Fish Condition	18
UNDERSTANDING LAKE AGING (EUTROPHICATION)	20
How can you minimize your water quality impacts?.....	22
REFERENCES.....	24

REPORT FIGURES29
APPENDIX A: 2005 NEWFOUND LAKE DATA SUMMARY A-1
APPENDIX B: DETERMINING WATER QUALITY CHANGES & TRENDSB-1
APPENDIX C: GLOSSARY OF LIMNOLOGICAL TERMS C-1

REPORT FIGURES

Figure 1. LLMP Objectives	1
Figure 2. National LLMP Support Volunteer Monitoring Programs	2
Figure 3. Algal Standing Crop: 1988-1992.....	3
Figure 4. Algal Standing Crop: 1986-1995.....	4
Figure 5. Monthly Precipitation (1980-2004).....	7
Figure 6. Monthly Snowfall (1982-2004).....	8
Figure 7. Monthly Temperature (1984-2004).....	9
Figure 8. Typical Temperature Conditions: Summer	12
Figure 9. Location of the Newfound Lake sampling stations: Sites 1 Deep, 2 Mayhew, 3 Pasquaney, 4 Loon Island, 5 Beachwood, 6 Fowler River, 7 Cockermonth River and 8 Follansbee Cove.	29
Figure 10. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll <i>a</i> trends for Site 2 Mayhew.....	31
Figure 11. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 2 Mayhew.	31
Figure 12. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll <i>a</i> trends for Site 3 Pasquaney Bay.	33
Figure 13. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 3 Pasquaney Bay.....	33
Figure 14. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll <i>a</i> trends for Site 4 Loon Island.....	35
Figure 15. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 4 Loon Island:.....	35
Figure 16. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll <i>a</i> trends for Site 5 Beachwood.	37
Figure 17. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 5 Beachwood.	37
Figure 18. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll <i>a</i> trends for Site 7 Cockermonth River.	39
Figure 19. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 7 Cockermonth River.	39

Figure 20. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll <i>a</i> trends for Site 8 Follansbee Cove.	41
Figure 21. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 8 Follansbee Cove.	41
Figure 22. Comparison of the annual Newfound Lake, Site 2 Mayhew, lay monitor Secchi Disk transparency data (1986-2005) that are presented as box and whisker plots.	43
Figure 23. Comparison of the annual Newfound Lake, Site 2 Mayhew, lay monitor chlorophyll <i>a</i> data (1986-2005) that are presented as box and whisker plots.	43
Figure 24. Comparison of the annual Newfound Lake, Site 3 Pasquaney, lay monitor Secchi Disk transparency data (1986-2005) that are presented as box and whisker plots.	45
Figure 25. Comparison of the annual Newfound Lake, Site 3 Pasquaney, lay monitor chlorophyll <i>a</i> data (1986-2005) that are presented as box and whisker plots.	45
Figure 26. Comparison of the annual Newfound Lake, Site 4 Loon Island, lay monitor Secchi Disk transparency data (1987-2005) that are presented as box and whisker plots.	47
Figure 27. Comparison of the annual Newfound Lake, Site 4 Loon Island, lay monitor chlorophyll <i>a</i> data (1987-2005) that are presented as box and whisker plots.	47
Figure 28. Comparison of the annual Newfound Lake, Site 5 Beachwood, lay monitor Secchi Disk transparency data (1999-2005) that are presented as box and whisker plots.	49
Figure 29. Comparison of the annual Newfound Lake, Site 5 Beachwood, lay monitor chlorophyll <i>a</i> data (1999-2005) that are presented as box and whisker plots.	49
Figure 30. Newfound Lake inter-site comparison of the 2005 lay monitor Secchi Disk transparency data that are presented as box and whisker plots.	51
Figure 31. Newfound Lake inter-site comparison of the 2005 lay monitor Chlorophyll <i>a</i> data that are presented as box and whisker plots.	51
Figure 32. Temperature and dissolved oxygen profiles collected in Newfound Lake, (A) Site 2 Mayhew and (B) Site 3 Pasquaney on August 2, 2005.	53
Figure 33. Temperature and dissolved oxygen profiles collected in Newfound Lake, (A) Site 4 Loon Island and (B) Site 5 Beachwood on August 2, 2005.	55
Figure 34. Temperature and dissolved profiles collected in Newfound Lake, (A) Site 7 Cockermouth River and (B) Site 8 Follansbee Cove on August 2, 2005.	57

INTRODUCTION

The New Hampshire Lakes Lay Monitoring Program

The 2005 sampling season marked the twenty-seventh anniversary for the NH Lakes Lay Monitoring Program (LLMP). The LLMP has grown from a university class project on Chocorua Lake and pilot study on the Squam Lakes to a comprehensive state-wide program with over 500 volunteer monitors and more than 100 lakes participating. Originally developed to establish a database for determining long-term trends of lake water quality for science and management, the program has expanded by taking advantage of the many resources that citizen monitors can provide (Figure 1).

The NH LLMP has gained an international reputation as a successful cooperative monitoring, education and research program. Current projects include: the use of volunteer generated data for non-point pollution studies using high tech analysis system (Geographic Information Systems and Satellite Remote Sensing), and intensive watershed monitoring for the development of watershed nutrient budgets, investigations of water quality and indicator organisms (food web analysis, fish condition, and stream invertebrates). The key ingredients responsible for the success of the program include innovative cost share funding and cost reduction, assurance of credible data, practical sampling protocols and, most importantly, the interest and motivation of our volunteer monitors.

The 2005 sampling season was another exciting year for the New Hampshire Lakes Lay Monitoring Program. National recognition for the high quality of work by you, the volunteer monitors, continued with awards, requests for program information and invitations to speak at national conferences (Table 8).

Figure 1. LLMP Objectives

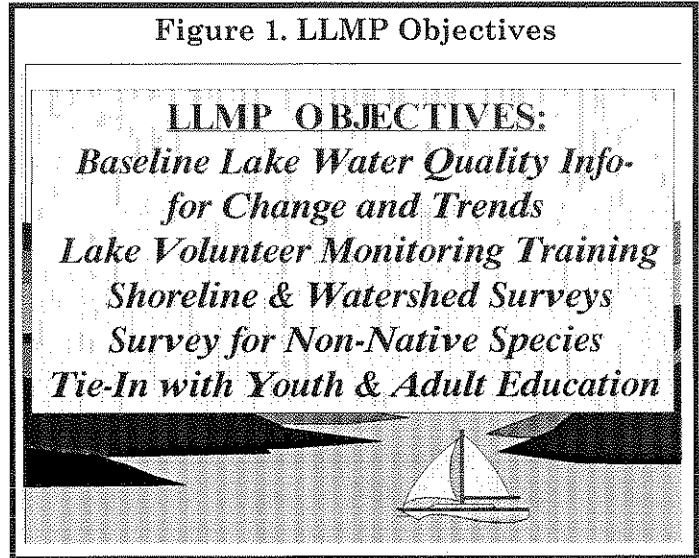


Table 8. Awards & Recognition

- 1983- NH Environmental Law Council Award
- 1984- Governor's Volunteer Award
- 1985- CNN Science & Technology Today
- 1988- Governor's "Gift" award funded
- 1990- NH Journal TV coverage NHPTV
- 1991- Renew America Award
Environmental Success Index
White House Reception / Briefing
- 1992- EPA Administrators Award
Environmental Exchange Network Listing
- 1993- NH Lakes Association Award
- 1994- EPA Office of Watersheds Award
- 1995- Winnepesaukee Watershed Project
- 1998- Governor's Proclamation for 20th Anniversary
- 1999- EPA Watershed Academy Host
- 2001- Lake Chocorua Project highlighted at national conferences (invited presentations)
- 2002- Chocorua Project receives Technical Excellence Award from the North American Lake Management Society
- 2003- UNH CE Maynard and Audrey Heckel Extension Fellowship awarded to LLMP
- 2004- Participatory Research Model of NH LLMP highlighted at National Water Quality Monitoring Conference
- 2005- LLMP Coordinator J. Schloss receives the prestigious Secchi Disk Award from the North American Lakes Management Society

We are excited by the results of teaming up students, educators and local lake residents through our Multidisciplinary Lakes Management course and our summer Community Mapping with GIS and Watershed Ecology courses that are held annually (the two latter mentioned courses are for educators, community leaders and other interested persons). Some of the lake management recommendations made as part of the student coursework requirements have been successfully implemented by lake associations.

Our active collaboration with the UNH Center for Freshwater Biology continues to drive relevant applied research: Our volunteer intensive Water / Nutrient Budget Study of the Squam Lakes Watershed has provided insights into the impacts of development, growth and septic systems on lake water quality. Combined with the knowledge gained from the Lake Chocorua Watershed Study we are closer to understanding what the level of nutrient conditions we can expect from different land uses.

We have also been a key player in testing an integrated pest management approach (see our "Special Topic" in this report) to exotic variable water milfoil control and have also recently initiated a study examining the potential to manage exotic milfoil growth using parasitic nematodes.

We continue the research initiated by collaborators Dr. John Sasner and Dr. Jim Haney focusing on how watershed development and our activities on the landscape play a role in creating potentially toxic algae blooms. Analogous to the 'red tide' of estuaries, certain blue-green algae (microscopic bacteria) can produce toxins that are health risks to animals and humans.

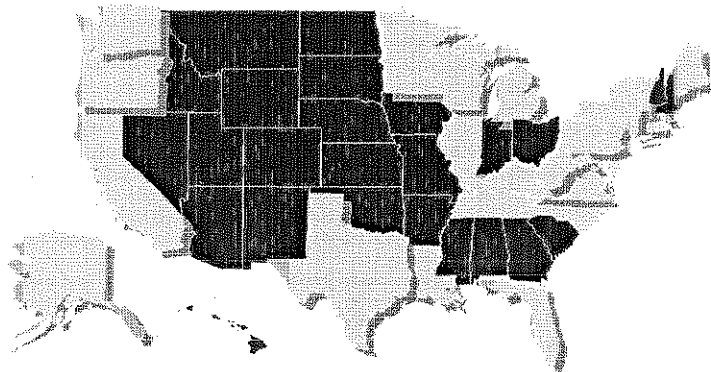
Additional ongoing research is focusing on the use of satellite imagery as well as on-lake optical devices as a means of determining the water transparency and amount of microscopic plant "algal" growth in our New Hampshire Lakes, particularly blue green algae. Water quality data, collected by the volunteer monitors, have served as ground truthed data to assess whether or not the satellite imagery shows promise. Data generated through this project have been presented at national conferences and are testament to the high quality data generated by our volunteer monitors.

Recent interest in the success of our NH LLMP participatory research model has resulted in invited presentations at national conferences and provided the basis of a series of articles in the Volunteer Monitor, the national newsletter with a distribution of over 10,000.

We continue to be listed as a model citizen-monitoring program on the Environmental Success Index of Renew America, the Environmental Network Clearinghouse

Figure 2. National LLMP Support to Volunteer Monitoring Programs

NH LLMP Directly involved with the Initiation, Expansion or Support of Volunteer Programs in 24 States.



Light gray shading denotes LLMP assisted states

and the National Awards Council for Environmental Sustainability. To date, the approach and methods of the NH LLMP have been adopted by new or existing programs in twenty-four states and eleven countries (Figure 2)!

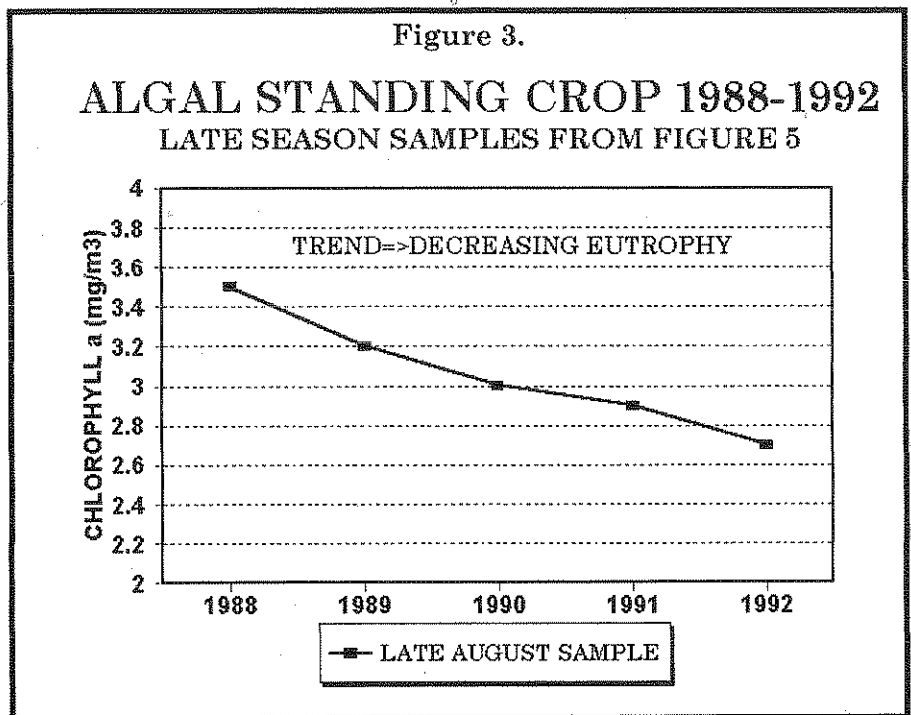
Importance of Long-term Monitoring

A major goal of our monitoring program is to identify any short or long-term changes in the water quality of the lake. Of major concern is the detection of cultural eutrophication: increases in the productivity of the lake, the amount of algae and plant growth, due to the addition of nutrients from human activities. Changes in the natural buffering capacity of the lakes in the program is also a topic of great concern, as New Hampshire receives large amounts of acid precipitation, yet most of our lakes contain little mineral content to neutralize this type of pollution.

For over two decades, weekly data collected from lakes participating in the New Hampshire Lakes Lay Monitoring Program have indicated there is quite a variation in water quality indicators through the

open water season (April through November) on the majority of lakes. Short-term differences may be due to variations in weather, lake use, or other chance events. Monthly sampling of a lake during a single summer provides some useful information, but there is a greater chance that important short-term events such as algal blooms or the lake's response to storm run-off will be missed. These short-term fluctuations may be unrelated to the actual long-term trend of a lake or they may be indicative of the changing status or "health" of a lake.

Consider the hypothetical data depicted in Figure 3. Limiting sampling to only once a year during August, from 1988 to 1992, produced a plot suggesting a decrease in eutrophication. However, the actual long-term trend of the lake, increasing eutrophication, can only be clearly discerned by frequent sampling over a ten year period (Figure 5). In this instance, the information necessary to distinguish between short-term fluctuations "noise" and long-term trends "signal" could only be accomplished through the frequent collection of water quality data over many years. To that end, the establishment of a long term database was essential to trend detection.



The number of seasons it takes to distinguish between the noise and the signal is not the same for each lake. Evaluation and interpretation of a long-term database will indicate that the water quality of the lake has worsened, improved, or remained the same. In addition, different areas of a lake may show a different response. As more data are collected, prediction of current and future trends can be made. No matter what the outcome, this information is essential for the intelligent management of your lake.

There are also short-term uses for lay monitoring data. The examination of different stations in a lake

can disclose the location of specific problems and corrective action can be initiated to handle the situation before it becomes more serious. On a lighter note, some associations post their weekly data for use in determining the best depths for finding fish!

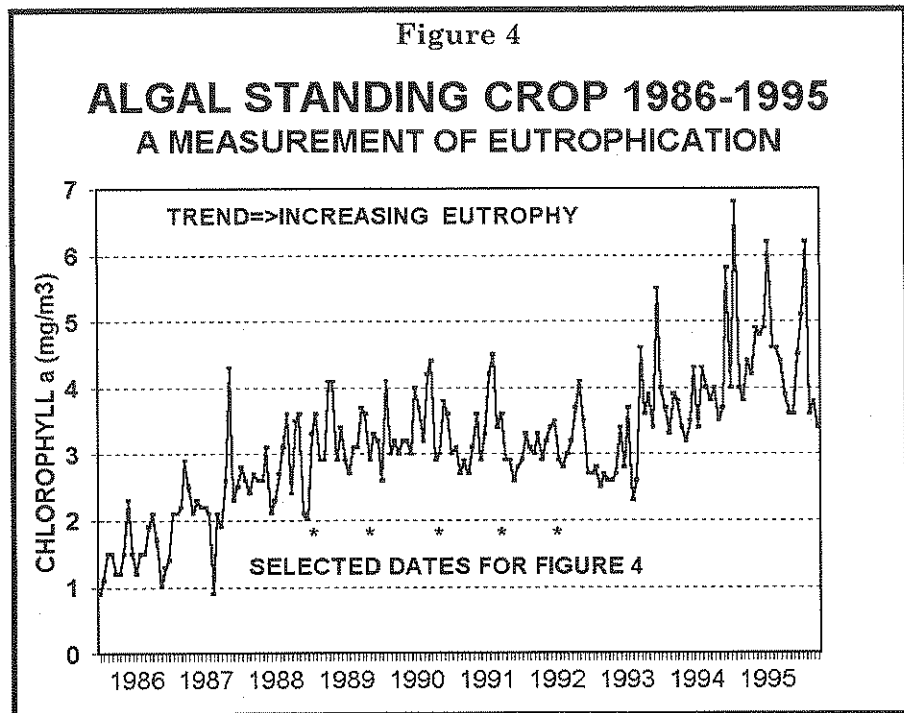
It takes a considerable amount of effort as well as a deep concern for one's lake to be a volunteer in the **NH Lakes Lay Monitoring Program**. Many times a monitor has to brave inclement weather or heavy boat traffic to collect samples. Sometimes it even may seem that one week's data is just the same as the next week's data. Yet every sampling provides important information on the variability of the lake.

We are pleased with the interest and commitment of our Lay Monitors and are proud that their work is what makes the **NH LLMP** the most extensive, and we believe, the best volunteer program of its kind.

Purpose and Scope of This Effort

2005 was the twenty-seventh year that water quality monitoring of the Squam Lakes was undertaken in conjunction with the **NH Lakes Lay Monitoring Program (NH LLMP)**. The monitoring program is designed to establish a long-term database to which future data could be compared and to identify localized problem areas where future educational and corrective efforts should be focused. Sampling emphasis was placed on 13 open water sampling stations around the lake while additional tributary sampling has also been undertaken historically at the major stream inlets to both Squam Lake and Little Squam Lake.

The primary purpose of annual lake reporting is to discuss results of the current monitoring season with emphasis on current conditions of New Hampshire lakes including the extent of eutrophication and the lakes' susceptibility to increasing acid precipitation. If there are additional water quality concerns we advise the lake association to



contact our program staff to discuss additional monitoring options. When applicable we also strive to place the recent results into a historical context using past NH LLMP data as well as historical data from other sources. This information is part of a large data base of historical and more recent data compiled and entered onto our computer files for New Hampshire lakes that include New Hampshire Fish and Game surveys of the 1930's, the surveys conducted by the New Hampshire Water Supply and Pollution Control Commission and the **CFB/FBG** surveys. However, care must be taken when comparing current results with early studies. Many complications arise due to methodological differences of the various analytical facilities and technological improvements in testing.

Climatic Summary - 2005

Water Quality and the Weather

Water quality variations are commonly observed over the course of the year and among years in our New Hampshire lakes, ponds, wetlands and streams. The most commonly noticed changes are those associated with decreasing water clarities, increasing algal growth (greenness), and increasing plant growth around the lake's periphery. Over the long haul, changes such as these are attributed to a lake's natural aging process; what is known as "**eutrophication**". However, short-term water quality changes such as those mentioned above are often encountered even in our most pristine lakes and ponds. These water quality changes often coincide with variations in weather patterns that include precipitation and temperature fluctuations and even variations in the sunlight intensity that can accelerate or suppress the photosynthetic process.

Climatic "swings" can have a profound effect on water quality, sometimes positive and other times negative. For instance, 1996 was a wet year relative to other years of LLMP water quality monitoring. This translated into reduced water clarities, elevated microscopic plant "algal" growth and increased total phosphorus concentrations for most participating LLMP lakes. "Excessive" runoff associated with wet periods often facilitates the transport of pollutants such as nutrients (including phosphorus), sediment, dissolved colored compounds, as well as toxic materials such as herbicides, automotive oils, etc. into water bodies. As a result, lakes often respond with shallower (less clear) water clarities and elevated algal abundance "greenness" during these periods as evidenced by historical monitoring through the NH LLMP. Similarly, short-term storm events can have a profound effect on the water quality. Take for instance the "100 year storm" (October 21-22, 1996) that blanketed southern New Hampshire with approximately 6 inches of rain over a 30-hour period. This storm resulted in increased sedimentation and organic matter loading into our lakes as materials were flushed into the water bodies from the adjacent uplands. Likewise, the heavy rains that saturated the soil and resulted in flood conditions in June 1998 (heaviest rains occurring on June 12 and 13) resulted in significantly shallower water transparency readings in the weeks to months that followed. While events such as the October 1996 and the June 1998 storms are short lived, they can have a profound effect on our water quality in the weeks to months that follow, particularly when nutrients that stimulate plant growth are retained in the lake.

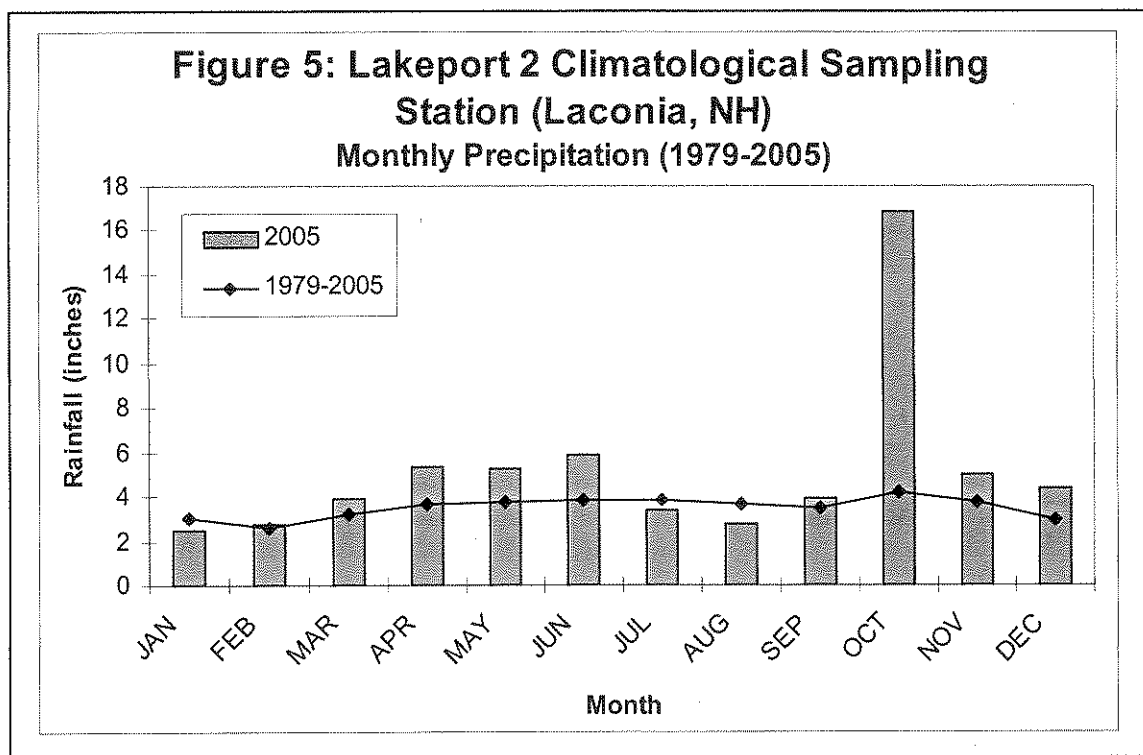
NH LLMP data collected during dry years such as 1985 and 2001, on the other hand, have coincided with improved water quality for many New Hampshire lakes. Reduced transport of pollutants into the lake often results in higher water quality measured as deeper water transparencies, lower microscopic plant "algae" concentrations and lower nutrient concentrations. Do all lakes experience poorer water quality as a result of heavy precipitation events? Simply stated, the answer is no. While most New Hampshire lakes are characterized by reduced water clarities, increased nutrients and elevated plant "algal" concentrations following periods, or years, of heavy precipitation, a handful of lakes actually benefit from these types of events. The water bodies that improve during wet periods are generally lakes characterized by high nutrient concentrations and high "algal" concentrations that are diluted by watershed runoff and thus

benefit during periods, or years, of heavy rainfall. However, these more nutrient enriched lakes remain susceptible to nutrients entering the lake from seepage sources such as poorly functioning septic systems.

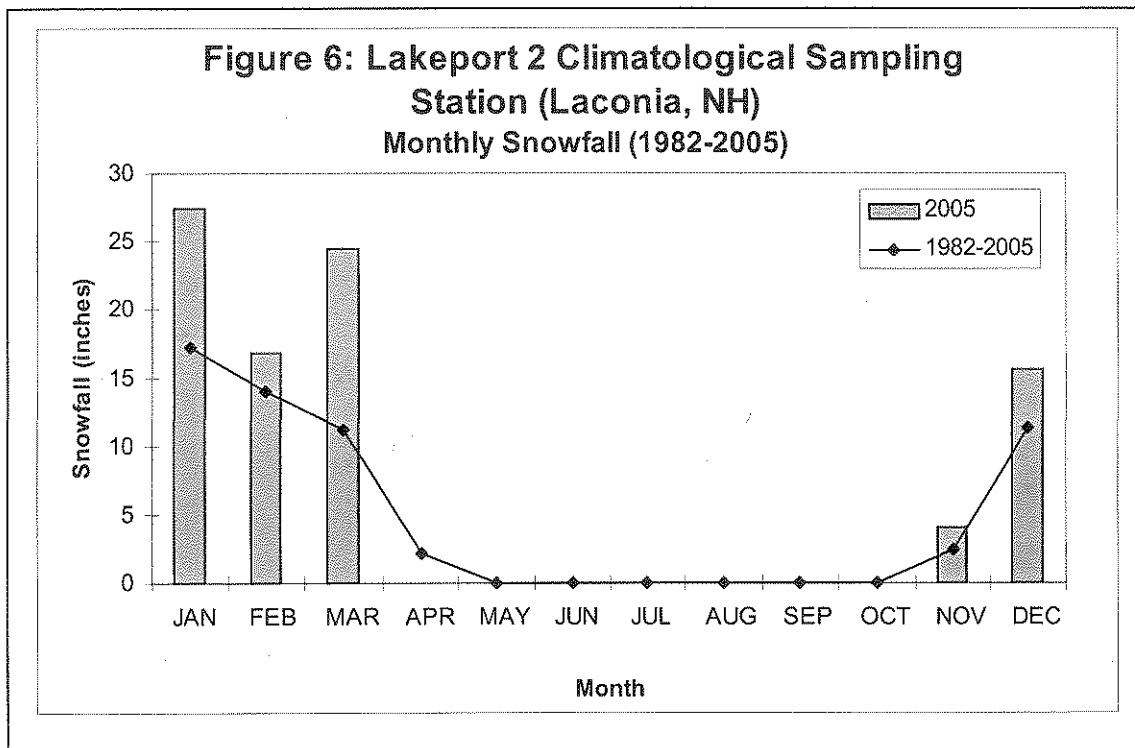
Precipitation (2005)

The 2005 annual precipitation (reported as “rainfall” water equivalent) measured 61.94 inches and was the highest “rainfall” amount that has been documented over the past 27 years: 1979-2005 (note: precipitation data are reported for the Lakeport 2 Climatological sampling station located in Laconia New Hampshire: 43°33'N and 71°28'W). The monthly precipitation totals documented during the months of January and February were near/below average although the timing of the rainfall events coincided with low temperatures that translated into relatively heavy periods of snowfall during January and February. The March precipitation was above average and was coupled with two snow-storms, and below average temperatures, during the first third of the month that culminated in of the additional snowpack accumulation that contributed to periods of heavy runoff later in the month of March and into the Month of May (Figures 5 & 6). Significant accumulations of winter snowpack can result in a period of heavy overland runoff in the spring that oftentimes coincides with increased sediment and nutrient loading that negatively impacts water quality. Above average rainfall continued into the month of June during which the rainfall was largely concentrated to the middle of the month. The subsequent months of July and August were characterized by below average rainfall followed by slightly above average rainfall during the month of September.

A series of storms swept through New Hampshire during the month of October during which the rainfall amount was nearly four times above the 27 year average documented between 1979 to 2005 (Figure 5). Futhermore, the October 2005 rainfall reached record levels in some locations in southern New Hampshire. The months of No-



vember and December rounded out the years with above average rainfall and above average snowfall.

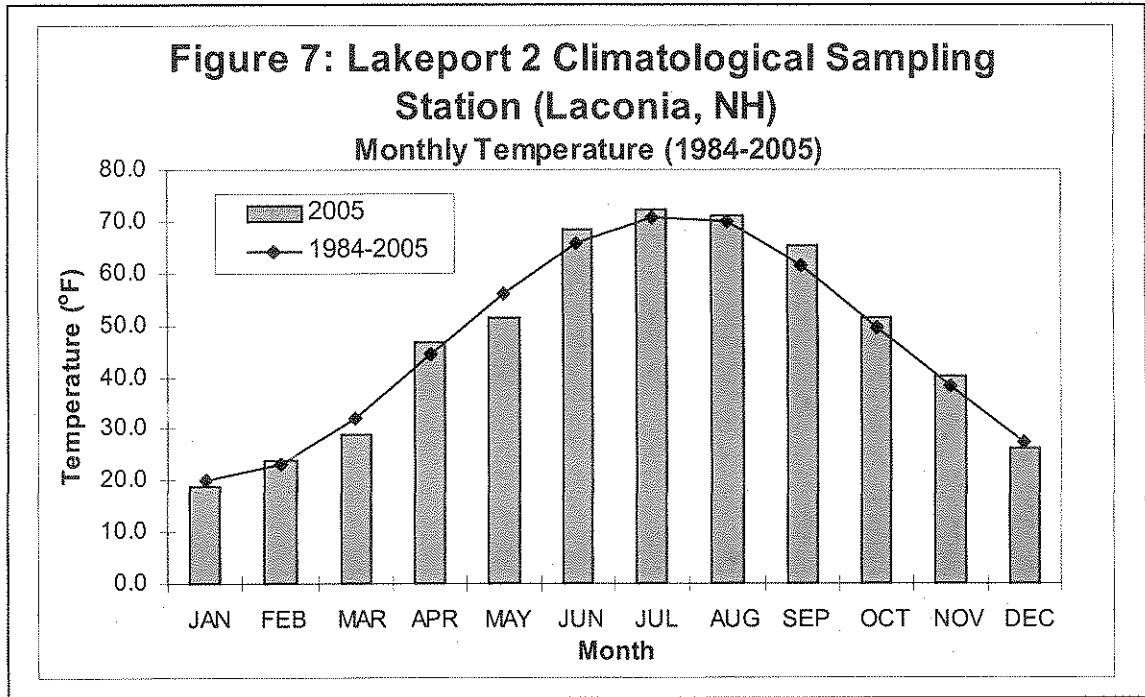


Temperature (2005)

Similar to the impact of precipitation extremes, temperature extremes can have far reaching effects on the water quality, particularly early in the year and during the summer months. Atypically warm spells can account for a rapid snowpack melt resulting in flooding and a massive influx of materials (e.g. nutrients, sediments) into our lakes during the late winter and early spring months. Early spring runoff periods coincide with minimal vegetative cover (that acts as a pollutant filter and soil stabilizer) and thus leaves the landscape highly susceptible to erosion. As we progress into the summer months, atypically warm periods can enhance both microscopic “algal” and macroscopic aquatic “weed” plant growth. During the summer growing season, above average temperatures often result in algal blooms that can reach nuisance proportions under optimal conditions. These nuisance blooms can include surface algal “scums” that cover the lake and wash up on the windward lakeshores.

During years such as 1994 and 1995, when above average temperatures characterized the summer months, participating NH LLMP lakes were generally characterized by increased algal concentrations, particularly in the shallows, where filamentous cotton candy like clouds of algae (i.e. *Mougeotia*) flourished. Other NH LLMP lakes had increased algal growth “greenness” and shallower water transparencies during these “hot” periods.

The January and February 2005 monthly temperatures were near the twenty two year average while the March, 2005 temperature was approximately 3°F below the 22 year average (Figure 7). Increasing temperatures and above average rainfall during



the months of April and May contributed to periods of heavy spring runoff. The monthly temperature averages documented between April and December 2005 oscillated from near average temperatures to slightly above or slightly below average temperatures (Figure 7). Below average temperatures during the month of May (over four °F below the twenty two year average) corresponded to some of the colder in-lake water temperatures documented in recent years through the month of June. The below average water temperatures that were documented into June might have suppressed some of the microscopic plant “algal” and macroscopic plant growth early in the 2005 sampling season.

Water Quality Impacts

Water Transparency and Dissolved “tea” Colored Water

As previously mentioned shallower water transparency readings are characteristic of most New Hampshire lakes during wet years and following short term precipitation events. Wet periods often coincide with greater concentrations of dissolved “tea” colored compounds (dissolved organic matter resulting from the breakdown of vegetation and soils) washed in from surrounding forests and wetlands. Dissolved water color is not indicative of water quality problems (although large increases in dissolved color sometimes follow large land clearing operations) but in some of our more pristine program lakes, it nevertheless has a large effect on water clarity changes. Data collected by the **Center for Freshwater Biology (CFB)** since 1985 indicate most lakes are characterized by higher dissolved “tea” colored water during wet years relative to years more typical in terms of annual precipitation levels. In some of our more highly “tea” colored lakes the early spring months are also characterized by higher dissolved color concentrations, relative to mid-summer levels, due to the heavy runoff periods that flush highly colored water into our lakes during the period of spring snowmelt and following heavy spring rains.

Sediment Loading

Sediments are continuously flushed into our lakes and ponds during periods of heavy watershed runoff, particularly during snowmelt and again during and following sporadic storm events during the summer and fall months. Many New Hampshire lakes experience water clarity decreases following storm events such as those described above. Lakes, ponds and rivers are particularly susceptible to sediment loadings in the early spring months when vegetated shoreside buffers, often referred to as riparian buffers, are reduced. With limited vegetation to trap sediments and suspended materials, a high percentage of the particulate debris and dissolved materials are flushed into the lake. Human activities such as logging, agriculture, construction and land clearing can also increase sediment displacement during and following heavy storm events throughout the year. These activities are often associated with excessive sediment loading in many of our lakes and ponds. As these materials (sediments) are transported into surface waters they can degrade water quality in a number of ways. When fine sediments (silt) enter a lake they tend to remain in the water column for relatively long periods of time. These suspended sediments can be abrasive to fish gills, ultimately leading to fish kills. Suspended sediments also reduce the available light necessary for plant growth that can result in plant die-offs and the subsequent oxygen depletion under extreme conditions.

As sediments settle out of the water column they can smother bottom dwelling aquatic organisms and fish spawning habitat. As the dead materials begin to decay the result can be noxious odors as well as stimulation of nuisance plant growth (i.e. scums along the lakebottom; new macroscopic plant growth). Note: one should keep in mind that nuisance plants such as water milfoil (*Myriophyllum heterophyllum*) will generally regenerate more rapidly than more favorable plant forms. This can result in more problematic weed beds than those present before the disturbance. Habitat changes associated with the accumulation of fine sediments and associated "muck" might also favor increased nuisance plant growth in the future. Another unfavorable attribute of sediment loading is that the sediments tend to carry with them other sorts of contaminant such as pathogens, nutrients and toxic chemicals (i.e. herbicides and pesticides).

Early symptoms of excessive sediment runoff include deposits of fine material along the lakebottom, particularly in close proximity to tributary inlets and disturbed regions previously discussed (i.e. construction sites, logging sites, etc.). Silt may be visible covering rocks or aquatic vegetation along the lakebottom. During periods of heavy overland runoff the water might appear brown and turbid which reflects the sediment load. As material collects along the lakebottom you might notice a change in the weed composition reflecting a change in the substrate type (note: aquatic plants will display natural changes in abundance and distribution, so be careful not to jump to hasty conclusions). If excessive sediment loading is suspected, take a closer look in these areas and assess whether or not the change is associated with sediment loading (look for the warning signs discussed above) or whether the changes might be attributable to other factors.

Nutrient Loading

Nutrient loading is often greatest during heavy precipitation events, particularly during the periods of heavy watershed runoff. Phosphorus is generally considered the limiting nutrient for excessive plant and algal growth in New Hampshire lakes. Elevated phosphorus concentrations are generally most visible when documented in our tributary inlets where nutrients are concentrated in a relatively small volume of water.

Much of the phosphorus entering our lakes is attached to particulate matter (i.e. sediments, vegetative debris), but may also include dissolved phosphorus associated with fertilizer applications and septic system discharge.

Microscopic "Algal" and Macroscopic "Weed" Plant Growth

Historical Lakes Lay Monitoring Program data indicate most lakes experience "algal blooms" during years with above average summer temperatures (June, July and August) while years with heavy precipitation are also associated with an increased frequency and occurrence of "algal blooms" among participating LLMP lakes. Algal blooms are often green water events associated with decreases in water clarity due to their ability to absorb and scatter light within the water column, but can also accumulate near the lake bottom in shallow areas as "mats" or on the water surface as "scums" and "clouds". During some years, such as 1996, the algal blooms are predominantly green water events composed of algae distributed within the water column. New Hampshire lakes were particularly susceptible to algal blooms in 1996 as a function of the heavy runoff associated with an atypically wet year. Wet years such as 1996 can be particularly hard on lakes where excessive fertilizer applications, agricultural practices and construction activities favor the displacement of nutrients into surface waters. The occasional formation of certain algal blooms is a naturally occurring phenomenon and is not necessarily associated with changes in lake productivity. However, increases in the occurrence of bloom conditions can be a sign of eutrophication (the "greening" of a lake). Shifts from benign (clean water) forms to nuisance (polluted water) cyanobacterial forms such as *Anabaena*, *Aphanizomenon* and *Oscillatoria*, can also be a warning sign that improper land use practices are contributing excessive nutrients into the lake.

Filamentous cotton-candy like "clouds" of the nuisance green algae, *Mougeotia* and related species, have been well documented in 1994 and 1995 when the temperatures during the months of June and July were well above normal. These algal "clouds" often develop within nearshore weed beds where they can be seen along the lakebottom and tend to flourish during warm periods. During cooler years, this type of algal growth is kept "in check" and generally does not reach nuisance proportions.

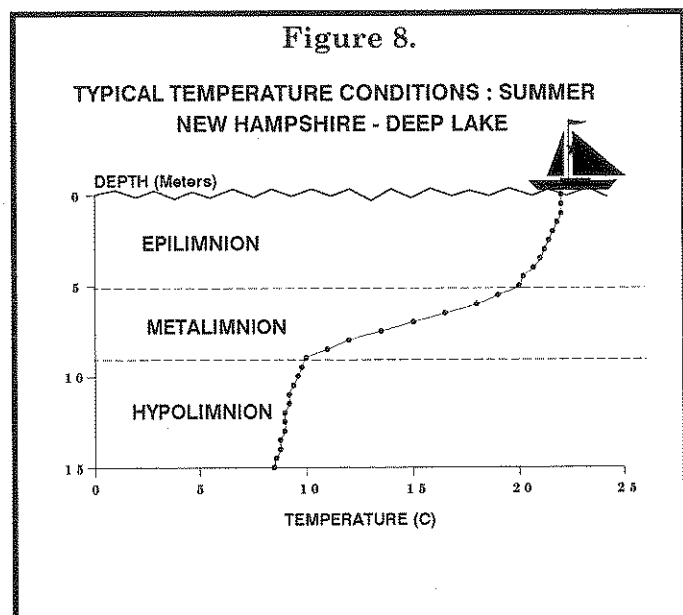
In other lakes, metalimnetic algae, algae which tend to grow in a thin layer along the thermocline gradient in a lake's middle depths, sometimes migrate up towards the lake surface causing a "bloom" event. If these algae are predominantly "nuisance" forms, like certain green or blue-green algae, they can be an early indication of nutrient loading.

DISCUSSION OF LAKE AND STREAM MONITORING MEASUREMENTS

The section below details the important concepts involved for the various testing procedures used in the New Hampshire Lakes Lay Monitoring Program. Certain tests or sampling performed at the time of the optional Center for Freshwater Biology field trip are indicated by an asterisk (*).

Thermal Stratification in the Deep Water Sites

Lakes in New Hampshire display distinct patterns of temperature stratification, that develop as the summer months progress, where a layer of warmer water (the **epilimnion**) overlies a deeper layer of cold water (**hypolimnion**). The layer that separates the two regions characterized by a sharp drop in temperature with depth is called the **thermocline** or **metalimnion** (Figure 8). Some shallow lakes may be continually mixed by wind action and will never stratify. Other lakes may only contain a developed epilimnion and metalimnion.



Water Transparency

Secchi Disk depth is a measure of the water transparency. The deeper the depth of Secchi Disk disappearance, the more transparent the lake water; light penetrates deeper if there is little dissolved and/or particulate matter (which includes both living and non-living particles) to absorb and scatter it.

In the shallow areas of many lakes, the Secchi Disk will hit bottom before it is able to disappear from view (what is referred to as a "Bottom Out" condition). Thus, Secchi Disk measurements are generally taken over the deepest sites of a lake. Transparency values greater than 4 meters are typical of clear, unproductive lakes while transparency values less than 2.5 meters are generally an indication of highly productive lakes. Water transparency values between 2.5 meters and 4 meters are generally considered indicative of moderately productive lakes.

Chlorophyll α

The chlorophyll α concentration is a measurement of the standing crop of phytoplankton and is often used to classify lakes into categories of productivity called trophic states. **Eutrophic** lakes are highly productive with large concentrations of algae and aquatic plants due to nutrient enrichment. Characteristics include accumulated organic matter in the lake basin and lower dissolved oxygen in the bottom waters. Summer chlorophyll α concentrations average above 7 mg m³ (7 milligrams per cubic meter; 7 parts per billion). **Oligotrophic** lakes have low productivity and low nutrient levels and average summer chlorophyll α concentrations that are generally less than 3 mg m³. These lakes generally have cleaner bottoms and high dissolved oxygen levels throughout. **Mesotrophic** lakes are intermediate in productivity with concentrations of chlorophyll α generally between 3 mg m³ and 7 mg m³. Testing is sometimes done to check for **metalimnetic algal populations**, algae that layer out at the thermocline and generally go undetected if only epilimnetic (point or integrated) sampling is undertaken. Chlorophyll concentrations of a water sample collected in the thermocline is compared to the integrated epilimnetic sample. Greater chlorophyll levels of the point sample, in conjunction with microscopic examination of the samples (see Phytoplankton section below), confirm the presence of such a population of algae. These populations should be monitored as they may be an indication of increased nutrient loading into the lake.

Turbidity *

Turbidity is a measure of suspended material in the water column such as sediments and planktonic organisms. The greater the turbidity of a given water body the lower the Secchi Disk transparency and the greater the amount of particulate matter present. Turbidity is measured as nephelometric turbidity units (NTU), a standardized method among researchers. Turbidity levels are generally low in New Hampshire reflecting the pristine condition of the majority of our lakes and ponds. Increasing turbidity values can be an indication of increasing lake productivity or can reflect improper land use practices within the watershed which destabilize the surrounding landscape and allow sediment flushing into the lake.

While Secchi Disk measurements will integrate the clarity of the water column from the surface waters down to the depth of disappearance, turbidity measurements are collected at discrete depths from the surface down to the lake bottom. Such discrete sampling can identify layering algal populations (previously discussed) that are undetectable when measuring Secchi Disk transparency alone.

Dissolved Color

The dissolved color of lakes is generally due to dissolved organic matter from **humic substances**, which are naturally-occurring polyphenolic compounds leached from decayed vegetation. Highly colored or "stained" lakes have a "tea" color. Such substances generally do not threaten water quality except as they diminish sunlight penetration into deep waters. Increases in dissolved watercolor can be an indication of increased development within the watershed as many land clearing activities (construction, deforestation, and the resulting increased run-off) add additional organic material to lakes. Natural fluctuations of dissolved color occur when storm events increase drainage from wetlands areas within the watershed. As suspended sediment is a diffi-

cult and expensive test to undertake, both dissolved color and chlorophyll information are important when interpreting the Secchi Disk transparency

Dissolved color is measured on a comparative scale that uses standard chloroplatinate dyes and is designated as a color unit or ptu. Lakes with color below 10 ptu are very clear, 10 to 20 ptu are slightly colored, 20 to 40 ptu are lightly tea colored, 40 to 80 ptu are tea colored and greater than 80 ptu indicates highly colored waters. Generally the majority of New Hampshire lakes have color between 20 to 30 ptu.

Total Phosphorus

Of the two "nutrients" most important to the growth of aquatic plants, nitrogen and phosphorus, it is generally observed that phosphorus is the more limiting to plant growth, and therefore the more important to monitor and control. Phosphorus is generally present in lower concentrations, and its sources arise primarily through human related activity in a watershed. Nitrogen can be fixed from the atmosphere by many bloom-forming blue-green bacteria, and thus it is difficult to control. The total phosphorus includes all dissolved phosphorus as well as phosphorus contained in or adhered to suspended particulates such as sediment and plankton. As little as 10 parts per billion of phosphorus in a lake can cause an algal bloom.

Generally, in the more pristine lakes, phosphorus values are higher after spring melt when the lake receives the majority of runoff from its surrounding watershed. The nutrient is used by the algae and plants which in turn die and sink to the lake bottom causing surface water phosphorus concentrations to decrease as the summer progresses. Lakes with nutrient loading from human activities and sources (Agriculture, Logging, Sediment Erosion, Septic Systems, etc.) will show greater concentrations of nutrients as the summer progresses or after major storm events.

Streamflow

Streamflow, when collected in conjunction with depth contour information, is a measure of the volume of water traversing a given stream stretch over a period of time and is often expressed as cubic meters per second. Knowledge of the streamflow is important when determining the amount of nutrients and other pollutants that enter a lake. Knowledge of the streamflow in conjunction with nutrient concentrations, for instance, will provide the information necessary to calculate phosphorus loading values and will in turn be useful in discerning the more impacted areas within a watershed.

pH *

The pH is a way of expressing the acidic level of lake water, and is generally measured with an electrical probe sensitive to hydrogen ion activity. The pH scale has a range of 1 (very acidic) to 14 (very "basic" or alkaline) and is logarithmic (i.e.: changes in 1 pH unit reflect a ten times difference in hydrogen ion concentration). Most aquatic organisms tolerate a limited range of pH and most fish species require a pH of 5.5 or higher for successful growth and reproduction.

Alkalinity

Alkalinity is a measure of the buffering capacity of the lake water. The higher the value the more acid that can be neutralized. Typically lakes in New Hampshire have

low alkalinities due to the absence of carbonates and other natural buffering minerals in the bedrock and soils of lake watersheds.

Decreasing alkalinity over a period of a few years can have serious effects on the lake ecosystem. In a study on an experimental acidified lake in Canada by Schindler, gradual lowering of the pH from 6.8 to 5.0 in an 8-year period resulted in the disappearance of some aquatic species, an increase in nuisance species of algae and a decline in the condition and reproduction rate of fish. During the first year of Schindler's study the pH remained unchanged while the alkalinity declined to 20 percent of the pre-treatment value. The decline in alkalinity was sufficient to trigger the disappearance of zooplankton species, which in turn caused a decline in the "condition" of fish species that fed on the zooplankton.

The analysis of alkalinity employed by the **Center for Freshwater Biology** includes use of a dilute titrant allowing an order of magnitude greater sensitivity and precision than the standard method. Two endpoints are recorded during each analysis. The first endpoint (gray color of dye; pH endpoint of 5.1) approximates low level alkalinity values, while the second endpoint (pink dye color; pH endpoint of 4.6) approximates the alkalinity values recorded historically, such as NH Fish and Game data, with the methyl-orange endpoint method.

The average alkalinity of lakes throughout New Hampshire is low, approximately 6.5 mg per liter (calcium carbonate alkalinity). When alkalinity falls below 2 mg per liter the pH of waters can greatly fluctuate. Alkalinity levels are most critical in the spring when acid loadings from snowmelt and run-off are high, and many aquatic species are in their early, and most susceptible, stages of their life cycle.

Specific Conductivity *

The specific conductance of a water sample indicates concentrations of dissolved salts. Leaking septic systems and deicing salt runoff from highways can cause high conductivity values. Fertilizers and other pollutants can also increase the conductivity of the water. Conductivity is measured in micromhos (the opposite of the measurement of resistance **ohms**) per centimeter, more commonly referred to as micro-Siemans (μS).

Dissolved Oxygen and Free Carbon Dioxide *

Oxygen is an essential component for the survival of aquatic life. Submergent plants and algae take in carbon dioxide and create oxygen through **photosynthesis** by day. **Respiration** by both animals and plants uses up oxygen continually and creates **carbon dioxide**. Dissolved oxygen profiles determine the extent of declining oxygen concentrations in the lower waters. High carbon dioxide values are indicative of low oxygen conditions and accumulating organic matter. For both gases, as the temperature of the water decreases, more gas can be dissolved in the water.

The typical pattern of clear, unproductive lakes is a slight decline in hypolimnetic oxygen as the summer progresses. Oxygen in the lower waters is important for maintaining a fit, reproducing, cold water fishery. Trout and salmon generally require oxygen concentrations above 5 mg per liter (parts per million) in the cool deep waters. On the other hand, carp and catfish can survive very low oxygen conditions. Oxygen above the lake bottom is important in limiting the release of nutrients from the sediments and minimizing the collection of undecomposed organic matter.

Bacteria, fungi and other **decomposers** in the bottom waters break down organic matter originating from the watershed or generated by the lake. This process uses up oxygen and produces carbon dioxide. In lakes where organic matter accumulation is high, oxygen depletion can occur. In highly stratified eutrophic lakes the entire hypolimnion can remain unoxygenated or **anaerobic** until fall mixing occurs.

The oxygen peaks occurring at surface and mid-lake depths during the day are quite common in many lakes. These characteristic **heterograde oxygen curves** are the result of the large amounts of oxygen, the by-product of photosynthesis, collecting in regions of high algal concentrations. If the peak occurs in the thermocline of the lake, metalimnetic algal populations (discussed above) may be present.

Underwater Light *

Underwater light available to photosynthetic organisms is measured with an **underwater photometer** which is much like the light meter of a camera (only waterproofed!). The **photic zone** of a lake is the volume of water capable of supporting photosynthesis. It is generally considered to be delineated by the water's surface and the depth that light is reduced to one percent surface irradiance by the absorption and scattering properties of the lake water. The one percent depth is sometimes termed the **compensation depth**. Knowledge of light penetration is important when considering lake productivity and in studies of submerged vegetation. Discontinuity (abrupt changes in the slope) of the profiles could be due to metalimnetic layering of algae or other particulates (discussed above). The underwater photometer allows the investigator to measure light at depths below the Secchi Disk depth to supplement the water clarity information.

Indicator Bacteria *

Certain disease causing organisms, pathogenic bacteria, viruses and parasites, can be spread through contact with polluted waters. Faulty septic systems, sewer leaks, combined sewer overflows and the illegal dumping of wastes from boats can contribute fecal material containing these pathogens. Typical water testing for pathogens involves the use of detecting coliform bacteria. These bacteria are not usually considered harmful themselves but they are relatively easy to detect and can be screened for quickly. Thus, they make good surrogates for the more difficult to detect pathogens.

Total coliform includes all coliform bacteria that arise from the gut of animals or from vegetative materials. **Fecal coliform** are those specific organisms that inhabit the gut of warm blooded animals. Another indicator organism **Fecal streptococcus** (sometimes referred to as **enterococcus**) also can be monitored. The ratio of fecal coliform to fecal strep may be useful in suggesting the type of animal source responsible for the contamination. In 1991, the State of New Hampshire changed the indicator organism of preference to *E. Coli* which is a specific type of fecal coliform bacteria thought to be a better indicator of human contamination. The new state standard requires Class A "bathing waters" to be under 88 organisms (referred to as colony forming units; cfu) per 100 milliliters of lakewater.

Ducks and geese are often a common cause of high coliform concentrations at specific lake sites. While waterfowl are important components to the natural and aesthetic qualities of lakes that we all enjoy, it is poor management practice to encourage these birds by feeding them. The lake and surrounding area provides enough healthy

and natural food for the birds and feeding them stale bread or crackers does nothing more than import additional nutrients into the lake and allows for increased plant growth. As birds also are a host to the parasite that causes "swimmers itch", waterfowl roosting areas offer a greater chance for infestation to occur. Thus while leaving offerings for our feathered friends is enticing, the results can prove to be detrimental to the lake system and to human health.

Phytoplankton *

The planktonic community includes microbial organisms that represent diverse life forms, containing photosynthetic as well as non-photosynthetic types, and including bacteria, algae, crustaceans and insect larvae (the insect larvae and zooplankton are discussed below in separate sections). Because planktonic algae or "phytoplankton" tend to undergo rapid seasonal cycles on a time scale of days and weeks, the levels of populations found should be considered to be most representative of the time of collection and not necessarily of other times during the ice-free season, especially the early spring and late fall periods.

The composition and concentration of phytoplankton can be indicative of the trophic status of a lake. Seasonal patterns do occur and must be considered. For example **diatoms**, tend to be most abundant in April-June and October-November, in the surface or epilimnetic layers of New Hampshire lakes. As the summer progresses, the dominant types might shift to **green algae** or **golden algae**. By late season **Blue-green bacteria** generally dominate. In nutrient rich lakes, nuisance green algae and/or bluegreen bacteria might dominate continually. After fall mixing diatoms might again be found to bloom.

Zooplankton *

There are three groups of zooplankton that are generally prevalent in lakes: the **protozoa**, **rotifers** and **crustaceans**. Most research has been devoted to the last two groups although protozoa may be found in substantial amounts. Of the rotifers and the crustaceans, time and budgetary constraints usually make it necessary to sample only the larger zooplankton (macrozooplankton; larger than 80 or 150 microns; 1 million microns make up a meter). Thus, zooplankton analysis is generally restricted only to the larger crustaceans. Crustacean zooplankton are very sensitive to pollutants and are commonly used to indicate the presence of toxic substances in water. The crustaceans can be divided into two groups, the **cladocerans** (which include the "water fleas") and the **copepods**.

Macrozooplankton are an important component in the lake system. The filter feeding of the herbivorous ("grazing") species may control the population size of selected species of phytoplankton. The larger zooplankton can be an important food source for juvenile and adult planktivorous fish. All zooplankton play a part in the recycling of nutrients within the lake. Like the phytoplankton, zooplankton, tend to undergo rapid seasonal cycles. Thus, the zooplankton population density and diversity should be considered to be most representative of the time of collection and not necessarily of other times during the ice-free season, especially the early spring and late fall periods.

Macroinvertebrates *

Macroinvertebrates generally refer to the aquatic insect community living near the bottom substrate (i.e. sediments) while other invertebrate groups such as the crayfish, leeches and the aquatic worms are also included. Like the phytoplankton and zooplankton, previously discussed, the macroinvertebrates undergo seasonal cycles and are most representative of conditions for particular periods of the year. The mayflies are probably the most well known example of a seasonal aquatic macroinvertebrate as mayfly populations metamorphosize into adults as the water temperatures increase in the spring and thus giving rise to the name "mayflies". Macroinvertebrates are also sensitive to environmental conditions such as streamflow, temperature and food availability and are most representative of particular habitats along the stream continuum (i.e. some organisms prefer slower moving stream reaches while others prefer rapidly flowing waters).

Macroinvertebrates are an essential component to a healthy aquatic habitat. Macroinvertebrates help decompose organic matter entering the system such as leaves and twigs and also serve as a food source for many fish species.

While some macroinvertebrates are capable of breathing air as we do, others have gills and utilize oxygen dissolved in the water much as fish do. Macroinvertebrates also vary in their tolerance to depleting dissolved oxygen concentrations making them a good indicator of pollutants coming into the water body. The caddis flies (Trichoptera), the mayflies (Ephemeroptera) and the stoneflies (Plecoptera) are often considered highly sensitive to pollution while the "true" flies (Diptera) are often considered highly tolerant to pollution. However, exceptions to the above categorizations are often encountered.

A variety of indices have been proposed to characterize water bodies over a gradient of pollution levels ranging from least polluted to most polluted scenarios and often designated by assigning a numerical delineator (i.e. 1 is least polluted while 10 is most polluted). Such an index, the Hilsenhoff Biotic Index (HBI), or a modification thereof, is commonly used by stream monitoring programs around the country. Macroinvertebrate data are useful in discerning the more impacted areas within the watershed where corrective efforts should be directed. Unlike chemical measurements that represent ambient conditions in the water body, the macroinvertebrate community composition integrates the water quality conditions over a longer period (months to years) and can identify "hot" spots missed by chemical sampling. If you are interested in more information regarding macroinvertebrate monitoring contact the LLMP coordinator.

Fish Condition

The assessment of fish species "health" is another biological indicator of water quality. Because fish are at the top of the food chain, their condition should reflect not only water quality changes that affect them directly but also those changes that affect their food supply. The fish condition index utilized by the **New Hampshire Fish Condition Program** is based on two components; fish scale analysis and a fish condition index.

Like tree trunks, fish scales have annual growth rings (annuli) that reflect their growth history and hence, provide a long-term record of past conditions in the lake. The fish condition index, based upon length and weight measurements, is a good indicator of the fish's health at the time of collection.

The resulting fish condition data can be compared among different lakes or among different years, or the index for a particular species can be compared to standard length-to-weight relationships that have been developed by fisheries biologists for many important fish species. In the end, the "health" of the various fish species reflects the overall water quality in the respective lake or pond.

Understanding Lake Aging (Eutrophication)

by: **Robert Craycraft** Educational Program Coordinator,
New Hampshire Lakes Lay Monitoring Program
University of New Hampshire
G18 Spaulding Hall, Durham, NH 03824
603-862-3696 FAX: 603-862-0107
email: bob.craycraft@unh.edu

and **Jeff Schloss** UNH Cooperative Extension Water Resources Specialist

A common concern among New Hampshire Lakes Lay Monitoring Program (NH LLMP) participants is a perceived increase in the density and abundance of aquatic plants in the shallows, increases in the amount of microscopic plant “algae” growth (detected as greener water), and water transparency decreases; what is known as **eutrophication**. Eutrophication is a natural process by which all lakes age and progress from clear, pristine lakes to green, nutrient enriched lakes on a geological time frame of thousands of years. Much like the fertilizers applied to our lawns, nutrients that enter our lakes stimulate plant growth and culminate in greener (and in turn less clear) waters. Some lakes age at a faster rate than others due to naturally occurring attributes: watershed area relative to lake area, slope of the land surrounding the lake, soil type, mean lake depth, etc. Since our New Hampshire lakes were created during the last ice-age which ended about 10,000 years ago, we should have a natural continuum of lakes ranging from extremely pristine to very enriched.

Classification criteria are often used to categorize lakes into what are known as **tro- phic states**, in other words, levels of lake plant and algae productivity or “greenness” Refer to Table 6 below for a summary of commonly used eutrophication parameters.

Table 6: Eutrophication Parameters and Categorization

Parameter	Oligotrophic “pristine”	Mesotrophic “transitional”	Eutrophic “enriched”
Chlorophyll a (ug/l) *	<3.0	3.0-7.0	>7.0
Water Transparency (meters) *	>4.0	2.5-4.0	<2.5
Total Phosphorus (ug/l) *	<15.0	15.0-25.0	>25.0
Dissolved Oxygen (saturation) #	high to moderate	moderate to low	low to zero
Macroscopic Plant (Weed) Abundance	low	moderate	high

* Denotes classification criteria employed by Forsberg and Ryding (1980).

Denotes dissolved oxygen concentrations near the lakebottom.

Oligotrophic lakes are considered “unproductive” pristine systems and are characterized by high water clarities, low nutrient concentrations, low algae concentrations, minimal levels of aquatic plant “weed” growth, and high dissolved oxygen concentrations near the lake bottom. **Eutrophic** lakes are considered “highly productive” enriched systems characterized by low water transparencies, high nutrient concentrations, high algae concentrations, large stands of aquatic plants and very low dissolved oxygen concentrations near the lake bottom. **Mesotrophic** lakes have qualities between those of oligotrophic and eutrophic lakes and are characterized by moderate water transparencies, moderate nutrient concentrations, moderate algae growth, moderate aquatic plant “weed” growth and decreasing dissolved oxygen concentrations near the lake bottom.

Is a pristine, oligotrophic, lake “better than” an enriched, eutrophic, lake? Not necessarily! As indicated above, lakes will naturally exhibit varying degrees of productivity. Some lakes will naturally be more susceptible to eutrophication than others due to their natural attributes and in turn have aged more rapidly. This is not necessarily a bad thing as our best bass fishing lakes tend to be more mesotrophic to eutrophic than oligotrophic; an ultra-oligotrophic lake (extremely pristine) will not support a very healthy cold water fishery. However, human related activities can augment the aging process (what is known as cultural eutrophication) and result in a transition from a pristine system to an enriched system in tens of years rather than the natural transitional period that should take thousands of years. Cultural eutrophication is particularly a concern for northern New England lakes where large tracts of once forested or agricultural lands are being developed, with the potential for increased sediment and nutrient loadings into our lakes which augment the eutrophication process.

Additionally, other pollutants such as heavy metals, herbicides, insecticides and petroleum products might also affect your lake’s “health”. A “healthy” lake, as far as eutrophication is concerned, is one in which the various aquatic plants and animals are minimally impacted so that nutrients and other materials are processed efficiently. We can liken this process to a well-managed pasture: nutrients stimulate the growth of grasses and other plants that are eaten by grazers like cows and sheep. As long as producers and grazers are balanced, a good amount of nutrients can be processed through the system. Impact the grazers and the grass will overgrow and nuisance weeds will appear, even if nutrients remain the same. In a lake, the producers are the algae and aquatic weeds while the grazers are the microscopic animals (**zooplankton**) and aquatic insects. These organisms can be very susceptible to a wide range of pollutants at very low concentrations. If impacted, the lake can become much more productive and the fishery will be impacted as well since these same organisms are an important food source for most fish at some stage of their life.

Development upon the landscape can negatively affect water quality in a number of ways:

- Removal of shore side vegetation and loss of wetlands - shore side vegetation (what is known as **riparian vegetation**) and wetlands provide a protective buffer that “traps” pollutants before reaching the lake. These buffers remove materials both chemically (through biological uptake) and physically (settling materials out). As riparian buffers are removed and wetlands lost, pollutant materials are more likely to enter the lake and in turn, favor declining water quality.
- Excessive fertilizer applications - fertilizers entering the lake can stimulate aquatic plant and algal growth and in extreme cases result in noxious algal blooms. Increases in algal growth tend to diminish water transparency and under extreme cases culminate in surface “scums” that can wash up on the shoreline producing unpleasant smells as the material decomposes. Excessive nutrient

concentrations also favor algal forms known to produce toxins which irritate the skin and under extreme conditions, are dangerous when ingested.

- Increased organic matter loading - organic matter (leaves, grass clippings, etc.) are a major source of nutrients in the aquatic environment. As the vegetative matter decomposes nutrients are "freed up" and can become available for aquatic plant and algal growth. In general, we are not concerned with this material entering the lake naturally (leaf senescence in the fall) but rather excessive loading of this material as occurs when residents dump or rake leaf litter and grass clippings into the lake. This material not only provides large nutrient reserves which can stimulate aquatic plant and algal growth but also makes great habitat for leaches and other potentially undesirable organisms in swimming areas.
- Septic problems - faulty septic systems are a big concern as they can be a primary source of water pollution around our lakes. Septic systems are loaded with nutrients and can also be a health threat when not functioning properly.
- Loss of vegetative cover and the creation of impervious surfaces - A forested watershed offers the best protection against pollutant runoff. Trees and tall vegetation intercept heavy rains that can erode soils and surface materials. The roots of these plants keep the soils in place, process nutrients and absorb moisture so the soils do not wash out. Impervious surfaces (paved roads, parking lots, building roofs, etc.) reduce the water's capacity to infiltrate into the ground, and in turn, go through nature's water purification system. As water seeps into the soil, pollutants are removed from the runoff through absorption onto soil particles. Biological processes detoxify pollutants and/or immobilize substances. Surface water runoff over impervious surfaces also increases water velocities that favor the transport of a greater load of suspended and dissolved pollutants into your lake.

How can you minimize your water quality impacts?

- Minimize fertilizer applications whenever possible. Most people apply far more fertilizers than necessary, with the excess eventually draining into your lake. This not only applies to those immediately adjacent to the lake but to everybody in the watershed. Pollutants in all areas of the watershed will ultimately make their way into your lake. Have your soil tested for a nominal fee (contact your county UNH Cooperative Extension Office for further information) to find out how much fertilizer and soil amendments are really needed. Sometimes just an application of crushed lime will release enough nutrients to fit the bill. If you do use fertilizer try to use low phosphorus, slow release nitrogen varieties. And remember that under the current NH Comprehensive Shoreline Protection Act (CSPA) you cannot apply any fertilizers or amendments within 25 feet of the shore.
- Don't dump leaf litter or leaves into the lake. Compost the material or take it to a proper waste disposal center. Do not fill in wetland areas. Do not create or enhance beach areas with sand (contains phosphorus, smothers aquatic habitat, fills in lake as it gets transported away by currents and wind).

- Septic systems will not function efficiently without the proper precautionary maintenance. Have your septic system inspected every two to four years and pumped out when necessary. Since the septic system is such an expensive investment often costing around \$10,000 for a complete overhaul, it is advantageous to assure proper care is taken to prolong the system's life. Additionally, following proper maintenance practices will reduce water quality degradation. Refer to:

Septic Systems, How they work and how to keep them working. \$1.00/ea University of New Hampshire Publications Center (603) 862-2346

Pipeline: Fall 1994 Vol. 15, No. 4. Maintaining Your Septic System-A Guide for Homeowners. http://www.nesc.wvu.edu/nsfc/pdf/pipline/PL_fall04.pdf

- Try to landscape and re-develop with consideration of how water flows on and off your property. Divert runoff from driveways, roofs and gutters to a level vegetated area or a rain garden so the water can be slowed, filtered and hopefully absorbed as recharge. Refer to:

A Guide to Developing and Re-Developing Shoreland Property in New Hampshire: A Blueprint to Help You Live by the Water. North Country Resource Conservation and Development Area, Inc. 103 Main Street-Suite #1., Meredith NH 03253-9266 (603) 279-6546

- Maintain shore side (riparian) vegetative cover when new construction is undertaken. For those who have pre-existing houses but lack vegetative buffers, consider shoreline plantings aimed at diminishing the pollution load into your lake. Refer to:

Planting Shoreland Areas (no charge) University of New Hampshire Cooperative Extension Publication Center. (603) 862-2346

A Guide to Developing and Re-Developing Shoreland Property in New Hampshire: A Blueprint to Help You Live by the Water. North Country Resource Conservation and Development Area, Inc. 103 Main Street-Suite #1., Meredith NH 03253-9266 (603) 279-6546

Buffers for Wetlands and Surface Waters: A Guidebook for New Hampshire Municipalities. Audubon Society of New Hampshire. 3 Silk Farm Road, Concord NH 03301 (603) 224-9909 (free for towns, \$5.00 for others).

- Review the New Hampshire Comprehensive Shoreland Protection Act (CSPA) if you have shoreland property. The CSPA sets legal regulations aimed at protecting water quality. If you have any questions regarding the act or need further information contact the *Shoreline Protection Act Coordinator* at (603) 271-3503.

REFERENCES

- American Public Health Association.(APHA) 1989. Standard Methods for the Examination of Water and Wastewater 17th edition. APHA, AWWA, WPCF.
- Baker, A.L. 1973. Microstratification of phytoplankton in selected Minnesota lakes. Ph. D. thesis, University of Minnesota.
- Carlson, R.E. 1977. A trophic state index for lakes. *Limnol. Oceanogr.* 22:361-379.
- Chase, V.P., L.S. Deming and F. Latawiec. 1995. Buffers for Wetlands and Surface Waters: A Guidebook for New Hampshire Municipalities. Audubon Society of New Hampshire.
- Craycraft, Robert and Jeffrey Schloss. Reports of the NH Lakes Lay Monitoring Program 1990-2001. UNH Center for Freshwater Biology and Cooperative Extension.
- Dates, Geoff and Jeffrey Schloss. 1998. DATA to INFORMATION: Data Management and Analysis for Coastal Monitoring Groups in New Hampshire and Maine-. Maine-NH Sea Grant and Gulf of Maine Council. September 1998. 90 pages. MSG-E-98-8. Manual.
- Edmondson, W.T. 1937. Food conditions in some New Hampshire lakes. In: Biological survey of the Androscoggin, Saco and coastal watersheds. (Report of E.E. Hoover.) New Hampshire Fish and Game Commission. Concord, New Hampshire.
- Estabrook, R.H., J.N. Connor, K.D. Warren, and M.R. Martin. 1987. New Hampshire Lakes and Ponds Inventory. Vol. III. Staff Report No. 153. New Hampshire Department of Environmental Services. Concord, New Hampshire.
- Estabrook, R.H., M.R. Martin and W.M. Henderson. 1988. New Hampshire Lakes and Ponds Inventory. Vol. IV. Staff Report No. 156. New Hampshire Department of Environmental Services. Concord, New Hampshire.
- Estabrook, R.H., M.R. Martin, P.M. McCarthy, D.J. Dubis, and W.M. Henderson. 1989. New Hampshire Lakes and Ponds Inventory. Vol. V. Staff Report No. 166. New Hampshire Department of Environmental Services. Concord, New Hampshire.
- Estabrook, R.H., P.M. McCarthy, M. O'Loan, W.M. Henderson, and D.J. Dubis. 1990. New Hampshire Lakes and Ponds Inventory. Vol. VI. NHDES-WSPCD-90-3. New Hampshire Department of Environmental Services. Concord, New Hampshire.
- Estabrook, R.H., M. O'Loan and W.M. Henderson. 1991. New Hampshire Lakes and Ponds Inventory. Vol. VII. NHDES-WSPCD-91-3. New Hampshire Department of Environmental Services. Concord, New Hampshire.
- Estabrook, R.H., M. O'Loan, W.M. Henderson and K.L. Perkins. 1992. New Hampshire Lakes and Ponds Inventory. Vol. VIII. NHDES-WSPCD-92-6. New Hampshire Department of Environmental Services. Concord, New Hampshire.

Estabrook, R.H., K. Faul and W.M. Henderson. 1993. New Hampshire Lakes and Ponds Inventory. Vol. IX. NHDES-WSPCD-93-3. New Hampshire Department of Environmental Services. Concord, New Hampshire.

Estabrook, R.H., K. Faul and W.M. Henderson. 1994. New Hampshire Lakes and Ponds Inventory. Vol. X. NHDES-WSPCD-94-4. New Hampshire Department of Environmental Services. Concord, New Hampshire.

Estabrook, R.H., W.M. Henderson and S. Ashley. 1996. New Hampshire Lakes and Ponds Inventory. Vol. XII. NHDES-WSPCD-96-6. New Hampshire Department of Environmental Services. Concord, New Hampshire.

Forsberg, C. and S.O. Ryding. 1980. Eutrophication parameters and trophic state indices in 30 Swedish waste-water receiving lakes. Arch. Hydrobiol. 89:189-207

Gallup, D.N. 1969. Zooplankton distributions and zooplankton-phytoplankton relationships in a mesotrophic lake. Ph.D. Thesis, University of New Hampshire.

Haney, J.F. and D.J. Hall. 1973. Sugar-coated Daphnia: a preservation technique for Cladocera. Limnol. Oceanogr. 18:331-333.

Hoover, E.E. 1936. Preliminary biological survey of some New Hampshire lakes. Survey report no. 1. New Hampshire Fish and Game Department. Concord, New Hampshire.

Hoover, E.E. 1937. Biological survey of the Androscoggin, Saco, and coastal watersheds. Survey report no. 2. New Hampshire Fish and Game Department. Concord, New Hampshire.

Hoover, E.E. 1938. Biological Survey of the Merrimack watershed. Survey report no. 3. New Hampshire Fish and Game Department. Concord, New Hampshire.

Hutchinson, G.E. 1967. A treatise on limnology, Vol. 2. John Wiley and Sons, New York.

Lind, O.T. 1979. Handbook of common methods in limnology. C.V. Mosby, St. Louis.

Lorenzen, M.W. 1980. Use of chlorophyll-Secchi Disk relationships. Limnol. Oceanogr. 25:371-372.

McCafferty, W.P. 1983. Aquatic Entomology: The Fishermen's and Ecologists' Illustrated Guide to Insects and their relatives. Jones and Bartlett Publishers. Boston MA.

Merritt, R.W. and K.W. Cummins. 1995. An Introduction to the Aquatic Insects of North America. Kendall/Hunt Publishing Company. Dubuque, Iowa

New Hampshire Water Supply and Pollution Control Commission. 1981. Classification and priority listing of New Hampshire lakes. Vol. II (Parts 1-6). Staff report no. 121. Concord, New Hampshire.

New Hampshire Water Supply and Pollution Control Commission. 1982. Classification and priority listing of New Hampshire lakes. Vol. III. Staff report no. 121. Concord, New Hampshire.

New Hampshire Water Supply and Pollution Control Commission. 1983. New Hampshire Lakes and Ponds Inventory. Vol. I. Staff report no. 133. Concord, New Hampshire.

New Hampshire Water Supply and Pollution Control Commission. 1985. New Hampshire Lakes and Ponds Inventory. Vol. II. Staff report no. 133. Concord, New Hampshire.

Newell, A.E. 1960. Biological survey of the lakes and ponds in Coos, Grafton and Carroll Counties. Survey report no. 8a. New Hampshire Fish and Game Department. Concord, New Hampshire.

Newell, A.E. 1970. Biological survey of the lakes and ponds in Cheshire, Hillsborough and Rockingham Counties. Survey report no. 8c. New Hampshire Fish and Game Department. Concord, New Hampshire.

Newell, A.E. 1977. Biological survey of the lakes and ponds in Sullivan, Merrimack, Belknap and Strafford Counties. Survey report no. 8b. New Hampshire Fish and Game Department. Concord, New Hampshire.

Schindler, D.W., et al. 1985. Long-term ecosystem stress: Effects of years of experimental acidification on a small lake. *Science*. 228:1395-1400.

Schloss, Jeffrey A. 2002. GIS Watershed Mapping: Developing and Implementing a Watershed Natural Resources Inventory. In: Handbook of Water Sensitive Ecological Planning & Design- Proceedings of an International Symposium. February 25 - 26, 2000 Harvard University Graduate School of Design. CRC Press, Boca Raton, FL, Chapter II.12.

Schloss, J. and J. Connor. 2001. Development of Statewide Nutrient (P) Loading Coefficients Through Geographic Information System Aided Analysis. Final Project Report. UNH Water Resources Research Center. Durham, NH.

Schloss, Jeffrey A. 2000. An Early Success of the Clean Water Action Plan: The Lake Chocorua Project. Proceedings of the 2nd National Water Quality Monitoring Council National Monitoring Conference: Monitoring for the Millennium. April 25-27, 2000 Austin Texas. Environmental Protection Agency, US Geological Survey, Ground Water Protection Council.

Schloss, Jeffrey A. 1999. Squam Lakes Watershed Natural Resources Inventory: A Case Study for GIS Analysis in a Multijurisdictional Watershed. 1999. Proceedings of the Tijuana River Watershed Workshop. May 5-8, 1999. Tijuana Mexico. NOAA Coastal Services Center publication.

Schloss, J.A., A.L. Baker and J.F. Haney. 1989. Over a decade of citizen volunteer monitoring in New Hampshire: The New Hampshire Lakes Lay Monitoring Program. *Lake and Reservoir Management*.

Sprules, W.G. 1980. Zoogeographic patterns in size structure of zooplankton communities with possible applications to lake ecosystem modeling and management. in W.C.

Kerfoot ed. Evolution and Ecology of Zooplankton Communities. University Press of New England. Dartmouth. pp. 642-656.

Uttermohl, H. 1958. Improvements in the quantitative methods of phytoplankton study. Mitt. int. Ver. Limnol. 9:1-25.

U.S. Environmental Protection Agency. 1979. A manual of methods for chemical analysis of water and wastes. Office of Technology Transfer, Cincinnati. PA-600/4-79-020.

Vollenweider, R.A. 1969. A manual on methods for measuring primary productivity in aquatic environments. International Biological Programme. Blackwell Scientific Publications, Oxford.

Warfel, H.E. 1939. Biological survey of the Connecticut Watershed. Survey Report 4. N.H. Fish and Game. Concord, New Hampshire.

Wetzel, R.G. 1983. Limnology. Saunders College Publishing, Philadelphia.

REPORT FIGURES

Figure 9. Location of the Newfound Lake sampling stations: Sites 1 Deep, 2 Mayhew, 3 Pasquaney, 4 Loon Island, 5 Beachwood, 6 Fowler River, 7 Cockermouth River and 8 Follansbee Cove.

Newfound Lake

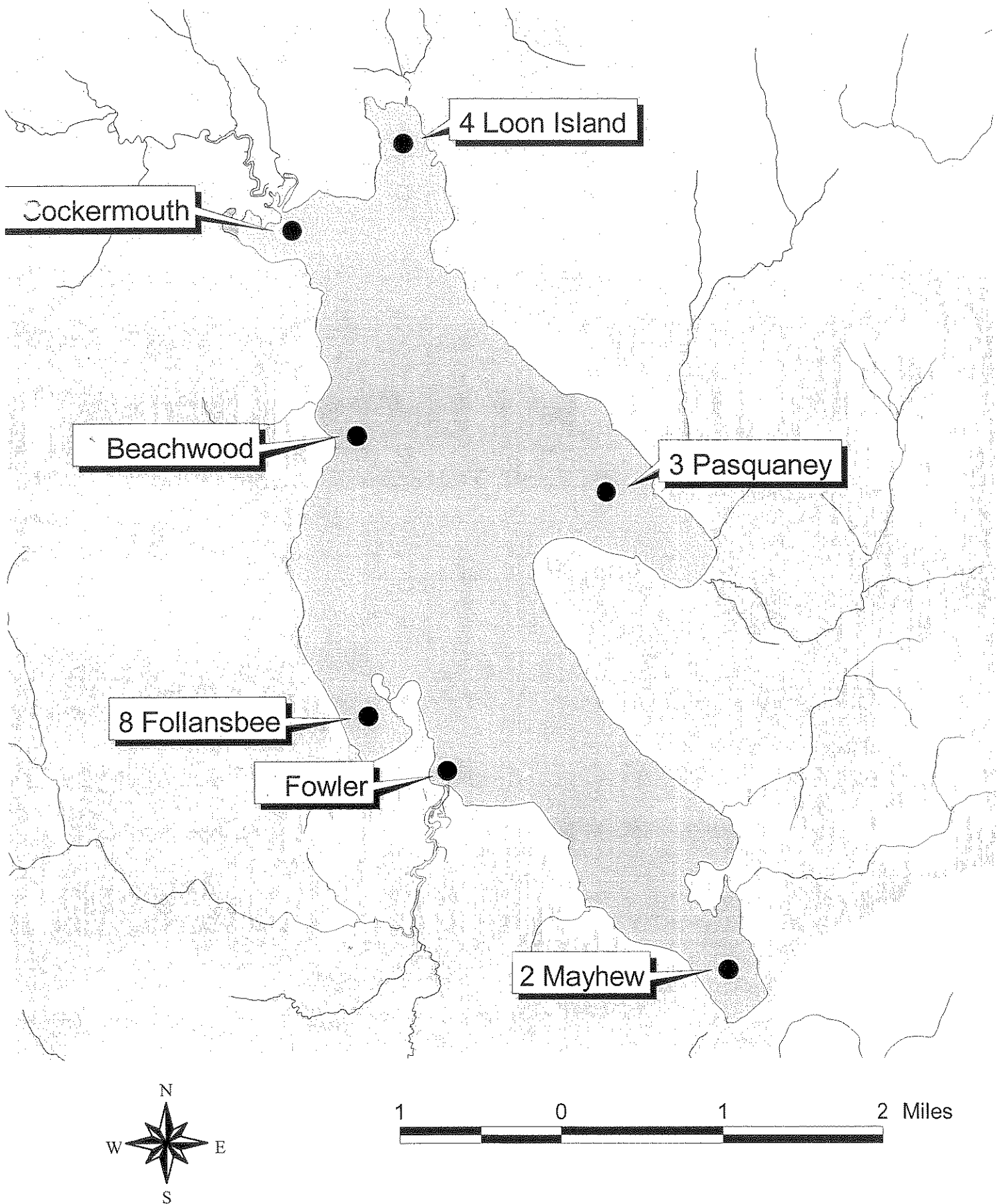
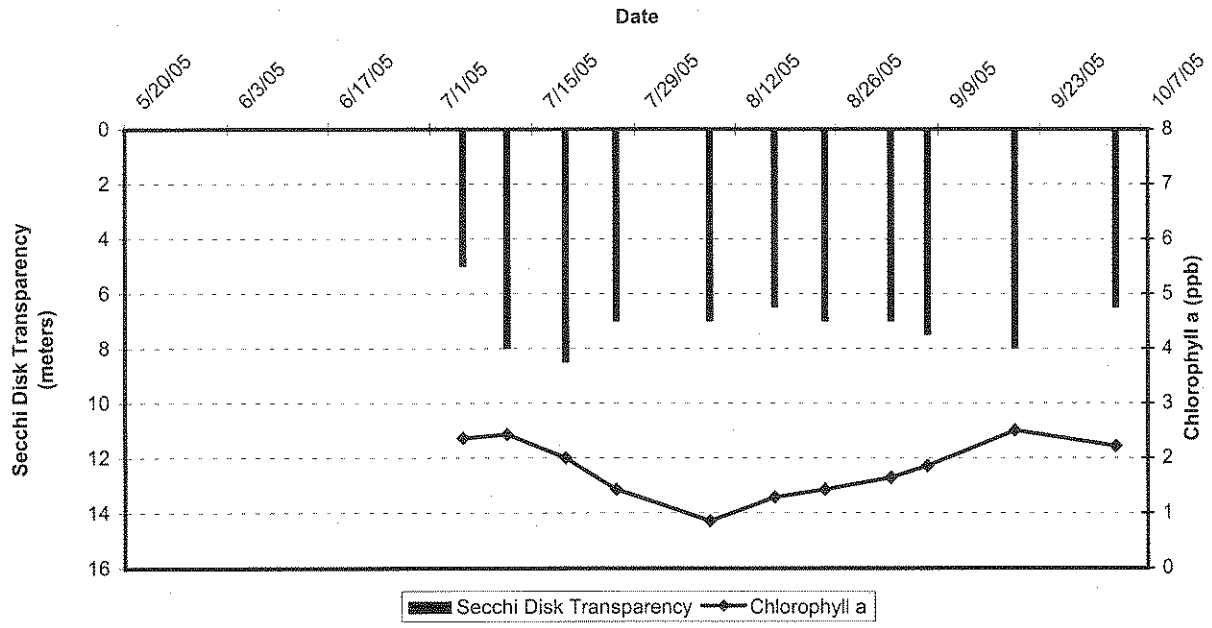


Figure 10. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll a trends for Site 2 Mayhew. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the chlorophyll a data are reported to the nearest 0.1 parts per billion (ppb).

Figure 11. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 2 Mayhew. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the dissolved color data are reported to the nearest 0.1 chlorophyll a unit (CPU).

Note: the overlay of the Secchi Disk data with chlorophyll a and dissolved color data is intended to provide a visual depiction of the impacts of chlorophyll a and dissolved color on water transparency measurements (e.g. higher chlorophyll a and dissolved color concentrations often correspond to shallower water transparencies).

Newfound -- Site 2 Mayhew (2005 Seasonal Data)



Newfound -- Site 2 Mayhew (2005 Seasonal Data)

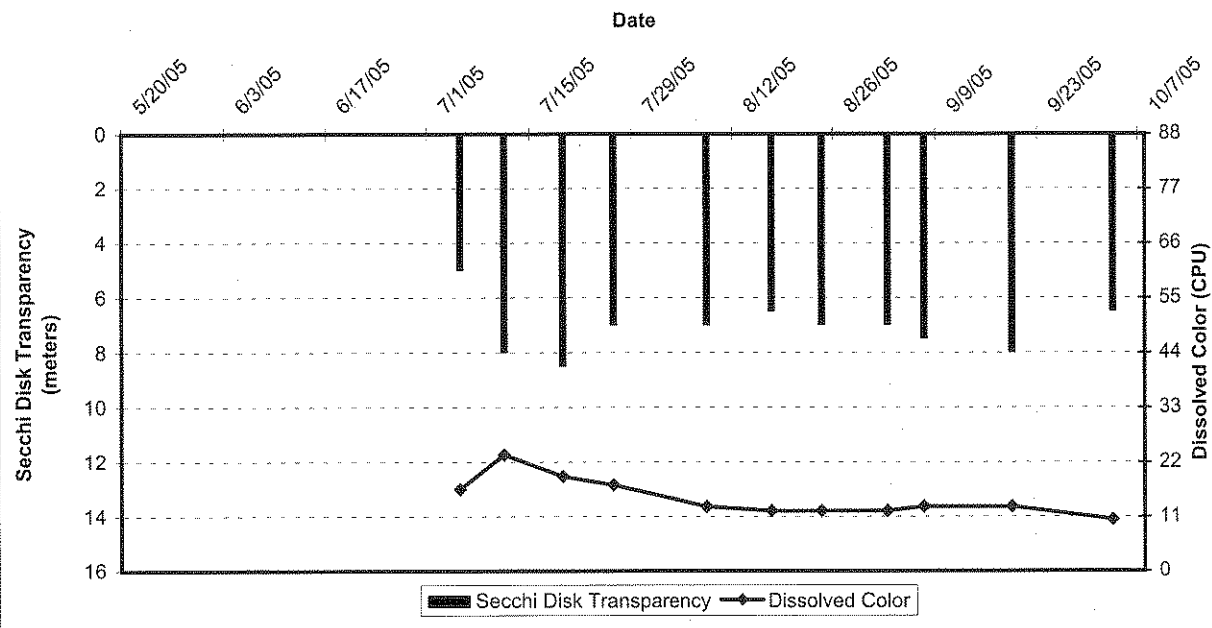
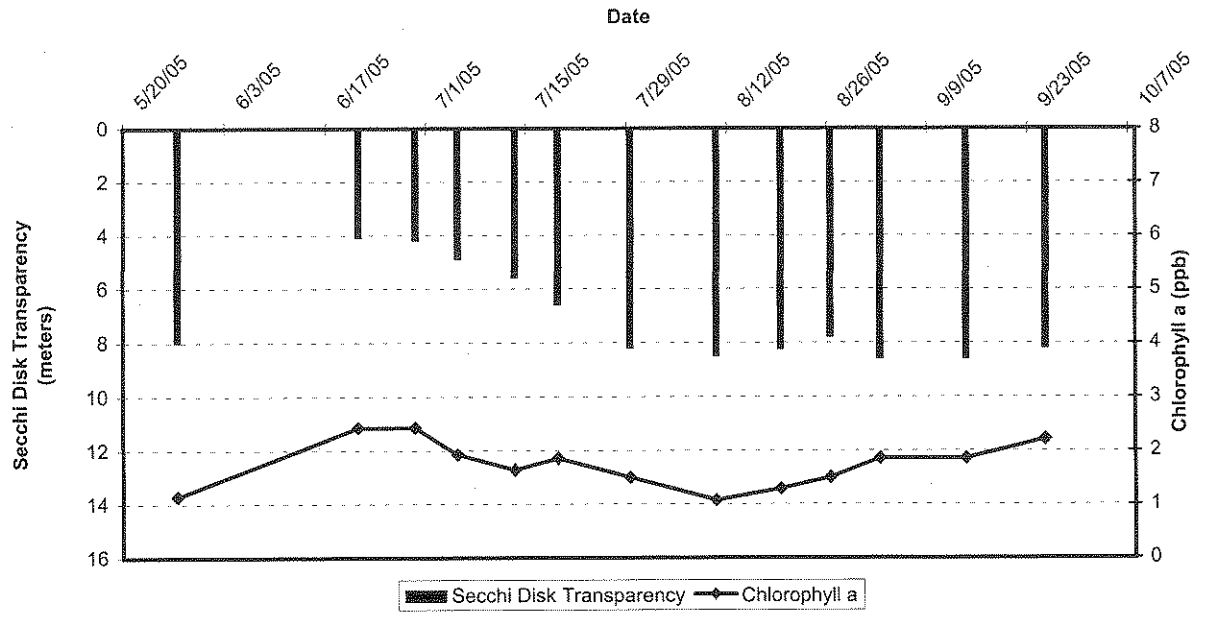


Figure 12. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll α trends for Site 3 Pasquaney Bay. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the chlorophyll α data are reported to the nearest 0.1 parts per billion (ppb).

Figure 13. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 3 Pasquaney Bay. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the dissolved color data are reported to the nearest 0.1 chloroplatinate unit (CPU).

Note: the overlay of the Secchi Disk data with chlorophyll α and dissolved color data is intended to provide a visual depiction of the impacts of chlorophyll α and dissolved color on water transparency measurements (e.g. higher chlorophyll α and dissolved color concentrations often correspond to shallower water transparencies).

Newfound -- Site 3 Pasquaney (2005 Seasonal Data)



Newfound -- Site 3 Pasquaney (2005 Seasonal Data)

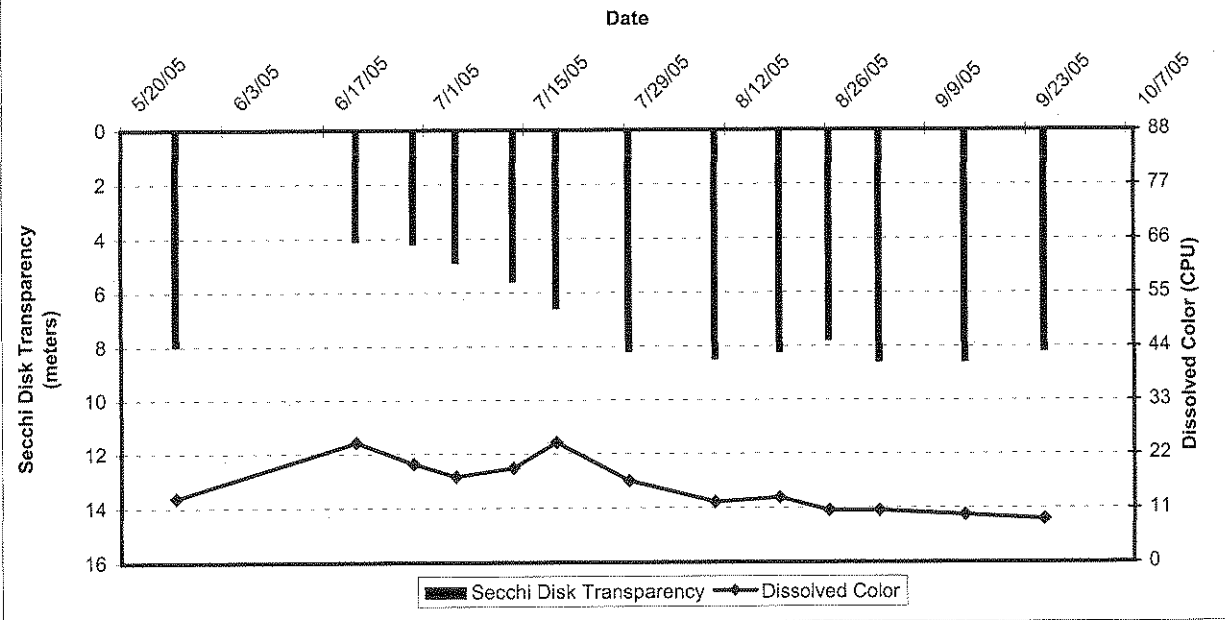
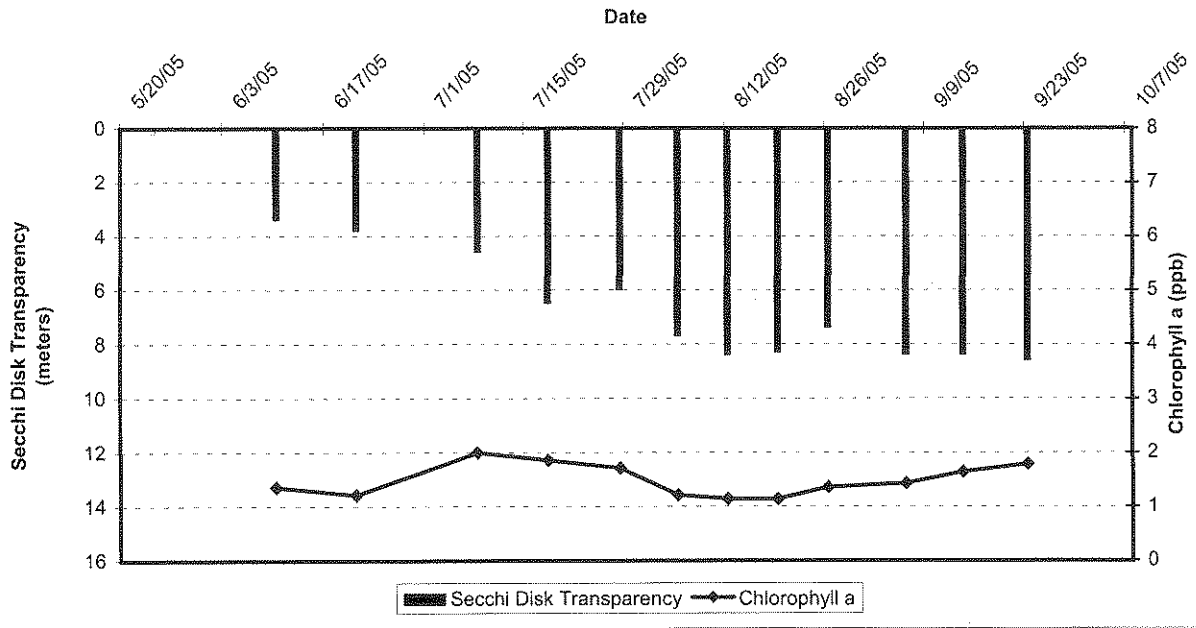


Figure 14. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll a trends for Site 4 Loon Island. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the chlorophyll a data are reported to the nearest 0.1 parts per billion (ppb).

Figure 15. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 4 Loon Island. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the dissolved color data are reported to the nearest 0.1 chloro-platinate unit (CPU).

Note: the overlay of the Secchi Disk data with chlorophyll a and dissolved color data is intended to provide a visual depiction of the impacts of chlorophyll a and dissolved color on water transparency measurements (e.g. higher chlorophyll a and dissolved color concentrations often correspond to shallower water transparencies).

Newfound -- Site 4 Loon Island (2005 Seasonal Data)



Newfound -- Site 4 Loon Island (2005 Seasonal Data)

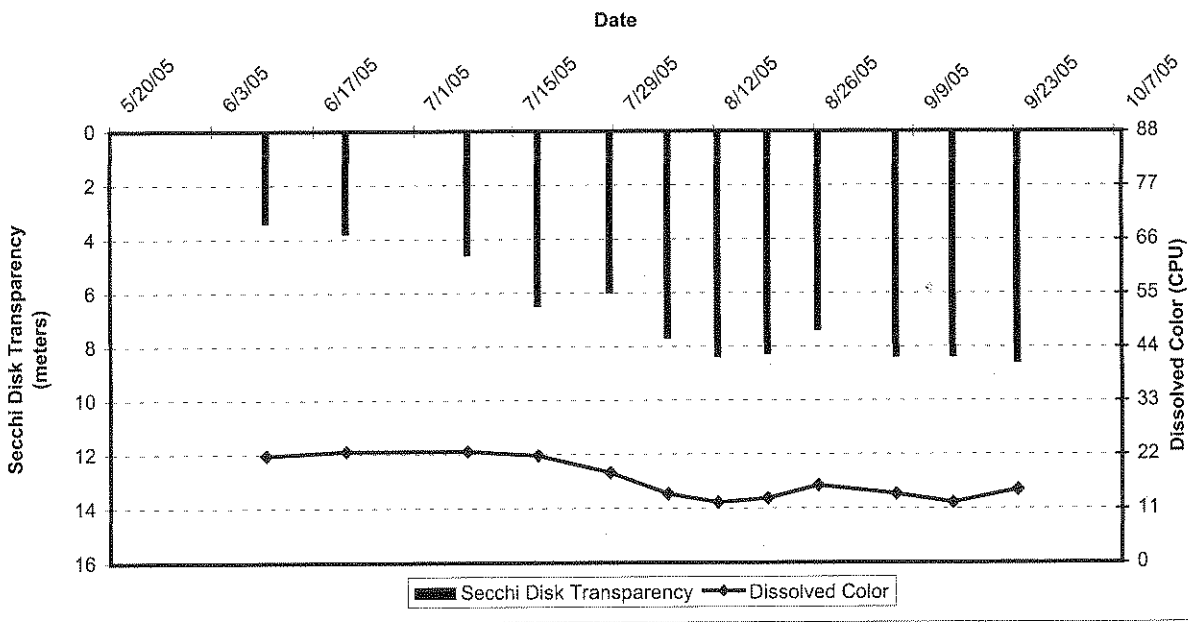
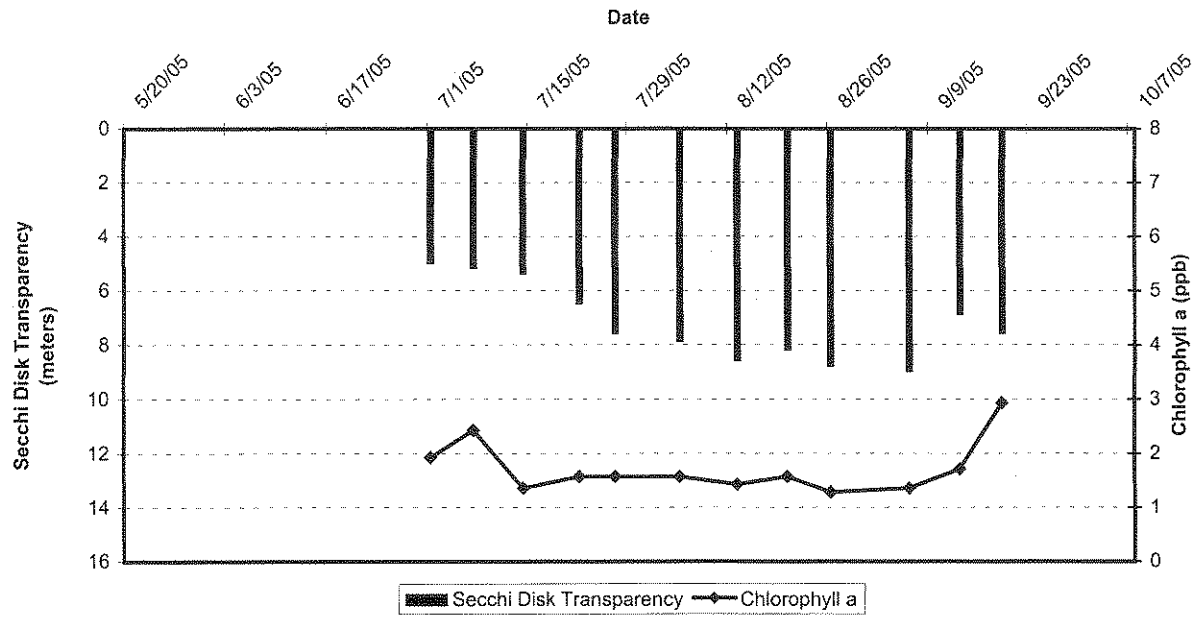


Figure 16. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll a trends for Site 5 Beachwood. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the chlorophyll a data are reported to the nearest 0.1 parts per billion (ppb).

Figure 17. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 5 Beachwood. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the dissolved color data are reported to the nearest 0.1 chloro-platinate unit (CPU).

Note: the overlay of the Secchi Disk data with chlorophyll a and dissolved color data is intended to provide a visual depiction of the impacts of chlorophyll a and dissolved color on water transparency measurements (e.g. higher chlorophyll a and dissolved color concentrations often correspond to shallower water transparencies).

Newfound -- Site 5 Beachwood (2005 Seasonal Data)



Newfound -- Site 5 Beachwood (2005 Seasonal Data)

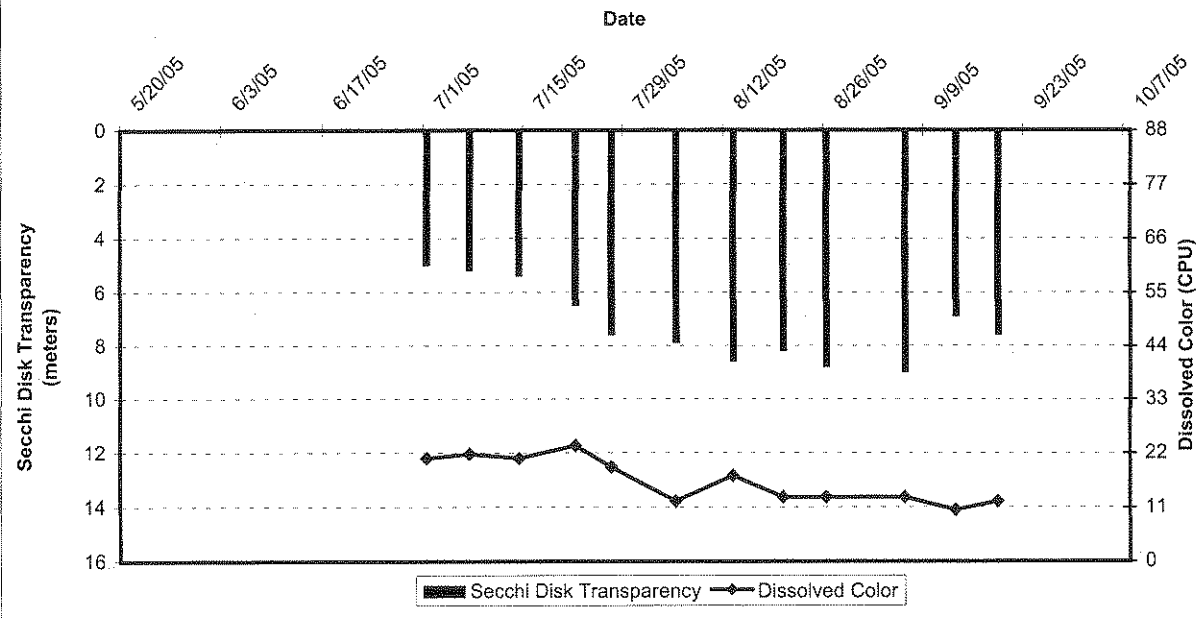
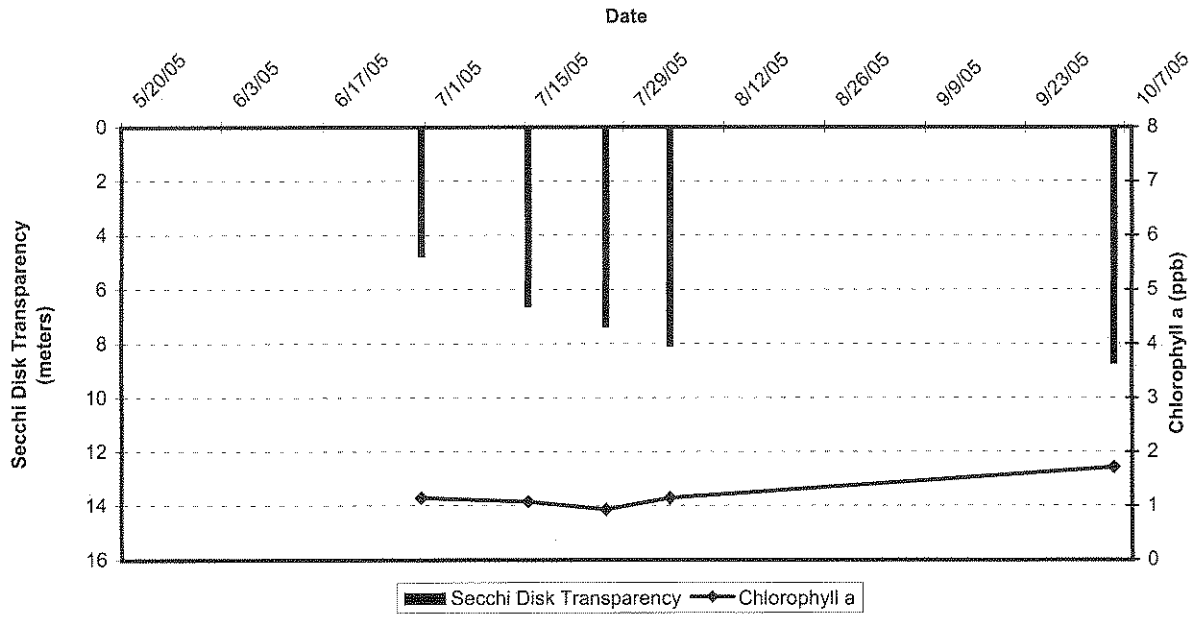


Figure 18. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll a trends for Site 7 Cockermouth River. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the chlorophyll a data are reported to the nearest 0.1 parts per billion (ppb).

Figure 19. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 7 Cockermouth River. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the dissolved color data are reported to the nearest 0.1 chloroplatinate unit (CPU).

Note: the overlay of the Secchi Disk data with chlorophyll a and dissolved color data is intended to provide a visual depiction of the impacts of chlorophyll a and dissolved color on water transparency measurements (e.g. higher chlorophyll a and dissolved color concentrations often correspond to shallower water transparencies).

Newfound -- Site 7 Cockermouth (2005 Seasonal Data)



Newfound -- Site 7 Cockermouth (2005 Seasonal Data)

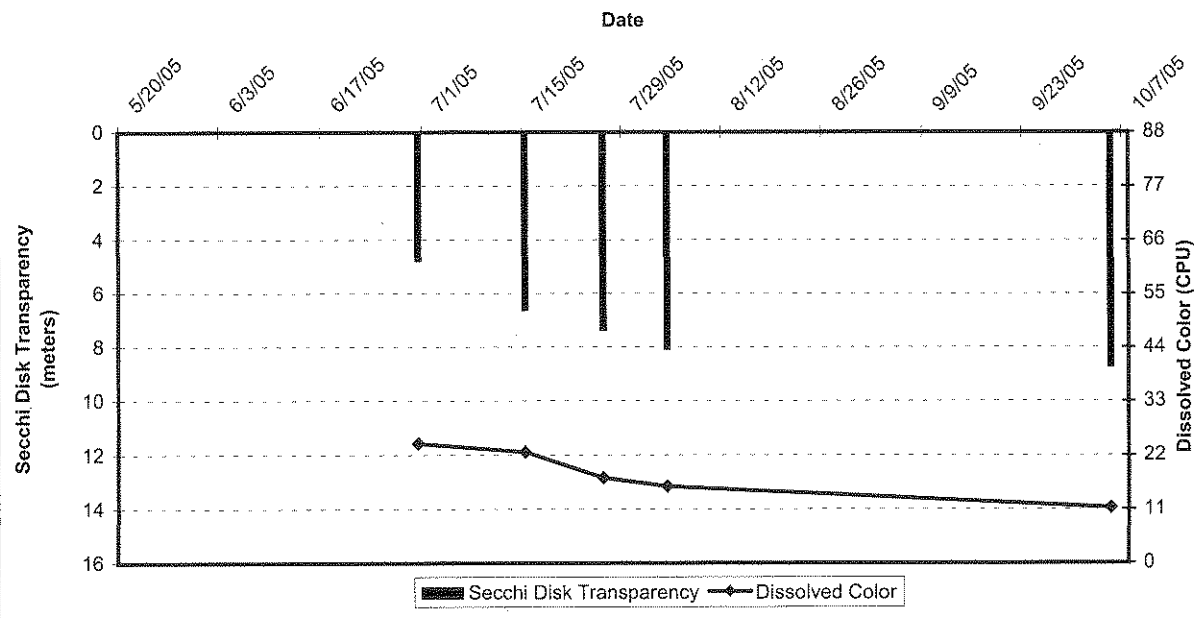
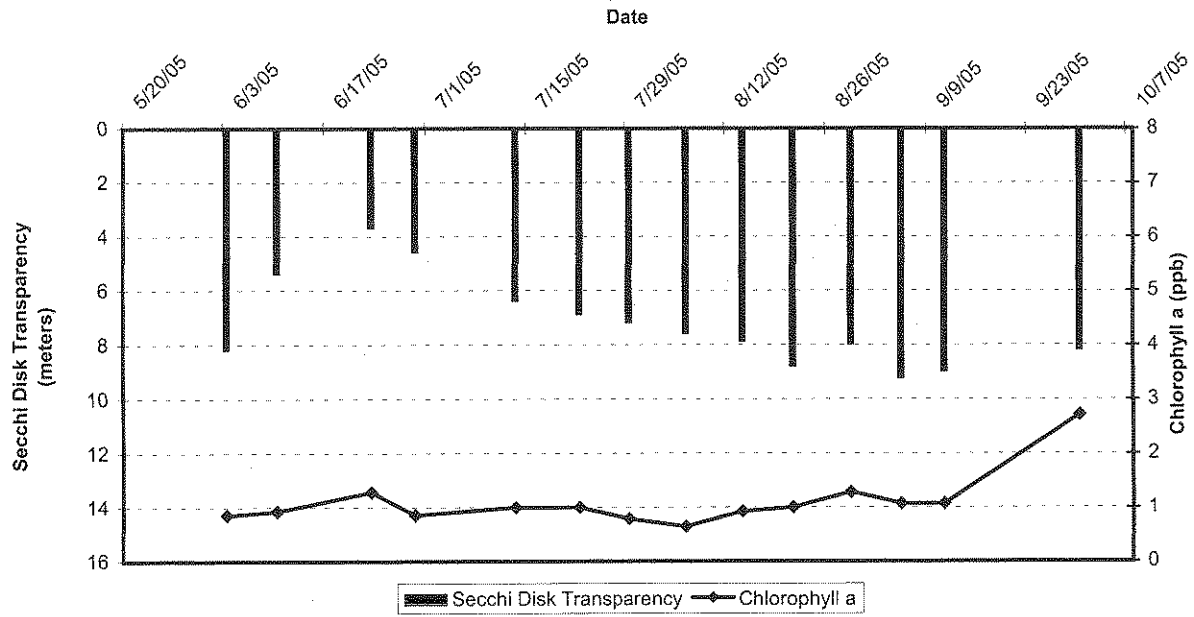


Figure 20. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and chlorophyll *a* trends for Site 8 Follansbee Cove. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the chlorophyll *a* data are reported to the nearest 0.1 parts per billion (ppb).

Figure 21. Newfound Lake, 2005. Seasonal Secchi Disk (water transparency) and dissolved color trends for Site 8 Follansbee Cove. The Secchi Disk transparency data are reported to the nearest 0.1 meters while the dissolved color data are reported to the nearest 0.1 chloroplatinate unit (CPU).

Note: the overlay of the Secchi Disk data with chlorophyll *a* and dissolved color data is intended to provide a visual depiction of the impacts of chlorophyll *a* and dissolved color on water transparency measurements (e.g. higher chlorophyll *a* and dissolved color concentrations often correspond to shallower water transparencies).

Newfound -- Site 8 Follansbee (2005 Seasonal Data)



Newfound -- Site 8 Follansbee (2005 Seasonal Data)

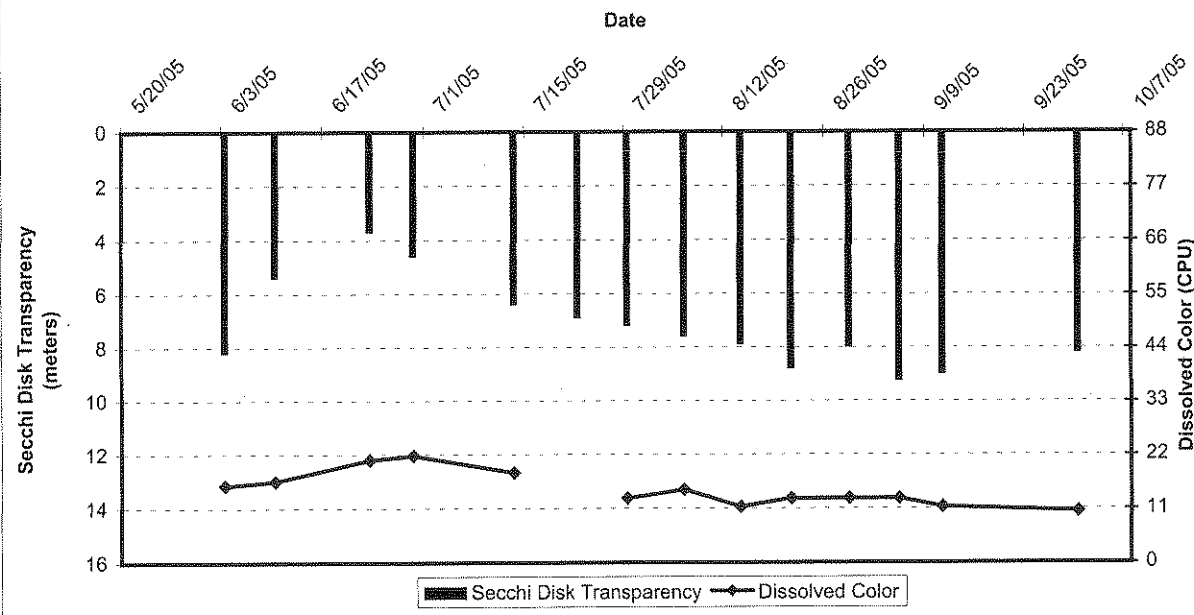
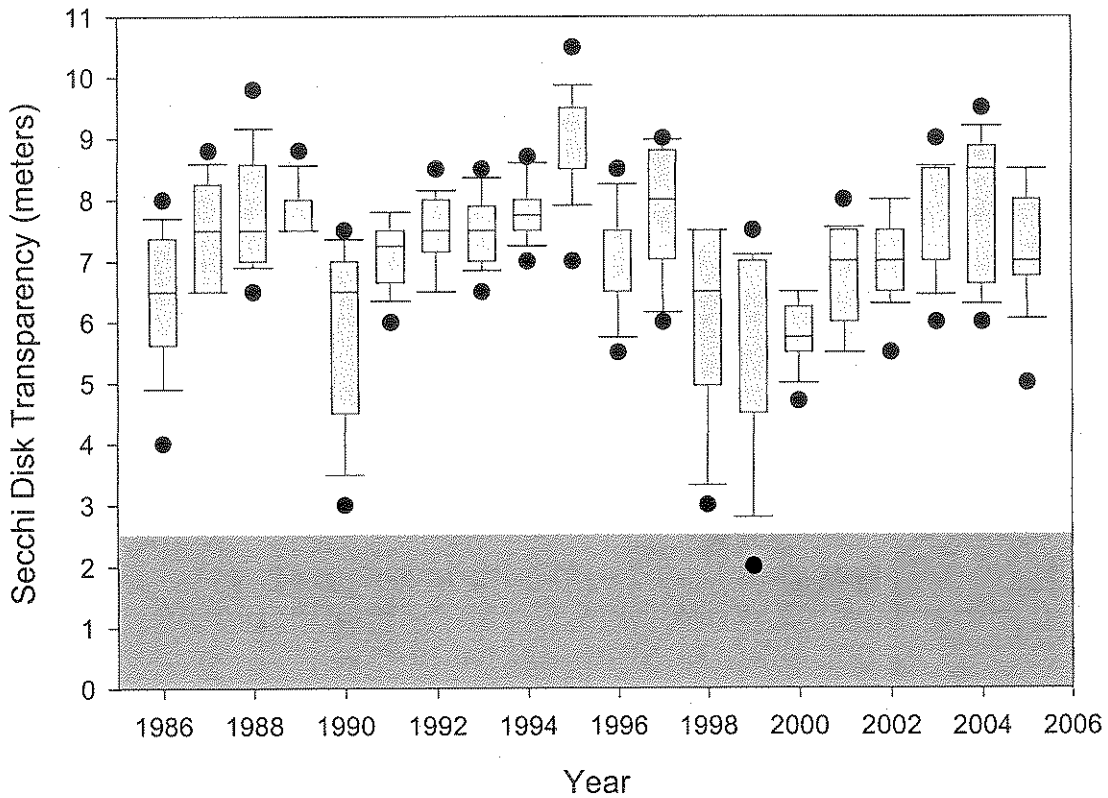


Figure 22. Comparison of the annual Newfound Lake, Site 2 Mayhew, lay monitor Secchi Disk transparency data (1986-2005) that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded regions on the graph are representative of water transparency conditions considered typical of an unproductive (clear), a moderately productive (light gray shading) and a highly productive (dark gray shading) lake.

Figure 23. Comparison of the annual Newfound Lake, Site 2 Mayhew, lay monitor chlorophyll α data (1986-2005) that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded region on the graph is representative of conditions considered typical of a moderately productive lake while the clear region of the graph represents the range considered typical of an unproductive lake.

Newfound Lake -- Site 2 Mayhew
Annual Secchi Disk Transparency Comparisons
Box and Whisker Plots: 1986-2005



Newfound Lake -- Site 2 Mayhew
Annual Chlorophyll a Comparisons
Box and Whisker Plots: 1986-2005

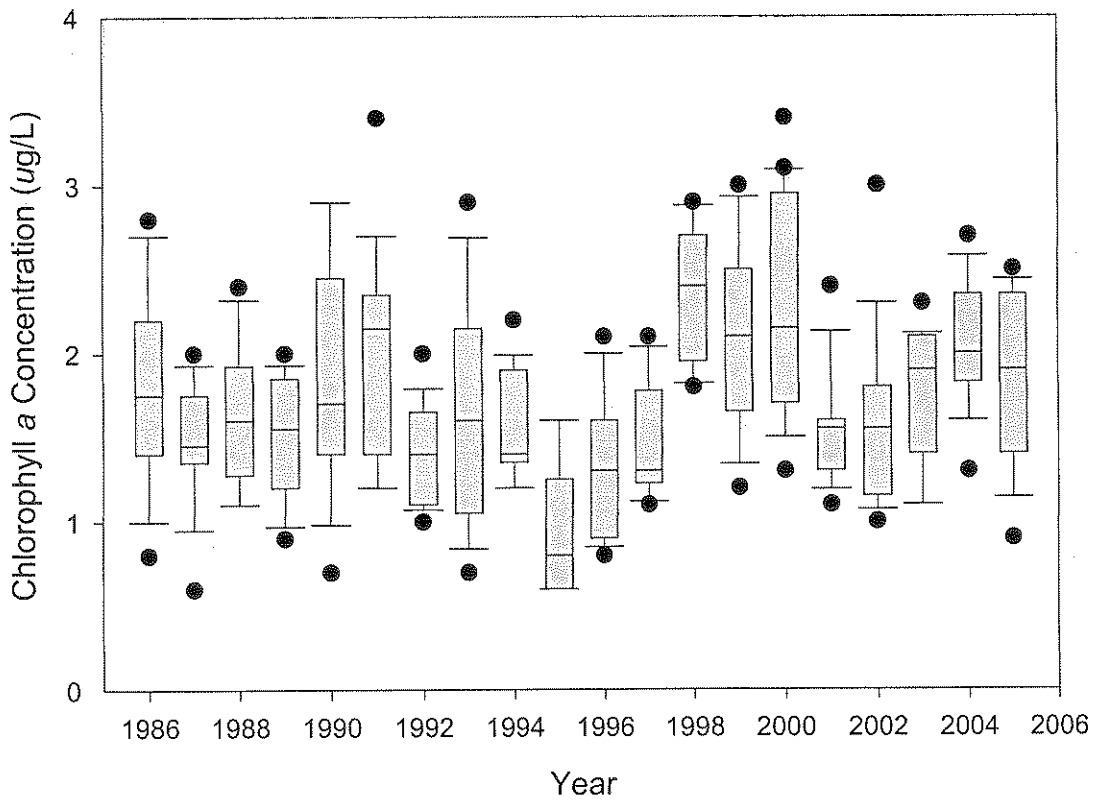
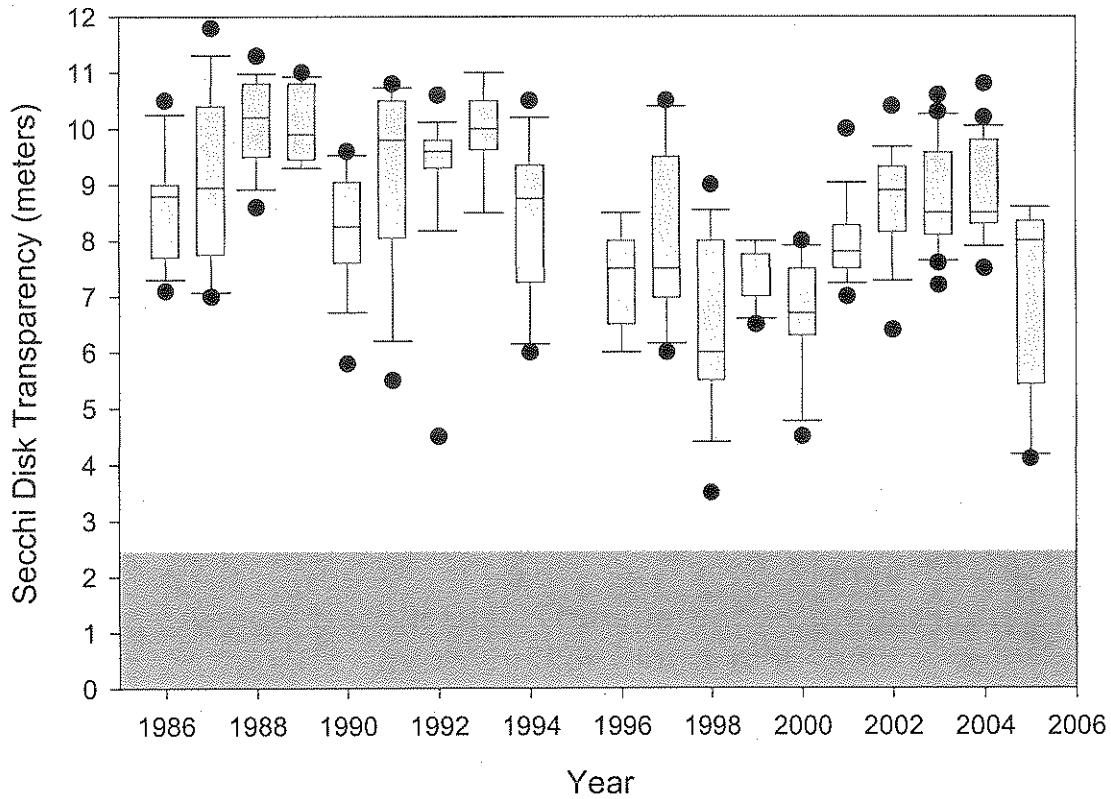


Figure 24. Comparison of the annual Newfound Lake, Site 3 Pasquaney, lay monitor Secchi Disk transparency data (1986-2005) that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded regions on the graph are representative of water transparency conditions considered typical of an unproductive (clear), a moderately productive (light gray shading) and a highly productive (dark gray shading) lake.

Figure 25. Comparison of the annual Newfound Lake, Site 3 Pasquaney, lay monitor chlorophyll *a* data (1986-2005) that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded region on the graph is representative of conditions considered typical of a moderately productive lake while the clear region of the graph represents the range considered typical of an unproductive lake.

**Newfound Lake - Site 3 Pasquaney
Annual Secchi Disk Transparency Comparisons
Box and Whisker Plots: 1986-2005**



**Newfound Lake -- Site 3 Pasquaney
Annual Chlorophyll a Comparisons
Box and Whisker Plots: 1986-2005**

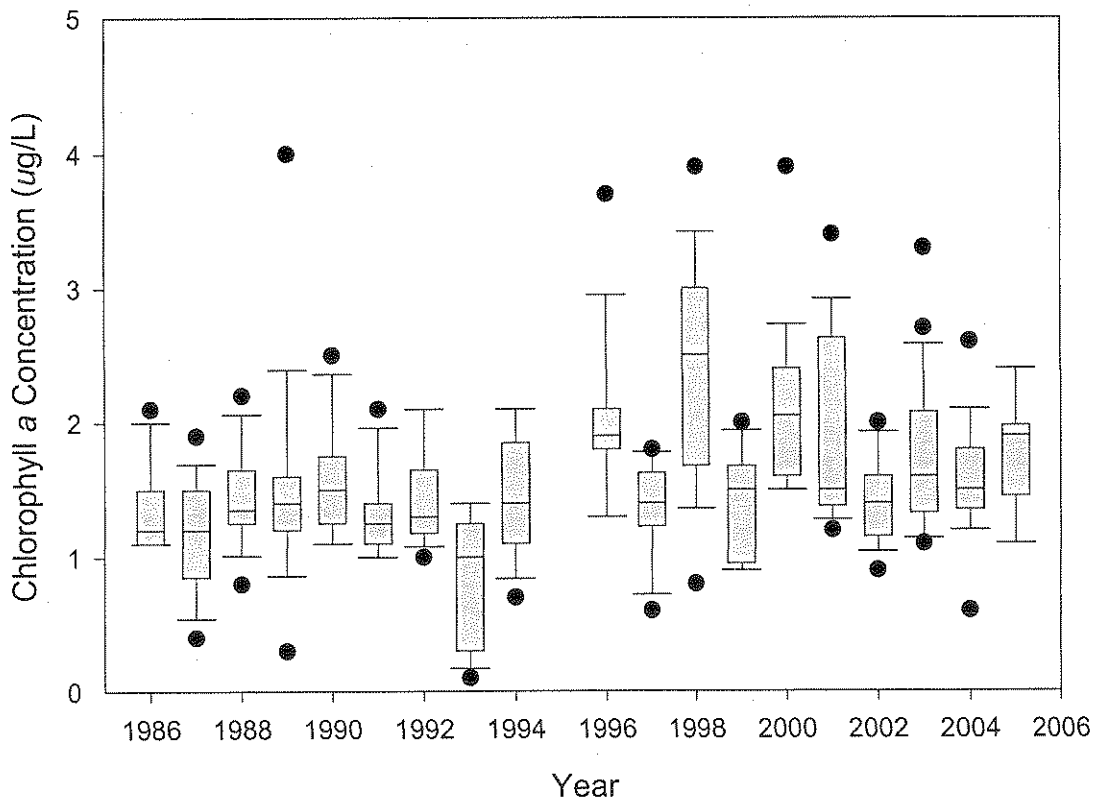
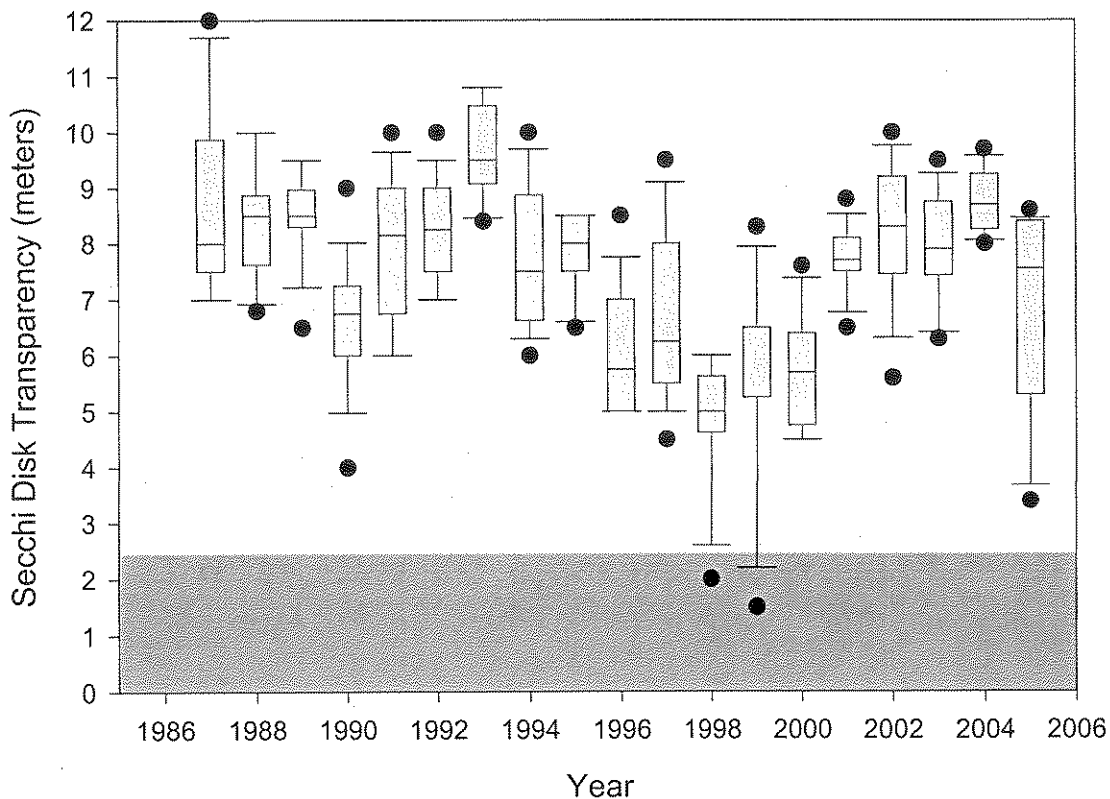


Figure 26. Comparison of the annual Newfound Lake, Site 4 Loon Island, lay monitor Secchi Disk transparency data (1987-2005) that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded regions on the graph are representative of water transparency conditions considered typical of an unproductive (clear), a moderately productive (light gray shading) and a highly productive (dark gray shading) lake.

Figure 27. Comparison of the annual Newfound Lake, Site 4 Loon Island, lay monitor chlorophyll *a* data (1987-2005) that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded region on the graph is representative of conditions considered typical of a moderately productive lake while the clear region of the graph represents the range considered typical of an unproductive lake.

**Newfound Lake - Site 4 Loon Island
Annual Secchi Disk Transparency Comparisons
Box and Whisker Plots: 1987-2005**



**Newfound Lake -- Site 4 Loon Island
Annual Chlorophyll a Comparisons
Box and Whisker Plots: 1987-2005**

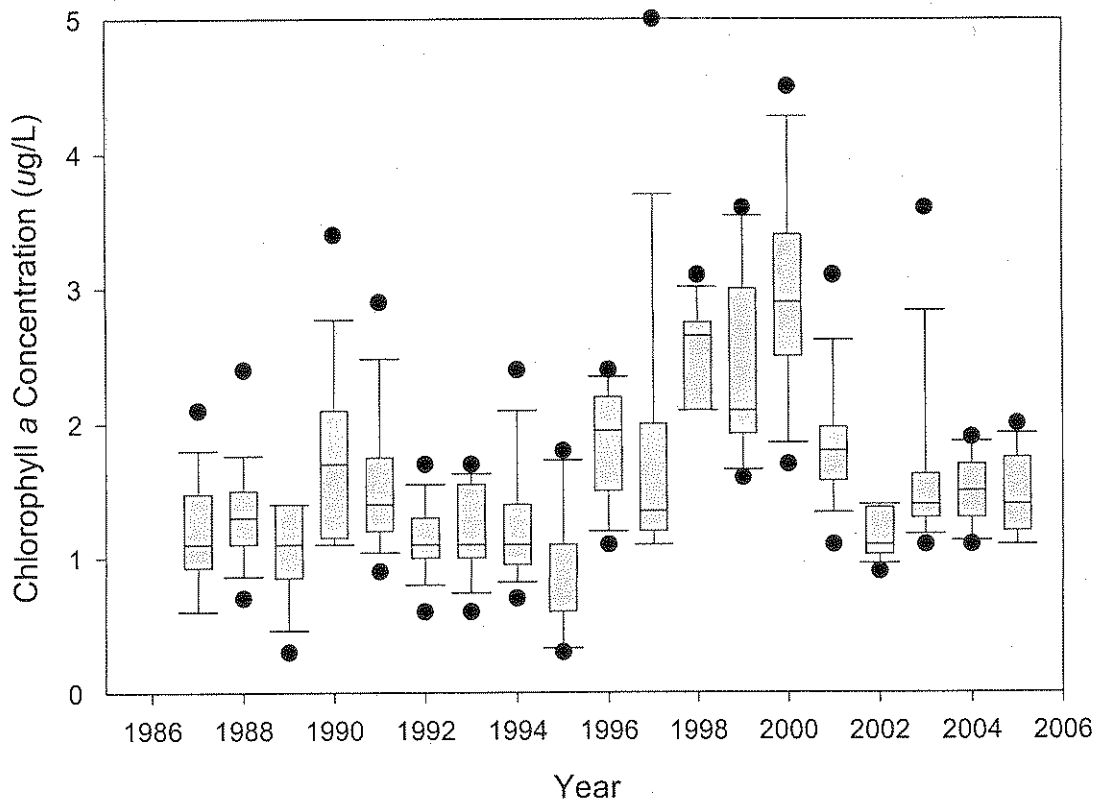
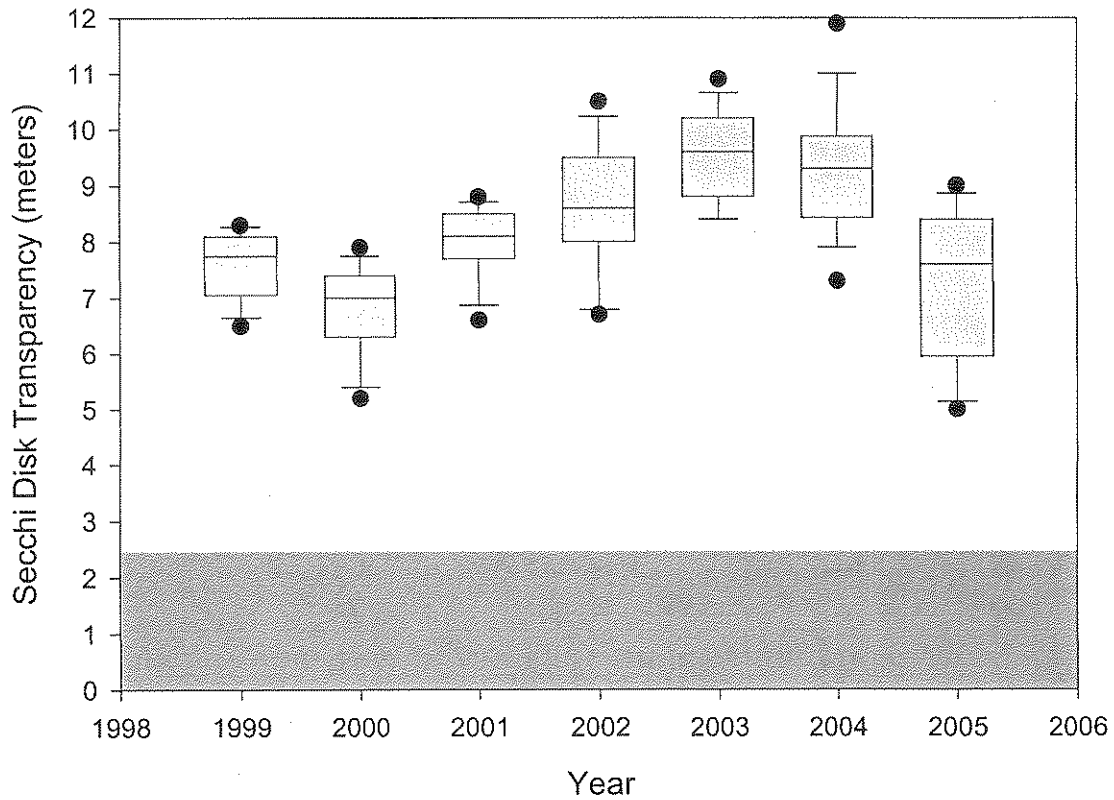


Figure 28. Comparison of the annual Newfound Lake, Site 5 Beachwood, lay monitor Secchi Disk transparency data (1999-2005) that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded regions on the graph are representative of water transparency conditions considered typical of an unproductive (clear), a moderately productive (light gray shading) and a highly productive (dark gray shading) lake.

Figure 29. Comparison of the annual Newfound Lake, Site 5 Beachwood, lay monitor chlorophyll *a* data (1999-2005) that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded region on the graph is representative of conditions considered typical of a moderately productive lake while the clear region of the graph represents the range considered typical of an unproductive lake.

**Newfound Lake - Site 5 Beachwood
Annual Secchi Disk Transparency Comparisons
Box and Whisker Plots: 1999-2005**



**Newfound Lake -- Site 5 Beachwood
Annual Chlorophyll a Comparisons
Box and Whisker Plots: 1999-2005**

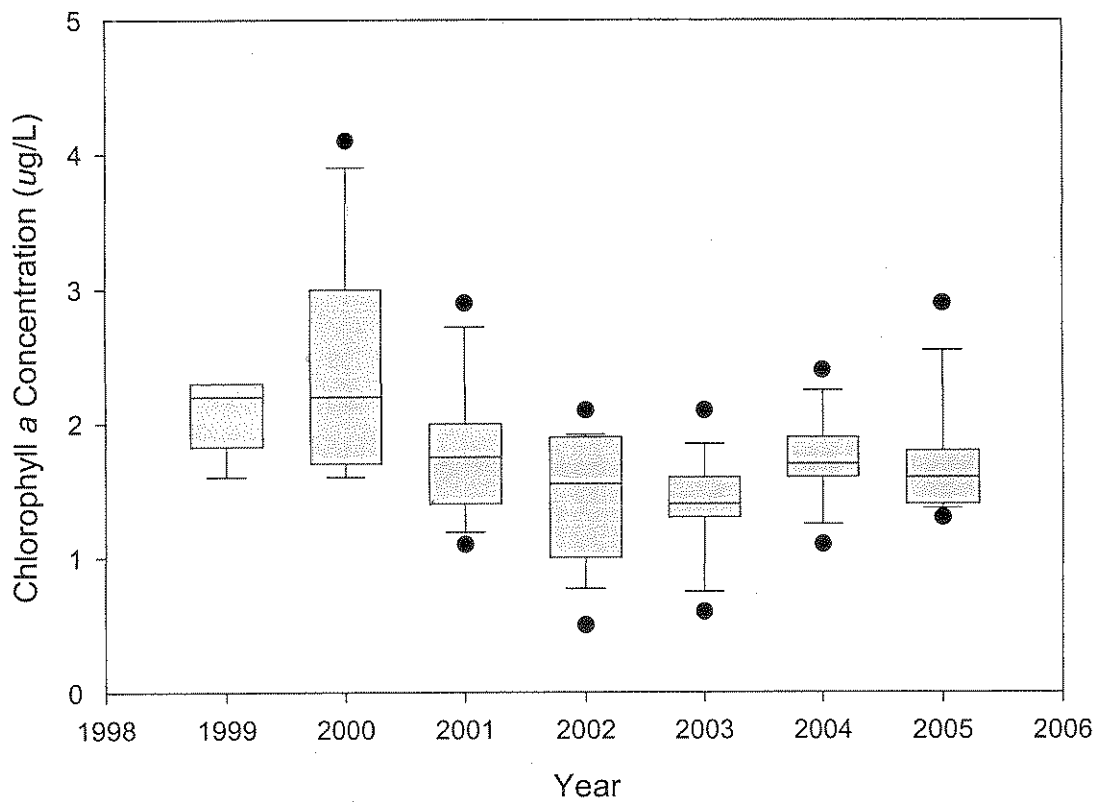
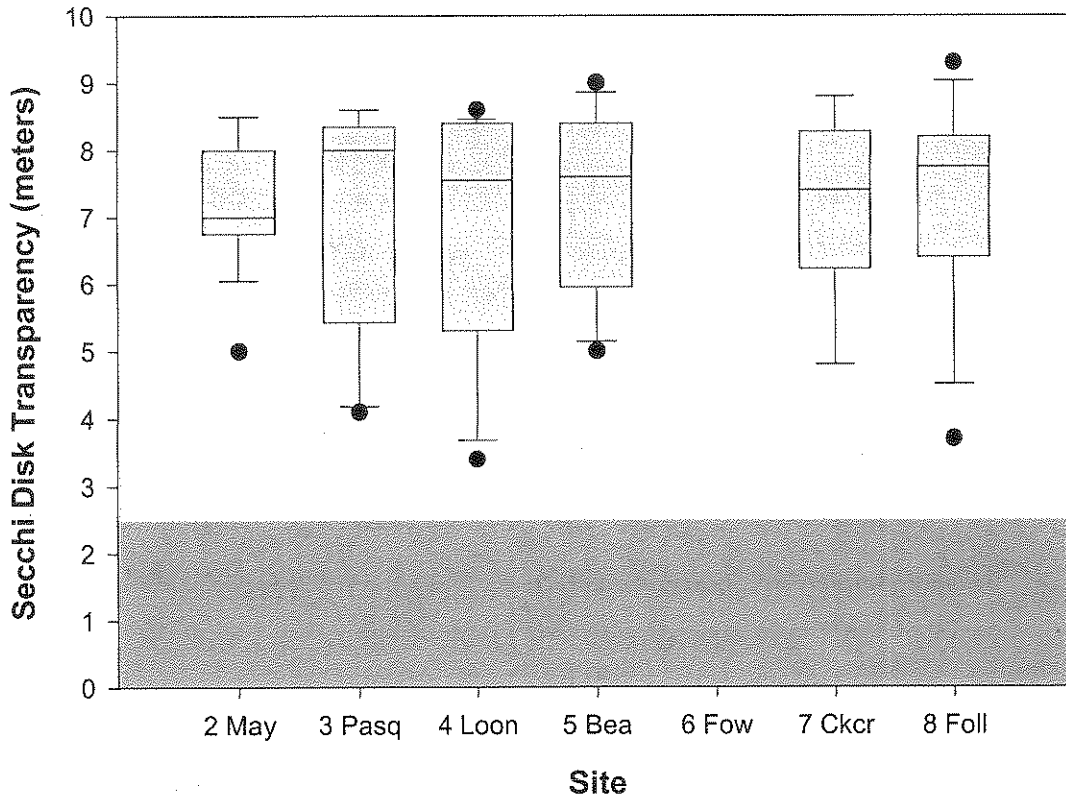


Figure 30. Newfound Lake inter-site comparison of the 2005 lay monitor Secchi Disk transparency data that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded regions on the graph are representative of water transparency conditions considered typical of an unproductive (clear), a moderately productive (light gray shading) and a highly productive (dark gray shading) lake.

Figure 31. Newfound Lake inter-site comparison of the 2005 lay monitor Chlorophyll a data that are presented as box and whisker plots. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. The gray shaded region on the graph is representative of conditions considered typical of a moderately productive lake while the clear region of the graph represents the range considered typical of an unproductive lake.

Newfound Lake - Inter-Site Comparison
 2005 Secchi Disk Transparency Comparisons
 Box and Whisker Plots



Newfound Lake - Inter-Site Comparison
 2005 Chlorophyll a Comparisons
 Box and Whisker Plots

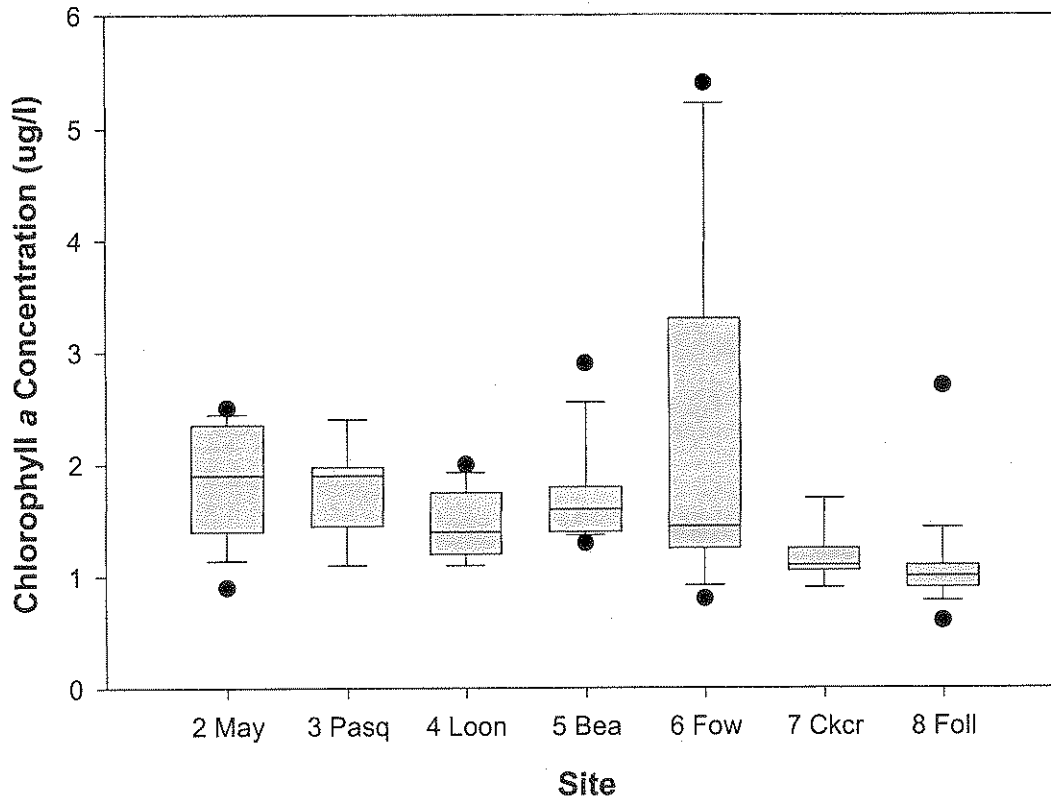
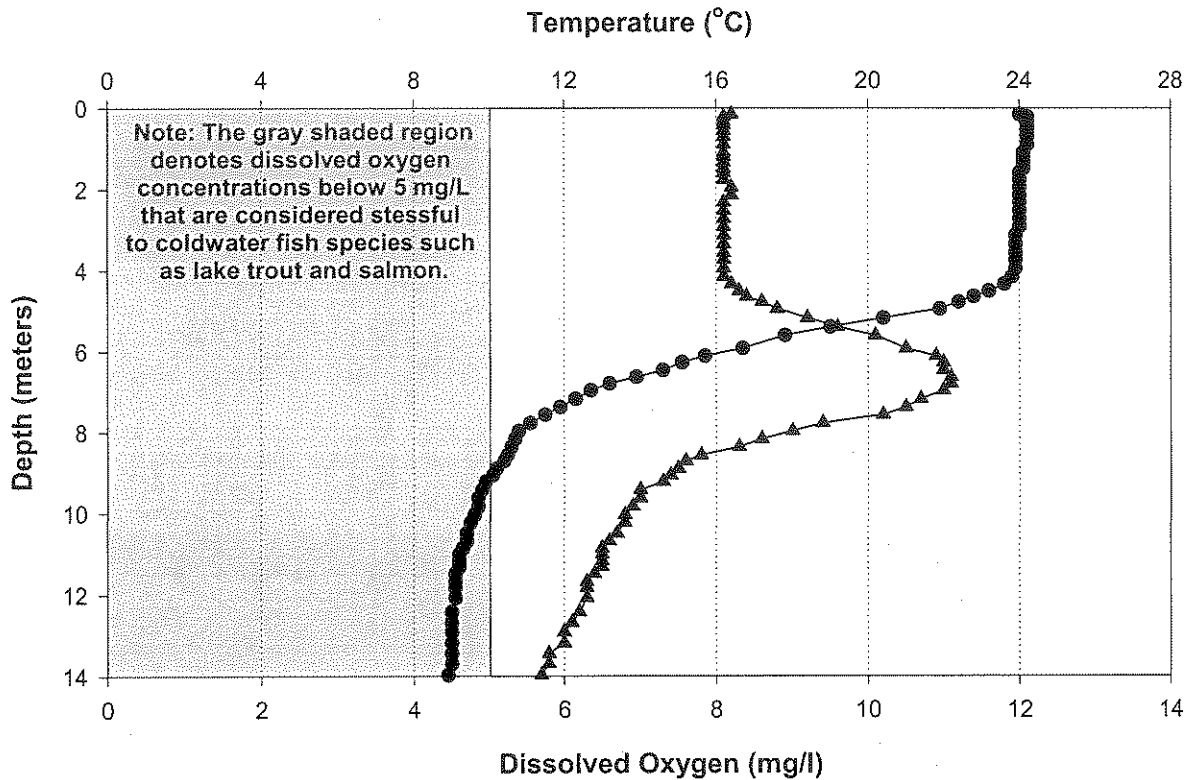


Figure 32. Temperature and dissolved oxygen profiles collected in Newfound Lake, (A) Site 2 Mayhew and (B) Site 3 Pasquaney on August 2, 2005. The gray shaded region on the graph denotes dissolved oxygen concentrations stressful to coldwater fish. The temperature and dissolved oxygen data were collected at increments of no greater than one-half meter and are reported as degrees Celsius ($^{\circ}\text{C}$) and parts per million (ppm), respectively.

Newfound - Site 2 Mayhew

August 2, 2005



Newfound - Site 3 Pasquaney Bay

August 2, 2005

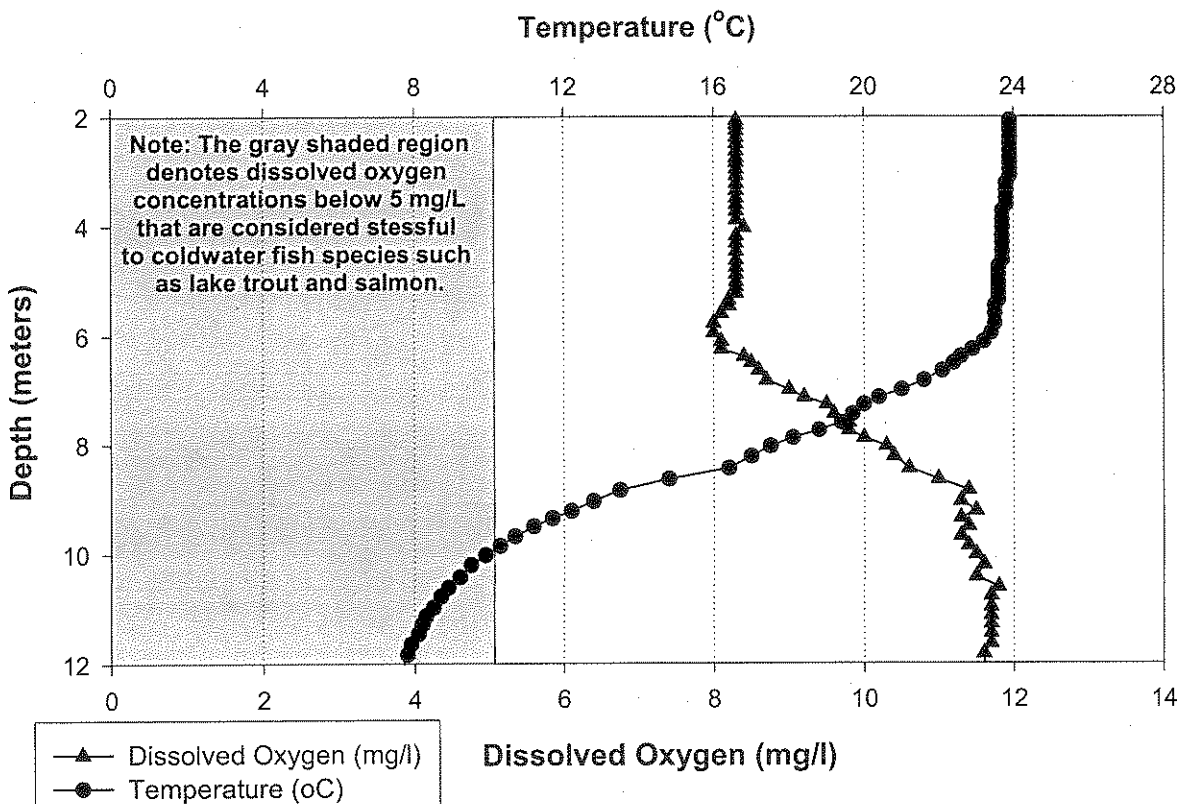
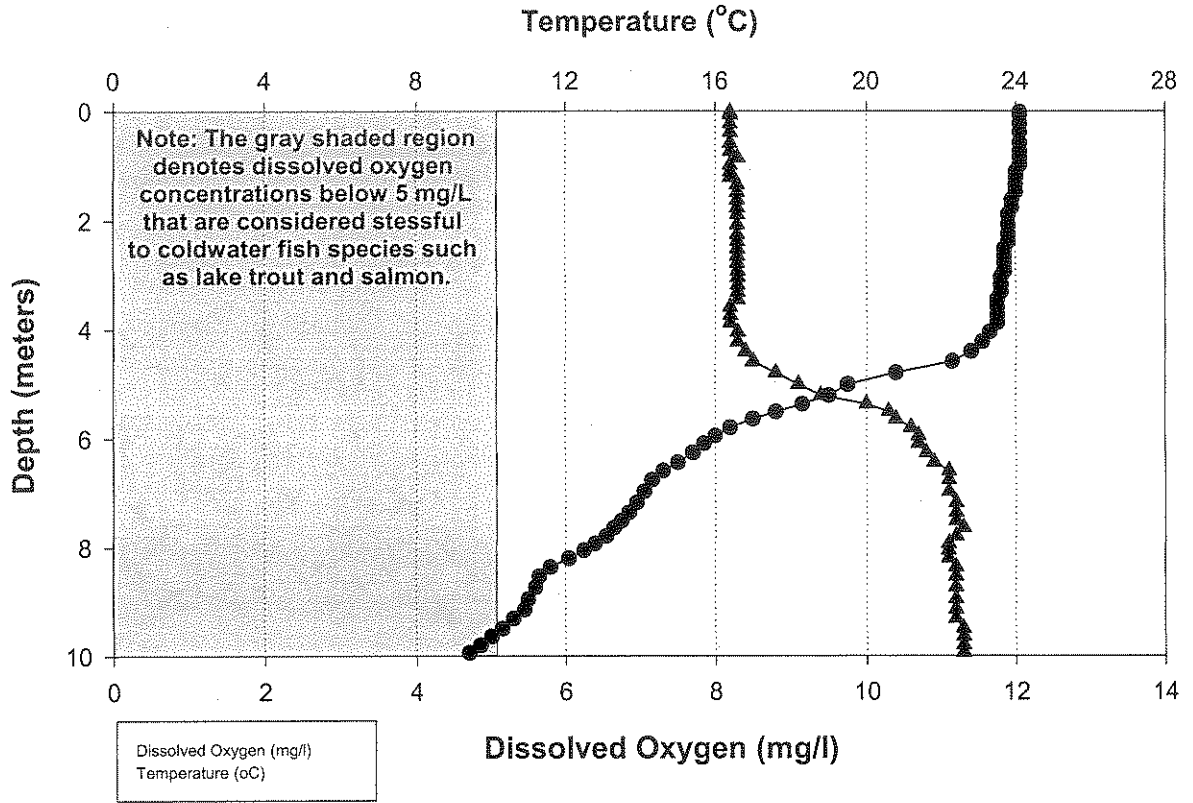


Figure 33. Temperature and dissolved oxygen profiles collected in Newfound Lake, (A) Site 4 Loon Island and (B) Site 5 Beachwood on August 2, 2005. The gray shaded region on the graph denotes dissolved oxygen concentrations stressful to coldwater fish. The temperature and dissolved oxygen data were collected at increments of no greater than one-half meter and are reported as degrees Celsius ($^{\circ}\text{C}$) and parts per million (ppm), respectively.

Newfound - Site 4 Loon Isl

August 2, 2005



Newfound - Site 5 Beachwood

August 2, 2005

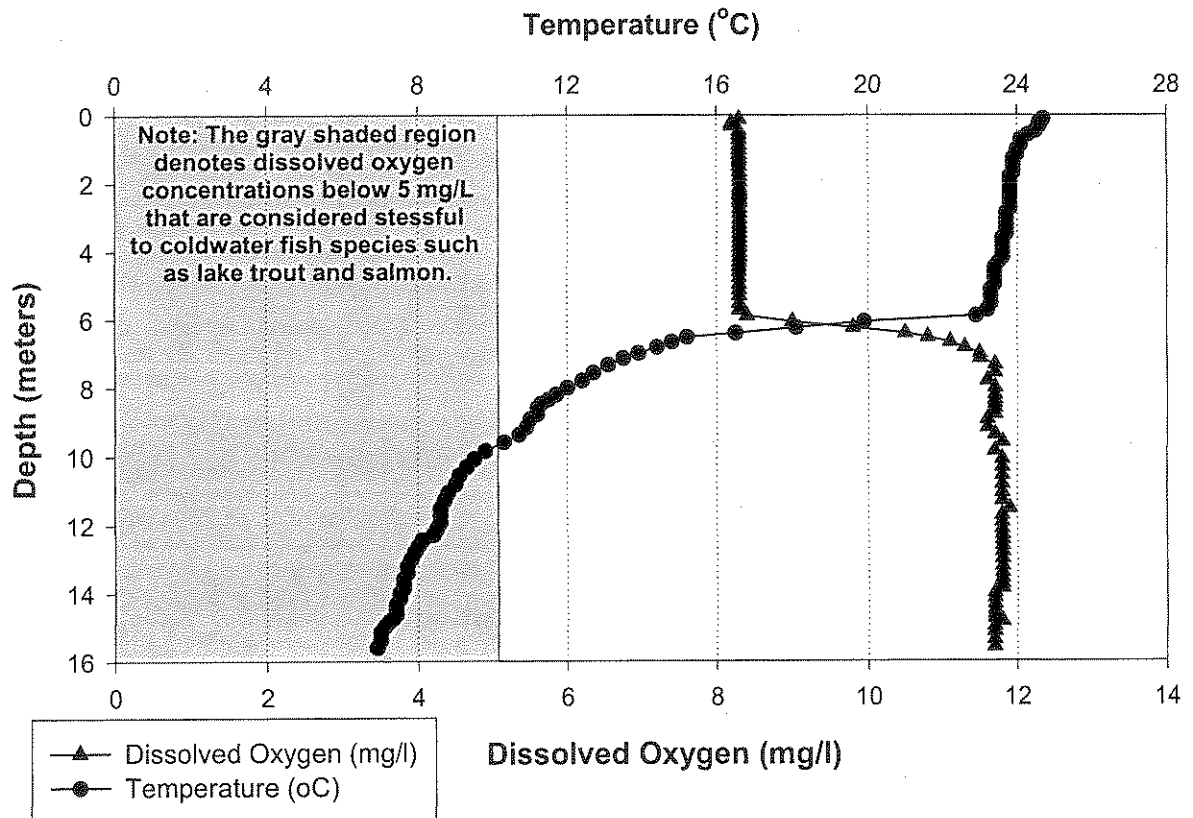
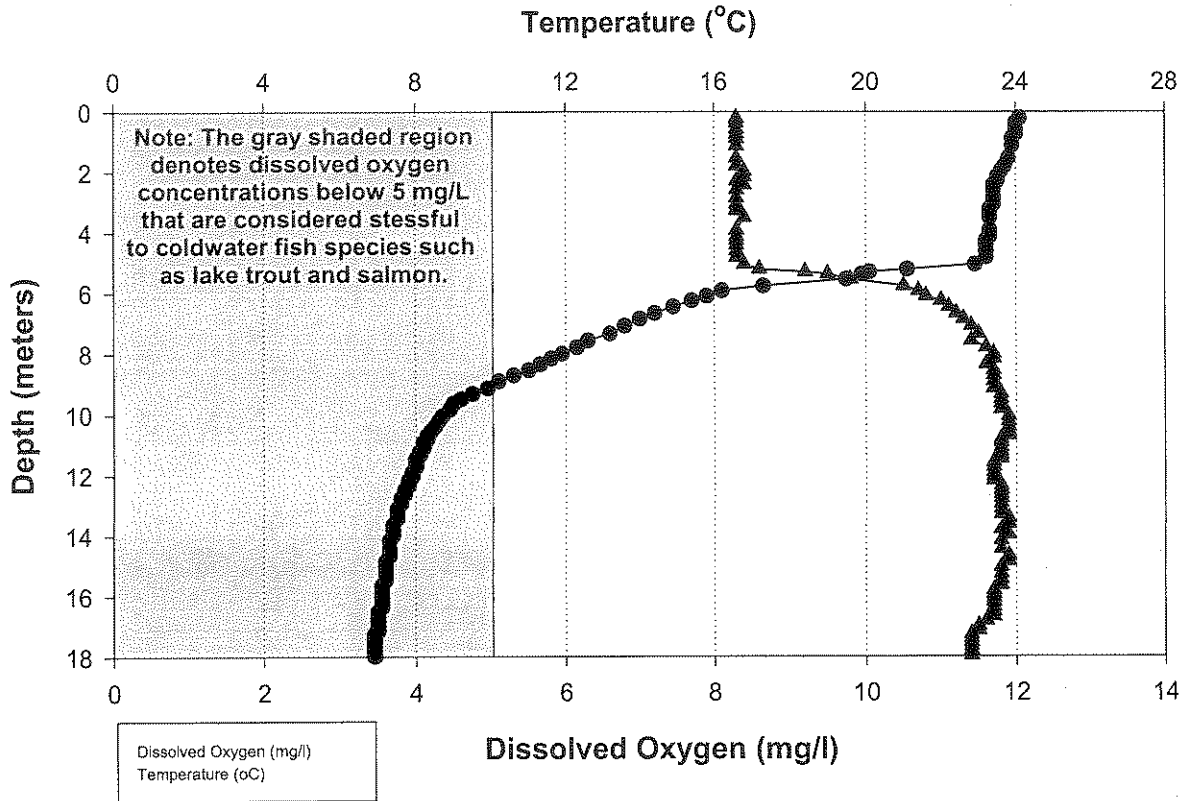
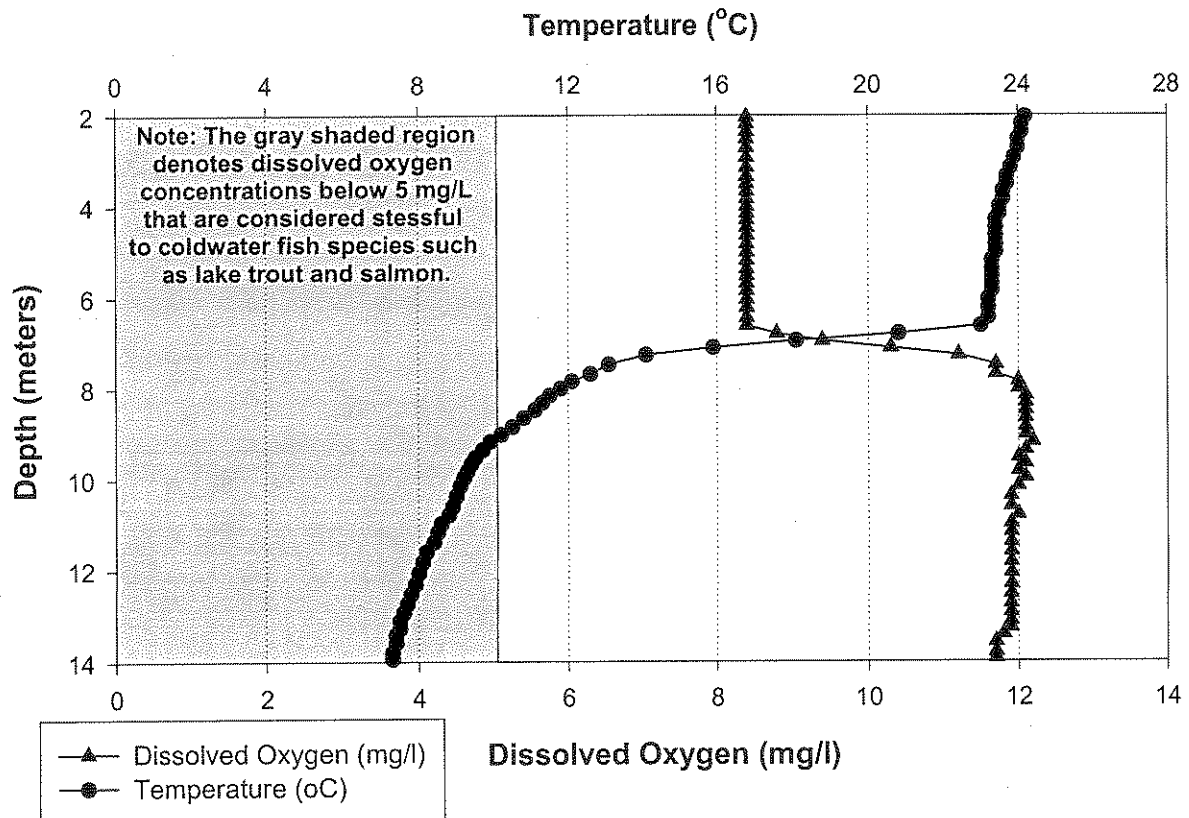


Figure 34. Temperature and dissolved profiles collected in Newfound Lake, (A) Site 7 Cockermouth River and (B) Site 8 Follansbee Cove on August 2, 2005. The gray shaded region on the graph denotes dissolved oxygen concentrations stressful to coldwater fish. The temperature and dissolved oxygen data were collected at increments of no greater than one-half meter and are reported as degrees Celsius ($^{\circ}\text{C}$) and parts per million (ppm), respectively.

**Newfound - Site 7 Cockermouth
August 2, 2005**



**Newfound - Site 8 Follansbee Cove
August 2, 2005**



APPENDIX A

Lakes Lay Monitoring Program, U.N.H. [Lay Monitor Data]

Newfound Lake, Bristol, Hebron, Alexandria and Bridgewater NH
-- subset of trophic indicators, all sites, 2005

Average transparency:	7.1 (2005: 68 values;	3.4 - 9.3 range)
Average chlorophyll:	1.6 (2005: 75 values;	0.6 - 5.4 range)
Average Color:	15.8 (2005: 73 values;	8.7 - 30.4 range)
Average Alkalinity (gray):	2.8 (2005: 67 values;	2.0 - 4.5 range)
Average alkalinity (pink):	3.5 (2005: 67 values;	2.6 - 5.6 range)
Total Phosphorus (streams)	7.7 (2005: 7 values;	3.2 - 15.1 range)

Site	Date	Secchi Disk Transparency (meters)	Chl <i>a</i> (ug/l)	Dissolved Color (cpu)	Total Phosphorus (ug/l)	Alkalinity gray end pt. @ pH 5.1 (mg/l)	Alkalinity pink end pt. @ pH 4.6 (mg/l)
2 Mayhew	7/5/05	5.0	2.4	16.5	-----	3.2	3.7
2 Mayhew	7/11/05	8.0	2.4	23.5	-----	3.2	3.7
2 Mayhew	7/19/05	8.5	2.0	19.1	-----	3.4	3.9
2 Mayhew	7/26/05	7.0	1.4	17.4	-----	2.7	3.1
2 Mayhew	8/8/05	7.0	0.9	13.0	-----	2.9	3.4
2 Mayhew	8/17/05	6.5	1.3	12.2	-----	2.8	3.3
2 Mayhew	8/24/05	7.0	1.4	12.2	-----	2.8	3.3
2 Mayhew	9/2/05	7.0	1.6	12.2	-----	2.9	3.4
2 Mayhew	9/7/05	7.5	1.9	13.0	-----	2.8	3.3
2 Mayhew	9/19/05	8.0	2.5	13.0	-----	2.7	3.2
2 Mayhew	10/3/05	6.5	2.2	10.4	-----	3.0	3.5
3 Pasquan	5/27/05	8.0	1.1	13.0	-----	-----	-----
3 Pasquan	6/21/05	4.1	2.4	24.3	-----	2.3	2.6
3 Pasquan	6/29/05	4.2	2.4	20.0	-----	2.7	3.3
3 Pasquan	7/5/05	4.9	1.9	17.4	-----	2.6	3.1
3 Pasquan	7/13/05	5.6	1.6	19.1	-----	2.7	3.1
3 Pasquan	7/19/05	6.6	1.9	24.3	-----	2.5	3.2
3 Pasquan	7/29/05	8.2	1.5	16.5	-----	2.3	2.9
3 Pasquan	8/10/05	8.5	1.1	12.2	-----	2.9	3.5
3 Pasquan	8/19/05	8.3	1.3	13.0	-----	2.9	3.5
3 Pasquan	8/26/05	7.8	1.5	10.4	-----	3.0	3.6
3 Pasquan	9/2/05	8.6	1.9	10.4	-----	2.9	3.4
3 Pasquan	9/14/05	8.6	1.9	9.6	-----	3.1	3.9
3 Pasquan	9/25/05	8.2	2.2	8.7	-----	3.4	3.9
4 LoonIsl	6/10/05	3.4	1.4	21.7	-----	2.5	3.3
4 LoonIsl	6/21/05	3.8	1.2	22.6	-----	2.3	2.8
4 LoonIsl	7/8/05	4.6	2.0	22.6	-----	2.2	2.8
4 LoonIsl	7/18/05	6.5	1.9	21.7	-----	2.0	2.6
4 LoonIsl	7/28/05	6.0	1.7	18.2	-----	2.2	2.7

Site	Date	Secchi Disk Transparency (meters)	Chl <i>a</i> (ug/l)	Dissolved Color (cpu)	Total Phosphorus (ug/l)	Alkalinity gray end pt. @ pH 5.1 (mg/l)	Alkalinity pink end pt. @ pH 4.6 (mg/l)
4 LoonIsl	8/5/05	7.7	1.2	13.9	-----	2.3	2.8
4 LoonIsl	8/12/05	8.4	1.1	12.2	-----	2.1	2.6
4 LoonIsl	8/19/05	8.3	1.1	13.0	-----	2.0	2.6
4 LoonIsl	8/26/05	7.4	1.4	15.6	-----	2.1	2.8
4 LoonIsl	9/6/05	8.4	1.4	13.9	-----	2.1	2.9
4 LoonIsl	9/14/05	8.4	1.6	12.2	-----	2.2	3.0
4 LoonIsl	9/23/05	8.6	1.8	14.8	-----	2.3	2.8
5 Beachwd	7/1/05	5.0	1.9	20.9	-----	2.3	3.1
5 Beachwd	7/7/05	5.2	2.4	21.7	-----	2.3	2.8
5 Beachwd	7/14/05	5.4	1.4	20.9	-----	2.4	2.8
5 Beachwd	7/22/05	6.5	1.6	23.5	-----	2.4	2.9
5 Beachwd	7/27/05	7.6	1.6	19.1	-----	2.3	2.7
5 Beachwd	8/5/05	7.9	1.6	12.2	-----	2.5	3.1
5 Beachwd	8/13/05	8.6	1.4	17.4	-----	2.7	3.2
5 Beachwd	8/20/05	8.2	1.6	13.0	-----	2.5	3.4
5 Beachwd	8/26/05	8.8	1.3	13.0	-----	3.4	4.0
5 Beachwd	9/6/05	9.0	1.4	13.0	-----	2.9	3.5
5 Beachwd	9/13/05	6.9	1.7	10.4	-----	2.3	3.1
5 Beachwd	9/19/05	7.6	2.9	12.2	-----	2.3	3.4
6 Fowler	6/4/05	-----	0.8	30.4	-----	-----	-----
6 Fowler	6/26/05	-----	4.8	15.6	-----	-----	-----
6 Fowler	7/12/05	-----	1.3	-----	-----	-----	-----
6 Fowler	7/26/05	-----	5.4	16.5	-----	-----	-----
6 Fowler	8/14/05	-----	1.2	12.2	-----	-----	-----
6 Fowler	8/27/05	-----	1.4	12.2	-----	-----	-----
6 Fowler	9/11/05	-----	1.8	11.3	-----	-----	-----
6 Fowler	10/3/05	-----	1.5	11.3	-----	-----	-----
7 Cocrmth	6/30/05	4.8	1.1	24.3	-----	3.6	4.3
7 Cocrmth	7/15/05	6.7	1.1	22.6	-----	4.0	5.1
7 Cocrmth	7/26/05	7.4	0.9	17.4	-----	3.1	3.8
7 Cocrmth	8/4/05	8.1	1.1	15.6	-----	3.5	4.5
7 Cocrmth	10/5/05	8.8	1.7	11.3	-----	3.4	4.4
8 Follansbee	6/3/05	8.2	0.9	15.6	-----	4.5	5.6
8 Follansbee	6/10/05	5.4	0.9	16.5	-----	3.4	4.3
8 Follansbee	6/23/05	3.7	1.3	20.9	-----	2.8	3.9
8 Follansbee	6/29/05	4.6	0.9	21.7	-----	3.1	4.2
8 Follansbee	7/13/05	6.4	1.0	18.2	-----	2.8	3.4
8 Follansbee	7/22/05	6.9	1.0	-----	-----	2.6	3.3
8 Follansbee	7/29/05	7.2	0.8	13.0	-----	3.0	3.6
8 Follansbee	8/6/05	7.6	0.6	14.8	-----	3.0	3.6
8 Follansbee	8/14/05	7.9	0.9	11.3	-----	3.0	3.7
8 Follansbee	8/21/05	8.8	1.0	13.0	-----	3.1	3.5
8 Follansbee	8/29/05	8.0	1.3	13.0	-----	3.1	3.8
8 Follansbee	9/5/05	9.3	1.1	13.0	-----	4.3	5.0
8 Follansbee	9/11/05	9.0	1.1	11.3	-----	3.5	4.5
8 Follansbee	9/30/05	8.2	2.7	10.4	-----	2.8	3.4

Site	Date	Secchi Disk Transparency (meters)	Chl <i>a</i> (ug/l)	Dissolved Color (cpu)	Total Phosphorus (ug/l)	Alkalinity gray end pt. @ pH 5.1 (mg/l)	Alkalinity pink end pt. @ pH 4.6 (mg/l)
Whitmor Brk	4/8/05	-----	-----	-----	5.3	-----	-----
Whitmor Brk	4/15/05	-----	-----	-----	6.9	-----	-----
Whitmor Brk	4/22/05	-----	-----	-----	3.2	-----	-----
Dock @ Pops	4/11/05	-----	-----	-----	12.8	-----	-----
Dock @ Pops	4/18/05	-----	-----	-----	4.8	-----	-----
Hemlock Brook	4/15/05	-----	-----	-----	15.1	-----	-----
Hemlock Brook	4/8/05	-----	-----	-----	6.0	-----	-----

<< end of 2005 data listing; 83 records >>

Lakes Lay Monitoring Program, U.N.H.
[Center for Freshwater Biology Data – August 2, 2005]

Site	Depth (meters)	Chl <i>a</i> (ug/L)	Dissolved Color (CPU)	Carbon Dioxide (mg/L)	Alkalinity gray end point @ pH 5.1 (mg/L)	Alkalinity pink end point @ pH 4.6 (mg/L)	Total Phosphorus (ug/L)
2 Mayhew	0.5	2.4	7.7	0.9	3.0	3.5	-----
2 Mayhew	6.0	5.1	9.4	1.2	3.0	3.5	7.9
2 Mayhew	13.5	-----	-----	4.5	3.5	4.1	8.1
2 Mayhew	0-4.0	2.1	9.4	-----	3.0	3.5	6.5
3 Pasquaney	11.5	-----	-----	1.9	3.0	3.5	6.9
3 Pasquaney	0-6.0	1.4	8.6	-----	2.9	3.4	11.8
4 Loon Island	9.5	-----	-----	1.8	3.0	3.6	6.5
4 Loon Island	0-4.0	3.3	6.0	-----	3.2	3.7	5.3
5 Beachwood	0.5	2.5	7.7	0.5	3.0	3.5	-----
5 Beachwood	7.5	2.1	10.3	1.0	3.0	3.5	4.6
5 Beachwood	15.0	-----	-----	1.9	2.9	3.5	5.7
5 Beachwood	0-5.5	2.2	7.7	-----	3.1	3.7	7.0
6 Fowler	0.5	1.5	9.4	-----	3.1	3.7	4.5
7 Cockermouth	0.5	2.6	6.0	0.6	3.1	3.6	-----
7 Cockermouth	7.5	2.9	11.2	1.3	3.0	3.6	6.6
7 Cockermouth	17.5	-----	-----	2.3	3.1	3.7	6.1
7 Cockermouth	0-5.0	2.9	8.6	-----	3.1	3.6	5.3
8 Follansbee	13.5	-----	-----	1.8	2.9	3.4	6.2
8 Follansbee	0-5.5	1.9	8.6	-----	3.1	3.6	5.6

Site Secchi Disk Tranparency (meters)

2 Mayhew	6.7 meters
3 Pasquaney	7.8 meters
4 Loon Island	7.7 meters
5 Beachwood	8.1 meters
7 Cockermouth	8.0 meters
8 Follansbee	8.8 meters

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (uS/cm)	pH (std units)	Turbidity (NTU)
2 Mayhew	0.15	24.0	8.2	-----	6.8	2.6
2 Mayhew	0.21	24.2	8.1	-----	6.8	2.6
2 Mayhew	0.33	24.2	8.1	-----	6.9	2.6
2 Mayhew	0.42	24.2	8.1	-----	6.9	2.6
2 Mayhew	0.55	24.2	8.1	-----	6.9	2.6
2 Mayhew	0.70	24.2	8.1	-----	6.9	2.6
2 Mayhew	0.89	24.2	8.1	-----	6.9	2.7
2 Mayhew	1.09	24.1	8.1	-----	6.9	2.6
2 Mayhew	1.19	24.1	8.1	-----	6.9	2.6
2 Mayhew	1.33	24.1	8.1	-----	6.9	2.6

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
2 Mayhew	1.48	24.1	8.1	----	6.9	2.6
2 Mayhew	1.59	24.0	8.1	----	6.8	2.5
2 Mayhew	1.76	24.0	8.1	----	6.8	2.6
2 Mayhew	1.95	24.0	8.2	39.5	6.8	2.6
2 Mayhew	2.14	24.0	8.2	----	6.9	2.5
2 Mayhew	2.32	24.0	8.1	----	6.9	2.6
2 Mayhew	2.52	24.0	8.1	----	6.9	2.5
2 Mayhew	2.72	24.0	8.1	----	6.9	2.5
2 Mayhew	2.92	24.0	8.1	----	6.9	2.5
2 Mayhew	3.13	23.9	8.1	----	6.9	2.7
2 Mayhew	3.35	23.9	8.1	----	6.9	2.6
2 Mayhew	3.56	23.9	8.1	----	6.9	2.5
2 Mayhew	3.73	23.9	8.1	----	6.9	2.5
2 Mayhew	3.94	23.9	8.1	39.5	6.9	2.5
2 Mayhew	4.15	23.8	8.1	----	6.9	2.5
2 Mayhew	4.33	23.6	8.2	----	6.9	2.5
2 Mayhew	4.50	23.2	8.3	----	6.9	2.6
2 Mayhew	4.64	22.8	8.4	----	6.9	2.9
2 Mayhew	4.77	22.4	8.6	----	6.9	2.8
2 Mayhew	4.94	21.9	8.8	----	6.9	2.7
2 Mayhew	5.17	20.4	9.2	----	6.9	2.8
2 Mayhew	5.39	19.0	9.6	----	7.0	2.8
2 Mayhew	5.60	17.8	10.1	----	7.0	2.8
2 Mayhew	5.92	16.7	10.5	40.8	6.9	3.0
2 Mayhew	6.11	15.7	10.9	----	7.0	3.0
2 Mayhew	6.27	15.1	11.0	----	6.9	3.1
2 Mayhew	6.46	14.6	11.0	----	6.9	3.1
2 Mayhew	6.63	13.9	11.1	----	6.9	3.0
2 Mayhew	6.79	13.2	11.1	----	6.9	3.1
2 Mayhew	6.96	12.7	11.0	----	6.9	3.3
2 Mayhew	7.17	12.3	10.7	----	6.9	3.3
2 Mayhew	7.37	11.9	10.5	----	6.9	3.4
2 Mayhew	7.56	11.5	10.2	----	6.9	4.0
2 Mayhew	7.77	11.1	9.4	----	6.9	3.8
2 Mayhew	7.96	10.8	9.0	43.7	6.8	3.8
2 Mayhew	8.16	10.7	8.6	----	6.8	3.7
2 Mayhew	8.36	10.6	8.3	----	6.8	3.5
2 Mayhew	8.56	10.5	7.8	----	6.8	3.5
2 Mayhew	8.71	10.4	7.6	----	6.8	3.5
2 Mayhew	8.90	10.2	7.5	----	6.8	3.5
2 Mayhew	9.06	10.1	7.4	----	6.8	3.5
2 Mayhew	9.22	9.9	7.3	----	6.7	3.5
2 Mayhew	9.42	9.8	7.0	----	6.7	3.5
2 Mayhew	9.62	9.7	7.0	----	6.7	3.1
2 Mayhew	9.83	9.7	6.9	----	6.7	3.7
2 Mayhew	10.03	9.6	6.8	44.1	6.7	3.7

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
2 Mayhew	10.22	9.5	6.8	----	6.7	3.7
2 Mayhew	10.49	9.4	6.7	----	6.7	3.7
2 Mayhew	10.68	9.4	6.6	----	6.7	3.6
2 Mayhew	10.85	9.3	6.5	----	6.7	3.5
2 Mayhew	11.00	9.2	6.5	----	6.7	3.8
2 Mayhew	11.16	9.2	6.5	----	6.7	3.6
2 Mayhew	11.31	9.2	6.5	----	6.6	3.6
2 Mayhew	11.48	9.1	6.4	----	6.6	3.6
2 Mayhew	11.67	9.1	6.3	----	6.6	3.6
2 Mayhew	11.82	9.1	6.3	----	6.6	3.7
2 Mayhew	12.07	9.1	6.3	44.0	6.6	3.7
2 Mayhew	12.42	9.0	6.2	----	6.6	3.8
2 Mayhew	12.68	9.0	6.1	----	6.6	3.8
2 Mayhew	12.91	9.0	6.0	----	6.6	3.9
2 Mayhew	13.18	9.0	6.0	----	6.6	4.0
2 Mayhew	13.44	9.0	5.8	----	6.6	4.0
2 Mayhew	13.69	9.0	5.8	----	6.5	4.3
2 Mayhew	13.97	8.9	5.7	44.3	6.5	4.5
3 Pasquaney	0.14	24.0	8.3	----	6.7	2.1
3 Pasquaney	0.17	24.0	8.3	----	6.7	2.0
3 Pasquaney	0.31	24.0	8.3	----	6.7	2.0
3 Pasquaney	0.48	24.0	8.4	----	6.7	2.0
3 Pasquaney	0.60	24.0	8.3	----	6.7	2.0
3 Pasquaney	0.72	23.9	8.3	----	6.7	2.0
3 Pasquaney	0.83	23.9	8.3	----	6.7	2.1
3 Pasquaney	0.92	23.9	8.3	----	6.7	2.0
3 Pasquaney	1.01	23.9	8.3	39.4	6.7	2.0
3 Pasquaney	1.11	23.9	8.4	----	6.7	2.0
3 Pasquaney	1.22	23.9	8.3	----	6.7	2.0
3 Pasquaney	1.29	23.9	8.4	----	6.7	2.0
3 Pasquaney	1.37	23.9	8.3	----	6.8	2.0
3 Pasquaney	1.45	23.9	8.3	----	6.8	2.1
3 Pasquaney	1.55	23.9	8.3	----	6.7	2.2
3 Pasquaney	1.66	23.9	8.3	----	6.8	2.1
3 Pasquaney	1.80	23.9	8.3	----	6.8	2.1
3 Pasquaney	1.94	23.9	8.3	39.3	6.8	2.1
3 Pasquaney	2.07	23.9	8.3	----	6.8	2.0
3 Pasquaney	2.15	23.9	8.3	----	6.8	2.0
3 Pasquaney	2.25	23.9	8.3	----	6.8	2.0
3 Pasquaney	2.38	23.9	8.3	----	6.8	2.1
3 Pasquaney	2.49	23.9	8.3	----	6.7	2.1
3 Pasquaney	2.63	23.9	8.3	----	6.8	2.0
3 Pasquaney	2.75	23.9	8.3	----	6.8	2.0
3 Pasquaney	2.84	23.9	8.3	----	6.7	2.0
3 Pasquaney	2.96	23.9	8.3	39.4	6.8	2.0
3 Pasquaney	3.08	23.9	8.3	----	6.8	2.0

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
3 Pasquaney	3.22	23.8	8.3	-----	6.8	2.0
3 Pasquaney	3.35	23.8	8.3	-----	6.8	2.0
3 Pasquaney	3.49	23.8	8.3	-----	6.8	2.0
3 Pasquaney	3.61	23.8	8.3	-----	6.8	2.0
3 Pasquaney	3.73	23.7	8.3	-----	6.8	2.0
3 Pasquaney	3.89	23.7	8.3	-----	6.8	2.4
3 Pasquaney	4.03	23.7	8.4	39.4	6.8	2.3
3 Pasquaney	4.18	23.7	8.3	-----	6.8	2.4
3 Pasquaney	4.31	23.7	8.3	-----	6.8	2.4
3 Pasquaney	4.46	23.7	8.3	-----	6.7	2.4
3 Pasquaney	4.62	23.7	8.3	-----	6.7	2.4
3 Pasquaney	4.75	23.6	8.3	-----	6.8	2.4
3 Pasquaney	4.88	23.6	8.3	-----	6.8	2.3
3 Pasquaney	5.01	23.6	8.3	39.5	6.7	2.3
3 Pasquaney	5.11	23.6	8.3	-----	6.8	2.4
3 Pasquaney	5.23	23.6	8.3	-----	6.8	2.4
3 Pasquaney	5.35	23.6	8.2	-----	6.8	2.4
3 Pasquaney	5.46	23.5	8.2	-----	6.8	2.4
3 Pasquaney	5.60	23.5	8.1	-----	6.7	2.5
3 Pasquaney	5.78	23.5	8.0	-----	6.7	2.5
3 Pasquaney	5.95	23.4	8.0	41.0	6.7	2.4
3 Pasquaney	6.11	23.2	8.1	-----	6.8	2.4
3 Pasquaney	6.25	22.9	8.1	-----	6.8	2.6
3 Pasquaney	6.38	22.6	8.4	-----	6.8	2.5
3 Pasquaney	6.50	22.4	8.5	-----	6.8	2.4
3 Pasquaney	6.64	22.1	8.6	-----	6.8	2.4
3 Pasquaney	6.82	21.6	8.7	-----	6.8	2.4
3 Pasquaney	6.99	21.0	9.0	39.3	6.8	2.5
3 Pasquaney	7.13	20.4	9.2	-----	6.8	2.4
3 Pasquaney	7.26	20.0	9.5	-----	6.8	2.5
3 Pasquaney	7.44	19.7	9.6	-----	6.8	2.5
3 Pasquaney	7.60	19.4	9.8	-----	6.8	2.4
3 Pasquaney	7.74	18.8	9.8	-----	6.8	2.4
3 Pasquaney	7.88	18.1	10.0	-----	6.8	2.4
3 Pasquaney	8.03	17.5	10.3	39.7	6.8	2.5
3 Pasquaney	8.22	17.0	10.4	-----	6.8	2.6
3 Pasquaney	8.44	16.4	10.6	-----	6.8	2.7
3 Pasquaney	8.64	14.8	11.0	-----	6.8	2.6
3 Pasquaney	8.84	13.5	11.4	-----	6.8	3.1
3 Pasquaney	9.04	12.8	11.3	41.3	6.8	3.2
3 Pasquaney	9.22	12.2	11.5	-----	6.8	3.1
3 Pasquaney	9.35	11.7	11.3	-----	6.8	3.1
3 Pasquaney	9.50	11.2	11.4	-----	6.8	3.1
3 Pasquaney	9.68	10.7	11.3	-----	6.8	3.2
3 Pasquaney	9.85	10.3	11.4	-----	6.8	3.2
3 Pasquaney	10.02	9.9	11.5	42.0	6.8	3.0

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
3 Pasquaney	10.20	9.5	11.6	----	6.8	3.1
3 Pasquaney	10.42	9.2	11.5	----	6.7	3.1
3 Pasquaney	10.61	8.9	11.8	----	6.7	3.0
3 Pasquaney	10.77	8.7	11.7	----	6.7	3.0
3 Pasquaney	10.97	8.5	11.7	42.4	6.7	2.9
3 Pasquaney	11.13	8.3	11.7	----	6.7	3.0
3 Pasquaney	11.28	8.2	11.7	----	6.7	3.0
3 Pasquaney	11.45	8.1	11.7	----	6.7	2.9
3 Pasquaney	11.64	7.9	11.7	----	6.7	2.9
3 Pasquaney	11.83	7.8	11.6	----	6.7	2.9
3 Pasquaney	12.02	7.7	11.6	42.5	6.7	3.1
3 Pasquaney	12.28	7.5	11.6	----	6.7	3.1
4 Loon Island	0.02	24.1	8.2	----	6.7	2.4
4 Loon Island	0.06	24.1	8.2	----	6.7	2.4
4 Loon Island	0.22	24.1	8.2	----	6.7	2.9
4 Loon Island	0.38	24.1	8.2	----	6.7	2.9
4 Loon Island	0.58	24.1	8.2	----	6.7	2.9
4 Loon Island	0.73	24.1	8.2	----	6.7	2.9
4 Loon Island	0.85	24.1	8.3	----	6.7	2.8
4 Loon Island	0.98	24.1	8.2	38.9	6.7	2.7
4 Loon Island	1.10	24.0	8.2	----	6.7	2.8
4 Loon Island	1.21	24.0	8.2	----	6.7	2.9
4 Loon Island	1.34	24.0	8.3	----	6.7	2.9
4 Loon Island	1.48	24.0	8.3	----	6.7	2.8
4 Loon Island	1.62	23.9	8.3	----	6.7	2.7
4 Loon Island	1.76	23.9	8.3	----	6.7	2.8
4 Loon Island	1.89	23.8	8.3	----	6.8	2.8
4 Loon Island	2.07	23.8	8.3	38.9	6.8	2.7
4 Loon Island	2.25	23.8	8.3	----	6.8	2.7
4 Loon Island	2.38	23.8	8.3	----	6.8	2.6
4 Loon Island	2.52	23.7	8.3	----	6.8	2.7
4 Loon Island	2.65	23.7	8.3	----	6.8	2.7
4 Loon Island	2.78	23.7	8.3	----	6.8	2.7
4 Loon Island	2.92	23.7	8.3	----	6.8	2.6
4 Loon Island	3.04	23.6	8.3	39.0	6.8	2.6
4 Loon Island	3.18	23.6	8.3	----	6.8	2.5
4 Loon Island	3.31	23.6	8.3	----	6.7	2.6
4 Loon Island	3.45	23.5	8.3	----	6.8	2.6
4 Loon Island	3.59	23.5	8.2	----	6.8	2.6
4 Loon Island	3.74	23.5	8.2	----	6.8	2.6
4 Loon Island	3.88	23.5	8.2	----	6.7	2.5
4 Loon Island	4.05	23.3	8.3	39.0	6.7	2.6
4 Loon Island	4.23	23.1	8.3	----	6.7	2.6
4 Loon Island	4.41	22.8	8.4	----	6.8	2.6
4 Loon Island	4.59	22.3	8.5	----	6.8	2.6
4 Loon Island	4.80	20.8	8.8	----	6.8	2.6

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (uS/cm)	pH (std units)	Turbidity (NTU)
4 Loon Island	5.01	19.5	9.1	39.4	6.8	3.9
4 Loon Island	5.21	19.0	9.4	-----	6.8	3.3
4 Loon Island	5.37	18.3	10.0	-----	6.8	3.0
4 Loon Island	5.51	17.6	10.3	-----	6.8	3.0
4 Loon Island	5.64	17.0	10.4	-----	6.8	2.9
4 Loon Island	5.80	16.4	10.6	-----	6.8	2.8
4 Loon Island	5.95	16.0	10.7	39.9	6.8	2.8
4 Loon Island	6.09	15.7	10.7	-----	6.8	2.7
4 Loon Island	6.26	15.4	10.8	-----	6.7	2.7
4 Loon Island	6.44	15.0	10.9	-----	6.7	2.7
4 Loon Island	6.59	14.6	11.1	-----	6.8	2.6
4 Loon Island	6.76	14.3	11.1	-----	6.8	2.7
4 Loon Island	6.97	14.1	11.1	40.2	6.7	2.7
4 Loon Island	7.18	13.9	11.2	-----	6.7	2.7
4 Loon Island	7.36	13.7	11.2	-----	6.7	2.7
4 Loon Island	7.51	13.5	11.2	-----	6.7	2.7
4 Loon Island	7.64	13.3	11.3	-----	6.7	2.7
4 Loon Island	7.79	13.1	11.2	-----	6.7	2.7
4 Loon Island	7.93	12.8	11.1	-----	6.7	3.0
4 Loon Island	8.05	12.5	11.1	40.7	6.7	3.1
4 Loon Island	8.20	12.1	11.1	-----	6.7	3.2
4 Loon Island	8.36	11.6	11.2	-----	6.7	3.2
4 Loon Island	8.53	11.3	11.2	-----	6.7	3.1
4 Loon Island	8.73	11.2	11.2	-----	6.6	3.3
4 Loon Island	8.95	11.0	11.2	41.3	6.6	3.3
4 Loon Island	9.14	10.9	11.2	-----	6.6	3.3
4 Loon Island	9.31	10.6	11.2	-----	6.6	3.4
4 Loon Island	9.49	10.3	11.3	-----	6.6	3.4
4 Loon Island	9.64	10.0	11.3	-----	6.6	3.4
4 Loon Island	9.80	9.7	11.3	-----	6.6	3.6
4 Loon Island	9.94	9.4	11.3	41.8	6.6	3.9
4 Loon Island	10.08	9.1	11.4	-----	6.6	4.6
4 Loon Island	10.22	8.9	11.5	-----	6.6	4.5
5 Beachwood	0.12	24.7	8.3	-----	6.8	1.2
5 Beachwood	0.23	24.6	8.2	-----	6.7	1.4
5 Beachwood	0.31	24.6	8.2	-----	6.7	1.4
5 Beachwood	0.46	24.5	8.3	-----	6.7	1.3
5 Beachwood	0.58	24.3	8.3	-----	6.8	1.3
5 Beachwood	0.71	24.1	8.3	-----	6.8	1.4
5 Beachwood	0.87	24.1	8.3	-----	6.8	1.4
5 Beachwood	1.01	24.0	8.3	38.1	6.8	1.3
5 Beachwood	1.15	24.0	8.3	-----	6.8	1.3
5 Beachwood	1.27	23.9	8.3	-----	6.8	1.3
5 Beachwood	1.37	23.9	8.3	-----	6.8	1.3
5 Beachwood	1.50	23.9	8.3	-----	6.8	1.3
5 Beachwood	1.66	23.9	8.3	-----	6.7	1.3

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
5 Beachwood	1.81	23.8	8.3	-----	6.7	1.3
5 Beachwood	1.95	23.8	8.3	38.1	6.7	1.3
5 Beachwood	2.12	23.8	8.3	-----	6.7	1.8
5 Beachwood	2.21	23.8	8.3	-----	6.7	1.6
5 Beachwood	2.29	23.8	8.3	-----	6.7	1.6
5 Beachwood	2.35	23.8	8.3	-----	6.7	1.5
5 Beachwood	2.47	23.8	8.3	-----	6.7	1.5
5 Beachwood	2.57	23.8	8.3	-----	6.7	1.4
5 Beachwood	2.73	23.8	8.3	-----	6.7	1.4
5 Beachwood	2.87	23.7	8.3	-----	6.7	1.4
5 Beachwood	3.04	23.7	8.3	38.2	6.7	1.5
5 Beachwood	3.19	23.7	8.3	-----	6.7	1.9
5 Beachwood	3.32	23.7	8.3	-----	6.7	1.8
5 Beachwood	3.47	23.7	8.3	-----	6.7	1.7
5 Beachwood	3.60	23.6	8.3	-----	6.7	1.6
5 Beachwood	3.74	23.6	8.3	-----	6.7	1.6
5 Beachwood	3.88	23.6	8.3	-----	6.7	1.5
5 Beachwood	4.03	23.6	8.3	38.2	6.7	1.6
5 Beachwood	4.17	23.6	8.3	-----	6.7	1.7
5 Beachwood	4.31	23.5	8.3	-----	6.7	1.6
5 Beachwood	4.45	23.4	8.3	-----	6.7	1.7
5 Beachwood	4.59	23.4	8.3	-----	6.7	1.7
5 Beachwood	4.77	23.4	8.3	-----	6.7	1.6
5 Beachwood	4.96	23.4	8.3	38.2	6.7	1.7
5 Beachwood	5.12	23.3	8.3	-----	6.7	1.7
5 Beachwood	5.30	23.3	8.3	-----	6.7	1.8
5 Beachwood	5.52	23.3	8.3	-----	6.7	1.7
5 Beachwood	5.72	23.2	8.3	-----	6.7	1.7
5 Beachwood	5.89	22.9	8.4	-----	6.8	1.7
5 Beachwood	6.06	19.9	9.0	38.2	6.9	1.7
5 Beachwood	6.24	18.1	9.8	-----	7.0	1.8
5 Beachwood	6.40	16.5	10.5	-----	7.0	1.8
5 Beachwood	6.52	15.2	10.8	-----	7.0	1.9
5 Beachwood	6.66	14.8	11.1	-----	7.0	1.8
5 Beachwood	6.82	14.4	11.3	-----	7.0	1.9
5 Beachwood	6.99	13.9	11.5	39.3	6.9	2.3
5 Beachwood	7.14	13.5	11.5	-----	6.9	2.2
5 Beachwood	7.33	13.1	11.7	-----	6.9	2.0
5 Beachwood	7.56	12.7	11.7	-----	6.9	2.0
5 Beachwood	7.79	12.4	11.6	-----	6.9	2.0
5 Beachwood	8.00	12.0	11.7	40.0	6.8	2.0
5 Beachwood	8.20	11.7	11.7	-----	6.8	2.1
5 Beachwood	8.35	11.5	11.7	-----	6.8	2.2
5 Beachwood	8.47	11.3	11.7	-----	6.8	2.2
5 Beachwood	8.59	11.2	11.7	-----	6.8	2.1
5 Beachwood	8.77	11.2	11.7	-----	6.8	2.2

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
5 Beachwood	8.92	11.0	11.6	40.3	6.8	2.4
5 Beachwood	9.16	10.9	11.6	----	6.7	2.4
5 Beachwood	9.37	10.7	11.7	----	6.7	2.6
5 Beachwood	9.59	10.3	11.8	----	6.7	2.5
5 Beachwood	9.84	9.8	11.7	----	6.7	2.4
5 Beachwood	10.08	9.5	11.8	40.8	6.7	2.5
5 Beachwood	10.31	9.3	11.8	----	6.7	2.5
5 Beachwood	10.55	9.1	11.8	----	6.7	2.5
5 Beachwood	10.81	9.0	11.8	----	6.7	2.6
5 Beachwood	11.06	8.8	11.8	41.0	6.7	2.8
5 Beachwood	11.29	8.7	11.8	----	6.7	2.8
5 Beachwood	11.52	8.6	11.9	----	6.6	2.8
5 Beachwood	11.71	8.6	11.8	----	6.6	2.8
5 Beachwood	11.91	8.6	11.8	41.2	6.6	2.8
5 Beachwood	12.11	8.5	11.8	----	6.6	2.8
5 Beachwood	12.28	8.4	11.8	----	6.6	2.8
5 Beachwood	12.44	8.1	11.8	----	6.6	2.6
5 Beachwood	12.60	8.0	11.8	----	6.6	2.7
5 Beachwood	12.78	7.9	11.8	----	6.6	2.7
5 Beachwood	12.99	7.8	11.8	41.3	6.6	3.4
5 Beachwood	13.21	7.7	11.8	----	6.6	3.2
5 Beachwood	13.40	7.7	11.8	----	6.6	3.2
5 Beachwood	13.57	7.6	11.8	----	6.6	3.1
5 Beachwood	13.73	7.6	11.8	----	6.6	10.7
5 Beachwood	13.86	7.6	11.8	----	6.6	7.3
5 Beachwood	14.00	7.5	11.7	41.5	6.6	5.3
5 Beachwood	14.15	7.5	11.7	----	6.6	4.4
5 Beachwood	14.33	7.4	11.7	----	6.6	3.9
5 Beachwood	14.50	7.4	11.7	----	6.6	3.6
5 Beachwood	14.63	7.4	11.7	----	6.5	4.2
5 Beachwood	14.76	7.3	11.7	----	6.5	3.7
5 Beachwood	14.85	7.2	11.8	----	6.5	3.7
5 Beachwood	14.98	7.1	11.7	----	6.5	3.5
5 Beachwood	15.14	7.0	11.7	----	6.5	3.3
5 Beachwood	15.37	7.0	11.7	----	6.5	3.3
5 Beachwood	15.59	6.9	11.7	----	6.5	3.3
7 Cockermonth	0.20	24.1	8.3	----	6.7	2.2
7 Cockermonth	0.22	24.1	8.3	----	6.7	2.1
7 Cockermonth	0.36	24.0	8.3	----	6.7	2.1
7 Cockermonth	0.56	24.0	8.3	----	6.7	2.1
7 Cockermonth	0.73	24.0	8.3	----	6.7	2.0
7 Cockermonth	0.88	23.9	8.3	38.9	6.7	1.9
7 Cockermonth	1.12	23.9	8.3	----	6.7	1.9
7 Cockermonth	1.53	23.8	8.3	----	6.7	2.0
7 Cockermonth	1.79	23.7	8.3	----	6.7	2.0
7 Cockermonth	1.97	23.6	8.4	39.0	6.8	2.0

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
7 Cockermonth	2.15	23.5	8.4	----	6.8	2.0
7 Cockermonth	2.29	23.5	8.3	----	6.8	2.0
7 Cockermonth	2.42	23.4	8.4	----	6.7	2.0
7 Cockermonth	2.58	23.4	8.3	----	6.7	2.0
7 Cockermonth	2.84	23.4	8.3	----	6.7	2.0
7 Cockermonth	3.06	23.4	8.3	39.2	6.7	2.1
7 Cockermonth	3.22	23.3	8.3	----	6.8	2.0
7 Cockermonth	3.27	23.3	8.3	----	6.7	2.1
7 Cockermonth	3.28	23.3	8.3	----	6.7	2.1
7 Cockermonth	3.50	23.3	8.4	----	6.7	2.1
7 Cockermonth	3.92	23.3	8.3	39.3	6.7	2.1
7 Cockermonth	4.13	23.3	8.3	----	6.7	2.0
7 Cockermonth	4.26	23.2	8.3	----	6.7	2.0
7 Cockermonth	4.41	23.2	8.3	----	6.7	2.0
7 Cockermonth	4.61	23.2	8.3	----	6.7	2.0
7 Cockermonth	4.82	23.2	8.3	----	6.7	2.0
7 Cockermonth	5.05	22.9	8.4	39.1	6.7	2.1
7 Cockermonth	5.21	21.1	8.6	----	6.8	2.2
7 Cockermonth	5.30	20.1	9.2	----	6.8	2.3
7 Cockermonth	5.37	19.9	9.5	----	6.8	2.3
7 Cockermonth	5.54	19.5	9.8	----	6.8	2.3
7 Cockermonth	5.76	17.3	10.5	----	6.8	2.3
7 Cockermonth	5.92	16.2	10.7	40.0	6.8	2.3
7 Cockermonth	6.10	15.8	10.8	----	6.8	2.3
7 Cockermonth	6.25	15.4	11.0	----	6.8	2.3
7 Cockermonth	6.45	14.9	11.1	----	6.8	2.3
7 Cockermonth	6.67	14.4	11.2	----	6.8	2.3
7 Cockermonth	6.85	14.0	11.3	----	6.8	2.4
7 Cockermonth	7.08	13.6	11.4	40.0	6.8	2.4
7 Cockermonth	7.34	13.2	11.5	----	6.8	2.6
7 Cockermonth	7.57	12.6	11.4	----	6.8	2.5
7 Cockermonth	7.79	12.3	11.6	----	6.8	2.5
7 Cockermonth	8.00	11.9	11.7	40.5	6.8	2.5
7 Cockermonth	8.16	11.6	11.7	----	6.8	2.5
7 Cockermonth	8.35	11.3	11.6	----	6.7	2.6
7 Cockermonth	8.54	11.0	11.7	----	6.7	2.6
7 Cockermonth	8.71	10.6	11.7	----	6.7	2.7
7 Cockermonth	8.90	10.2	11.7	41.2	6.7	2.7
7 Cockermonth	9.13	9.9	11.7	----	6.7	2.7
7 Cockermonth	9.31	9.5	11.8	----	6.7	2.8
7 Cockermonth	9.48	9.2	11.8	----	6.7	2.7
7 Cockermonth	9.63	9.0	11.8	----	6.7	2.8
7 Cockermonth	9.82	8.9	11.8	----	6.7	2.9
7 Cockermonth	10.04	8.7	11.9	41.8	6.7	2.9
7 Cockermonth	10.21	8.6	11.9	----	6.7	2.8
7 Cockermonth	10.38	8.5	11.9	----	6.7	2.8

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
7 Cockermonth	10.53	8.4	11.9	----	6.7	2.8
7 Cockermonth	10.68	8.3	11.9	----	6.7	3.2
7 Cockermonth	10.79	8.3	11.8	----	6.7	3.0
7 Cockermonth	10.90	8.2	11.8	41.9	6.6	3.0
7 Cockermonth	11.07	8.2	11.8	----	6.6	3.1
7 Cockermonth	11.21	8.1	11.8	----	6.6	3.3
7 Cockermonth	11.32	8.1	11.8	----	6.6	3.3
7 Cockermonth	11.45	8.0	11.8	----	6.6	3.2
7 Cockermonth	11.56	8.0	11.7	----	6.6	3.2
7 Cockermonth	11.71	8.0	11.7	----	6.6	3.3
7 Cockermonth	11.89	7.9	11.7	----	6.6	3.2
7 Cockermonth	12.03	7.9	11.7	42.0	6.6	3.2
7 Cockermonth	12.17	7.8	11.7	----	6.6	3.3
7 Cockermonth	12.34	7.8	11.8	----	6.6	3.4
7 Cockermonth	12.48	7.7	11.8	----	6.6	3.4
7 Cockermonth	12.62	7.7	11.8	----	6.6	3.4
7 Cockermonth	12.76	7.6	11.8	----	6.6	3.7
7 Cockermonth	12.93	7.6	11.8	42.1	6.6	3.7
7 Cockermonth	13.11	7.5	11.8	----	6.6	3.6
7 Cockermonth	13.29	7.5	11.8	----	6.6	3.7
7 Cockermonth	13.43	7.5	11.9	----	6.6	4.5
7 Cockermonth	13.62	7.4	11.9	----	6.6	4.2
7 Cockermonth	13.79	7.4	11.8	----	6.6	3.9
7 Cockermonth	13.97	7.4	11.9	42.1	6.6	3.8
7 Cockermonth	14.15	7.3	11.8	----	6.6	3.8
7 Cockermonth	14.45	7.3	11.8	----	6.6	4.2
7 Cockermonth	14.66	7.3	11.9	----	6.5	4.0
7 Cockermonth	14.85	7.2	11.9	----	6.5	4.0
7 Cockermonth	15.04	7.2	11.8	----	6.5	4.7
7 Cockermonth	15.26	7.2	11.8	----	6.5	4.5
7 Cockermonth	15.45	7.2	11.8	----	6.5	4.3
7 Cockermonth	15.64	7.1	11.8	----	6.5	4.2
7 Cockermonth	15.81	7.1	11.7	----	6.5	4.2
7 Cockermonth	15.94	7.1	11.7	----	6.5	4.1
7 Cockermonth	16.07	7.1	11.7	----	6.5	4.1
7 Cockermonth	16.19	7.1	11.7	----	6.5	4.1
7 Cockermonth	16.35	7.1	11.7	----	6.5	4.1
7 Cockermonth	16.51	7.0	11.7	----	6.5	4.1
7 Cockermonth	16.68	7.0	11.7	----	6.5	4.1
7 Cockermonth	16.82	7.0	11.6	----	6.5	4.1
7 Cockermonth	16.98	7.0	11.5	----	6.5	4.8
7 Cockermonth	17.13	7.0	11.5	----	6.5	5.1
7 Cockermonth	17.26	6.9	11.4	----	6.5	5.1
7 Cockermonth	17.40	6.9	11.4	----	6.5	5.2
7 Cockermonth	17.53	6.9	11.4	----	6.5	5.2
7 Cockermonth	17.66	6.9	11.4	----	6.5	5.1

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
7 Cockermonth	17.80	6.9	11.4	----	6.5	5.2
7 Cockermonth	17.97	6.9	11.4	----	6.5	5.3
8 Follansbee	0.09	24.4	8.3	----	6.8	1.6
8 Follansbee	0.10	24.4	8.3	38.6	6.8	1.7
8 Follansbee	0.20	24.4	8.3	----	6.8	1.7
8 Follansbee	0.35	24.4	8.3	----	6.8	1.7
8 Follansbee	0.47	24.4	8.3	----	6.8	1.7
8 Follansbee	0.56	24.4	8.3	----	6.8	1.7
8 Follansbee	0.63	24.4	8.3	----	6.8	1.7
8 Follansbee	0.69	24.4	8.3	----	6.8	1.8
8 Follansbee	0.77	24.4	8.3	----	6.8	1.7
8 Follansbee	0.87	24.4	8.3	----	6.8	1.7
8 Follansbee	0.97	24.4	8.3	38.7	6.8	1.7
8 Follansbee	1.11	24.3	8.3	----	6.8	1.8
8 Follansbee	1.28	24.3	8.3	----	6.8	1.7
8 Follansbee	1.44	24.3	8.3	----	6.7	1.7
8 Follansbee	1.58	24.3	8.3	----	6.7	1.7
8 Follansbee	1.77	24.3	8.3	----	6.7	1.6
8 Follansbee	1.90	24.2	8.3	----	6.7	1.6
8 Follansbee	2.04	24.2	8.4	38.7	6.7	1.6
8 Follansbee	2.19	24.1	8.4	----	6.7	1.6
8 Follansbee	2.36	24.1	8.4	----	6.7	1.7
8 Follansbee	2.52	24.0	8.4	----	6.7	1.7
8 Follansbee	2.71	24.0	8.4	----	6.7	1.7
8 Follansbee	2.93	23.9	8.4	38.6	6.8	1.7
8 Follansbee	3.15	23.8	8.4	----	6.8	1.7
8 Follansbee	3.35	23.7	8.4	----	6.8	1.7
8 Follansbee	3.51	23.7	8.4	----	6.8	1.7
8 Follansbee	3.67	23.6	8.4	----	6.8	1.7
8 Follansbee	3.84	23.6	8.4	----	6.8	1.7
8 Follansbee	4.01	23.5	8.4	38.7	6.8	1.7
8 Follansbee	4.16	23.5	8.4	----	6.8	1.8
8 Follansbee	4.30	23.4	8.4	----	6.8	1.8
8 Follansbee	4.47	23.4	8.4	----	6.8	1.8
8 Follansbee	4.64	23.4	8.4	----	6.8	1.8
8 Follansbee	4.83	23.4	8.4	----	6.7	1.8
8 Follansbee	5.01	23.4	8.4	38.6	6.7	1.8
8 Follansbee	5.19	23.3	8.4	----	6.7	2.0
8 Follansbee	5.38	23.3	8.4	----	6.7	1.9
8 Follansbee	5.53	23.3	8.4	----	6.7	1.9
8 Follansbee	5.69	23.3	8.4	----	6.7	1.9
8 Follansbee	5.85	23.3	8.4	----	6.8	1.8
8 Follansbee	6.04	23.2	8.4	38.5	6.8	2.0
8 Follansbee	6.22	23.2	8.4	----	6.7	1.9
8 Follansbee	6.44	23.2	8.4	----	6.7	1.9
8 Follansbee	6.62	23.0	8.4	----	6.8	2.0

Site	Depth (meters)	Temperature (°C)	Dissolved Oxygen (mg/L)	Specific Conductivity @ 25°C (µS/cm)	pH (std units)	Turbidity (NTU)
8 Follansbee	6.79	20.8	8.8	----	6.9	2.0
8 Follansbee	6.95	18.1	9.4	----	7.0	2.2
8 Follansbee	7.10	15.9	10.3	----	7.0	2.2
8 Follansbee	7.26	14.1	11.2	----	7.0	2.3
8 Follansbee	7.47	13.1	11.7	----	7.0	2.3
8 Follansbee	7.68	12.6	11.7	----	7.0	2.3
8 Follansbee	7.84	12.1	12.0	----	7.0	2.3
8 Follansbee	8.00	11.8	12.0	40.6	6.9	2.3
8 Follansbee	8.15	11.5	12.1	----	6.9	2.4
8 Follansbee	8.31	11.3	12.1	----	6.9	2.3
8 Follansbee	8.47	11.1	12.1	----	6.9	2.7
8 Follansbee	8.64	10.8	12.1	----	6.9	2.5
8 Follansbee	8.84	10.5	12.1	----	6.9	2.4
8 Follansbee	9.01	10.2	12.1	41.1	6.8	2.6
8 Follansbee	9.17	9.9	12.2	----	6.8	2.7
8 Follansbee	9.35	9.7	12.1	----	6.8	2.6
8 Follansbee	9.52	9.5	12.0	----	6.8	2.5
8 Follansbee	9.65	9.4	12.1	----	6.8	2.5
8 Follansbee	9.81	9.3	12.0	----	6.8	3.4
8 Follansbee	9.96	9.2	12.1	41.3	6.8	3.2
8 Follansbee	10.14	9.1	12.0	----	6.8	2.9
8 Follansbee	10.36	9.0	11.9	----	6.7	2.7
8 Follansbee	10.59	8.9	11.9	----	6.7	2.7
8 Follansbee	10.78	8.8	12.0	----	6.7	2.7
8 Follansbee	10.97	8.6	11.9	41.5	6.7	2.8
8 Follansbee	11.16	8.5	11.9	----	6.7	2.7
8 Follansbee	11.38	8.4	11.9	----	6.7	2.7
8 Follansbee	11.59	8.2	11.9	----	6.7	2.6
8 Follansbee	11.82	8.1	11.9	----	6.7	2.6
8 Follansbee	12.06	8.0	11.9	41.7	6.7	3.1
8 Follansbee	12.31	7.9	11.9	----	6.7	2.9
8 Follansbee	12.52	7.8	11.9	----	6.7	3.0
8 Follansbee	12.74	7.7	11.9	----	6.7	2.9
8 Follansbee	12.93	7.6	11.9	----	6.6	3.0
8 Follansbee	13.11	7.5	11.9	41.9	6.6	3.0
8 Follansbee	13.27	7.5	11.9	----	6.6	3.0
8 Follansbee	13.41	7.4	11.8	----	6.6	3.6
8 Follansbee	13.58	7.4	11.7	----	6.6	3.5
8 Follansbee	13.80	7.3	11.7	----	6.6	3.5
8 Follansbee	13.93	7.3	11.7	----	6.6	3.5
8 Follansbee	14.02	7.2	11.6	----	6.6	3.5
8 Follansbee	14.08	7.2	11.5	----	6.6	3.5

APPENDIX B

DETERMINING WATER QUALITY CHANGES AND TRENDS

Box and Whisker Plots

Quick Overview:

The 2005 summary New Hampshire Lakes Lay Monitoring Program (NH LLMP) reports include *box-and-whisker* plots that have replaced the annual graphs that historically depicted the minimum, average and maximum values. The *box-and-whisker* plot provides a visual representation of how the data are spread out and how much variation there is. Thus, the *box-and-whisker* plots will provide more detail into how your data are distributed.

Basically, these plots show how the data group together for a given year. The line in the “box” represents the sample median, the extent of the “box” represents a statistical range for comparison to another year, the “whiskers” show the boundaries of what could be considered the representative range of all the samples, and any points above or below the whiskers show atypical readings or “outliers” that represent an extreme condition or difference from that year’s data range. An algae bloom event may cause this type of outlier to occur in the chlorophyll data (high point) or Secchi disk clarity (low point).

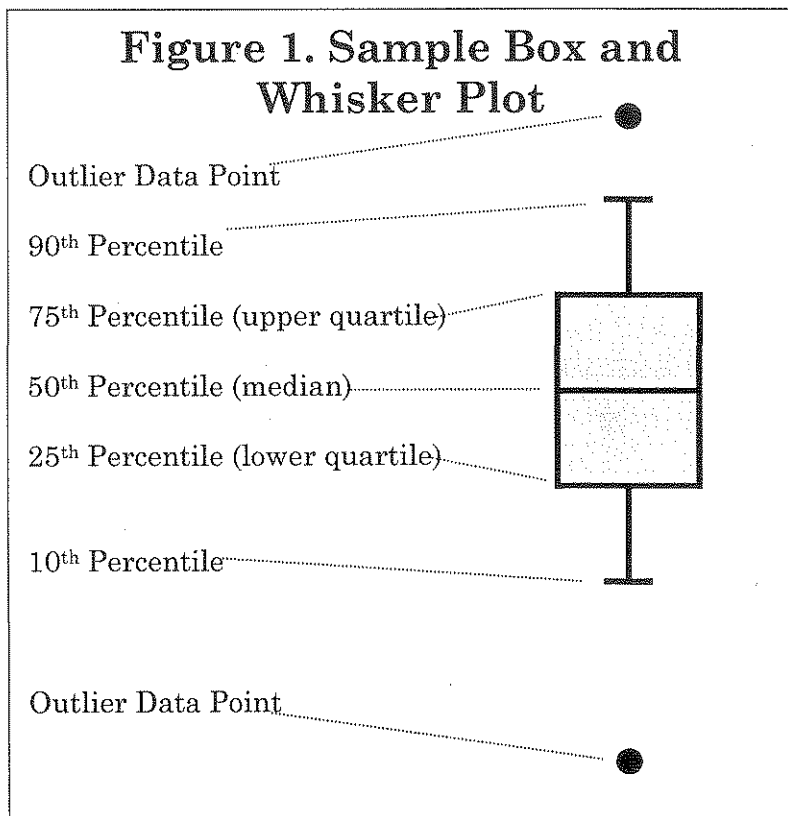
We recommend that each NH LLMP participating group plan on collecting weekly or biweekly measurements throughout the sampling season to ensure that enough data are available for this type of statistical analysis. We suggest that at least 8 data collections per year occur and generally set 10 measurements per year as a sampling effort goal per site.

We can employ the appropriate statistical techniques for detecting the extent that change is occurring when the sampling effort recommendations are followed. Your report summary should include box and whisker plots as well as a basic interpretation for your lake. If you have additional questions on interpreting your results feel free to call the Educational Program Coordinator (Bob Craycraft) at 603-862-3696.

The Details:

In the sections below we further describe the use of the box and whisker plot for those that are interested on how they are determined and how they are interpreted:

The **box-and-whisker plot** is good at showing the **extreme values** and the range of middle values of your data (Figure 1). The box depicts the middle values of a variable, while the **whiskers** stretch to demonstrate the values between which 80% of the data points will fall. The filled circles then reflect the “outlier” data points that fall outside of the whiskers and reflect values that are atypically high or atypically low relative to the other data measured for a given year.



The box-and-whisker plots can be summarized as a graphic that displays the following important features of the data when they are arranged in order from least to greatest:

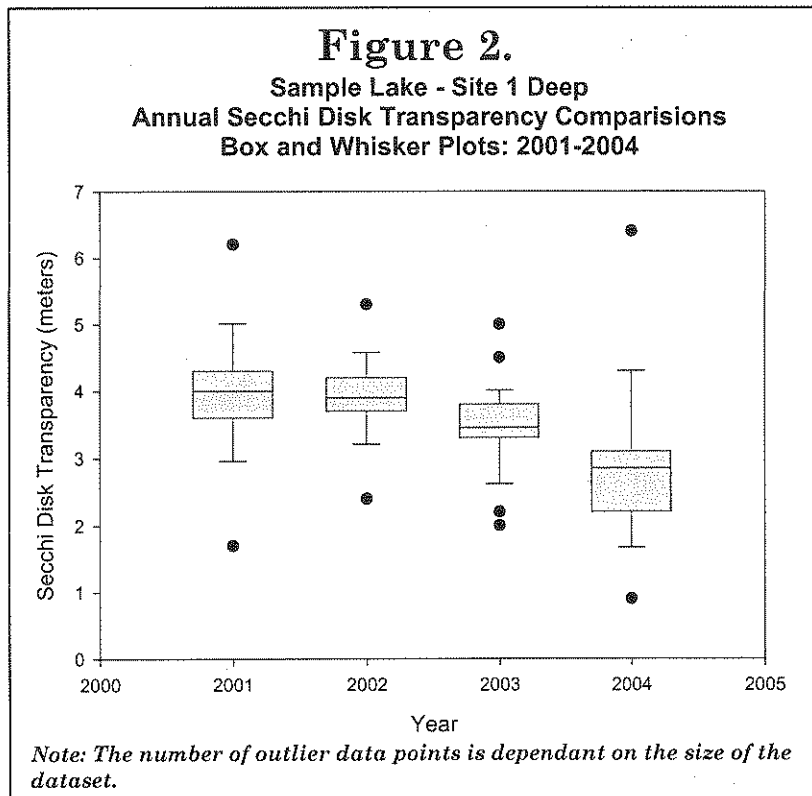
- Median (50th percentile) – the middle of the data.
- Lower Quartile (25th percentile) – the point below which 25% of the data points are located.
- Upper Quartile (75th percentile) – the point below which 75% of the data points are located.
- 90th Percentile – the point below which 90% of the data points are located.
- 10th Percentile – the point below which 10% of the data points are located.
- Outlier Data points – data points that represent the upper 10% or the lowest 10% of the data collected for a specific year.

Note: A minimum number of data points is required to compute each feature documented above. At least three points are required to compute the Lower and the Upper Quartiles, five points are needed to compute the 10th percentile, and six points are needed to compute the 90th percentile. In the event that insufficient data points have been collected, features will not be graphed due to the inability to reliably calculate the respective attribute.

Sample box-and-whisker plot interpretation:

A sample *box-and-whisker* plot is depicted in Figure 2 and it provides an opportunity to assess the usefulness of this type of plot at interpreting water quality monitoring data. The imaginary data depicted in Figure 2 reflect the annual water transparency measurements recorded between the years 2001 and 2004. As you can glean from Figure 2, the distribution of the water clarity measurements has shifted to less clear conditions between 2001 and 2004. The median values, as well as the upper and lower quartiles (what is represented by the gray shaded box) have gradually shifted to less clear conditions over the four year span. The data points that lie between the upper and lower quartiles reflect 50% of the data collected for a given year and can provide insight into whether or not the water quality data are varying significantly between or among years. In extreme cases, when the gray shaded regions do not overlap between successive years or among years, one can quickly determine that the data distribution is significantly different for those years where the middle data (gray shading) does not overlap. Such differences can reflect long-term trends or can be a reflection of extreme climatic conditions for a given year such as atypically wet or atypically dry conditions that can have a profound impact on water quality.

Additional evaluation of the data can include a review of the 10th and the 90th percen-



tiles (the whiskers) that provide additional insight into the distribution of the data. In this case, the trends exhibited by the 10th and the 90th percentiles are following the pattern of decreasing Secchi Disk Transparency as is exhibited by boxes (gray shaded regions). Outlier data points that fall outside of the “whiskers” can also be insightful. Such extreme values can be an early indicator of coming trends or can be an early warning sign of potential water quality problems. For instance, the occurrence of atypically shallow Secchi Disk transparency measurements can be an indication of short-term water quality problems such as excessive sediment loading or an algal bloom. If such problems are not contended with, but are instead left unattended, the longer-term impact could result in an increase in the magnitude

and frequency of the water transparency reductions that, in turn, would result in a decreasing trend as evidenced by a shift of the "Boxes" to shallower water transparencies. There might also be occasions when the Secchi Disk transparency outliers reflect atypically clear water clarity. Such outliers can be a sign that conditions are improving or, as is often the case, the water quality is responding to short-term climatic variations that can have a profound impact on the water quality data. For instance, the outlier data point of 6.4 meters that was documented in 2004 (Figure 2) is counter intuitive to the long-term trend of decreasing water quality. Plausible explanations for such an anomaly could be due to short-term overgrazing of algae by zooplankton (typical for moderate to highly productive lakes), an abrupt shift in climate that might have favored clearer water (cloudy days or cooler water) or perhaps there was some sort of human intervention, such as a fish stocking or lake treatment that would have resulted in clearer water clarities.

Your 2005 non-technical summary in this report includes a basic interpretation of the box-and whisker plots that are specific to your lake. However, since you have personal knowledge of the conditions of your lake and local events that might influence the water quality measurements, you might have additional insight into the cause of the water quality fluctuations that have not been discussed in the report. Should you want to discuss the water quality results further, or provide additional information that you feel is important, please contact Bob Craycraft by phone, (603) 862-3696, or by email, bob.craycraft@unh.edu. Since the *box-and-whisker* plots are being included for only the second time in the 2005 summary reports we would appreciate your feedback regarding your thought on these graphs and whether they are appropriate for our volunteer monitoring audience.

APPENDIX C

GLOSSARY OF LIMNOLOGICAL TERMS

Aerobe- Organisms requiring oxygen for life. All animals, most algae and some bacteria require oxygen for respiration.

Algae- See phytoplankton.

Alkalinity- Total concentration of bicarbonate and hydroxide ions (in most lakes).

Anaerobe- Organisms not requiring oxygen for life. Some algae and many bacteria are able to respire or ferment without using oxygen.

Anoxic- A system lacking oxygen, therefore incapable of supporting the most common kind of biological respiration, or of supporting oxygen-demanding chemical reactions. The deeper waters of a lake may become anoxic if there are many organisms depleting oxygen via respiration, and there is little or no replenishment of oxygen from photosynthesis or from the atmosphere.

Benthic- Referring to the bottom sediments.

Bacterioplankton- Bacteria adapted to the "open water" or "planktonic" zone of lakes, adapted for many specialized habitats and include groups that can use the sun's energy (phytoplankton), some that can use the energy locked in sulfur or iron, and others that gain energy by decomposing dead material.

Bicarbonate- The most important ion (chemical) involved in the buffering system of New Hampshire lakes.

Buffering- The capacity of lakewater to absorb acid with a minimal change in the pH. In New Hampshire the chemical responsible for buffering is the bicarbonate ion. (See pH.)

Chloride- One of the components of salts dissolved in lakewater. Generally the most abundant ion in New Hampshire lakewater, it may be used as an indicator of raw sewage or of road salt.

Chlorophyll a- The main green pigment in plants. The concentration of chlorophyll *a* in lakewater is often used as an indicator of algal abundance.

Circulation- The period during spring and fall when the combination of low water temperature and wind cause the water column to mix freely over its entire depth.

Density- The weight per volume of a substance. The more dense an object, the heavier it feels. Low-density liquids will float on higher-density liquids.

Dimictic- The thermal pattern of lakes where the lake circulates, or mixes, twice a year. Other patterns such as polymictic (many periods of circulation per year) are uncommon in New Hampshire. (See also meromictic and holomictic).

Dystrophy- The lake trophic state in which the lakewater is highly stained with humic acids (reddish brown or yellow stain) and has low productivity. Chlorophyll *a* concentration may be low or high.

Epilimnion- The uppermost layer of water during periods of thermal stratification. (See lake diagram).

Eutrophy- The lake trophic state in which algal production is high. Associated with eutrophy is low Secchi Disk depth, high chlorophyll *a*, and high total phosphorus. From an esthetic viewpoint these lakes are "bad" because water clarity is low, aquatic plants are often found in abundance, and cold-water fish such as trout and salmon are usually not present. A good aspect of eutrophic lakes is their high productivity in terms of warm-water fish such as bass, pickerel, and perch.

Free CO₂- Carbon dioxide that is not combined chemically with lake water or any other substances. It is produced by respiration, and is used by plants and bacteria for photosynthesis.

Holomixis- The condition where the entire lake is free to circulate during periods of overturn. (See meromixis.)

Humic Acids- Dissolved organic compounds released from decomposition of plant leaves and stems. Humic acids are red, brown, or yellow in color and are present in nearly all lakes in New Hampshire. Humic acids are consumed only by fungi, and thus are relatively resistant to biological decomposition.

Hydrogen Ion- The "acid" ion, present in small amounts even in distilled water, but contributed to rain-water by atmospheric processes, to ground-water by soils, and to lakewater by biological organisms and sediments. The active component of "acid rain". See also "pH" the symbolic value inversely and exponentially related to the hydrogen ion.

Hypolimnion- The deepest layer of lakewater during periods of thermal stratification. (See lake diagram)

Lake- Any "inland" body of relatively "standing" water. Includes many synonyms such as ponds, tarns, loches, billabongs, bogs, marshes, etc.

Lake Morphology- The shape and size of a lake and its basin.

Littoral- The area of a lake shallow enough for submerged aquatic plants to grow.

Meromixis- The condition where the entire lake fails to circulate to its deepest points; caused by a high concentration of salt in the deeper waters, and by peculiar landscapes (small deep lakes surrounded by hills and/or forests. (Contrast holomixis.)

Mesotrophy- The lake trophic state intermediate between oligotrophy and eutrophy. Algal production is moderate, and chlorophyll α , Secchi Disk depth, and total phosphorus are also moderate. These lakes are esthetically "fair" but not as good as oligotrophic lakes.

Metalimnion- The "middle" layer of the lake during periods of summer thermal stratification. Usually defined as the region where the water temperature changes at least one degree per meter depth. Also called the thermocline.

Mixis- Periods of lakewater mixing or circulation.

Mixotrophy- The lake condition where the water is highly stained with humic acids, but algal production and chlorophyll α values are also high.

Oligotrophy- The lake trophic state where algal production is low, Secchi Disk depth is deep, and chlorophyll α and total phosphorus are low. Esthetically these lakes are the "best" because they are clear and have a minimum of algae and aquatic plants. Deep oligotrophic lakes can usually support cold-water fish such as lake trout and land-locked salmon.

Overturn- See circulation or mixis

pH- A measure of the hydrogen ion concentration of a liquid. For every decrease of 1 pH unit, the hydrogen ion concentration increases 10 times. Symbolically, the pH value is the "negative logarithm" of the hydrogen ion concentration. For example, a pH of 5 represents a hydrogen ion concentration of 10^{-5} molar. [Please thank the chemists for this lovely symbolism -- and ask them to explain it in lay terms!] In any event, the higher the pH value, the lower the hydrogen ion concentration. The range is 0 to 14, with 7 being neutral 1 denoting high acid condition and 14 denoting very basic condition.

Photosynthesis- The process by which plants convert the inorganic substances carbon dioxide and water into organic glucose (sugar) and oxygen using sunlight as the energy source. Glucose is an energy source for growth, reproduction, and maintenance of almost all life forms.

Phytoplankton- Microscopic algae which are suspended in the "open water" zone of lakes and ponds. A major source of food for zooplankton. Common examples include: diatoms, euglenoids, dinoflagellates, and many others. Usually included are the blue-green bacteria.

Parts per million- Also known as "ppm". This is a method of expressing the amount of one substance (solute) dissolved in another (solvent). For example, a solution with 10 ppm of oxygen has 10 pounds of oxygen for every 999,990 pounds (500 tons) of water. Domestic sewage usually contains from 2 to 10 ppm phosphorus.

Parts per billion- Also known as "ppb". This is only 1/1000 of ppm, therefore much less concentrated. As little as 1 ppb of phosphorus will sustain growth of algae. As little as 10 ppb phosphorus will cause algal blooms! Think of the ratio as 1 milligram (1/28000 of an ounce) of phosphorus in 25 barrels of water (55 gallon drums)! Or, 1 gallon of septic waste diluted into 10,000 gallons of lakewater. It adds up fast!

Plankton- Community of microorganisms that live suspended in the water column, not attached to the bottom sediments or aquatic plants. See also "bacterioplankton" (bacteria), "phytoplankton" (algae) and "zooplankton" (microcrustaceans and rotifers).

Saturated- When a solute (such as water) has dissolved all of a substance that it can. For example, if you add table salt to water, a point is reached where any additional salt fails to dissolve. The water is then said to be saturated with table salt. In lakewater, gaseous oxygen can dissolve, but eventually the water becomes saturated with oxygen if exposed sufficiently long to the atmosphere or another source of oxygen.

Specific Conductivity- A measure of the amount of salt present in lakewater. As the salt concentration increases, so does the specific conductivity (electrical conductivity).

Stratum- A layer or "blanket". Can be used to refer to one of the major layers of lakewater such as the epilimnion, or to any layers of organisms or chemicals that may be present in a lake.

Thermal Stratification- The process by which layers are built up in the lake due to heating by the sun and partial mixing by wind.

Thermocline- Region of temperature change. (See metalimnion.)

Total Phosphorus- A measure of the concentration of phosphorus in lakewater. Includes both free forms (dissolved), and chemically combined form (as in living tissue, or in dead but suspended organisms).

Trophic Status- A classification system placing lakes into similar groups according to their amount of algal production. (See Oligotrophy, Mesotrophy, Eutrophy, Mixotrophy, and Dystrophy for definitions of the major categories)

Z- A symbol used by limnologists as an abbreviation for depth.

Zooplankton- Microscopic animals in the planktonic community. Some are called "water fleas", but most are known by their scientific names. Scientific names include: *Daphnia*, *Cyclops*, *Bosmina*, and *Kellicottia*.